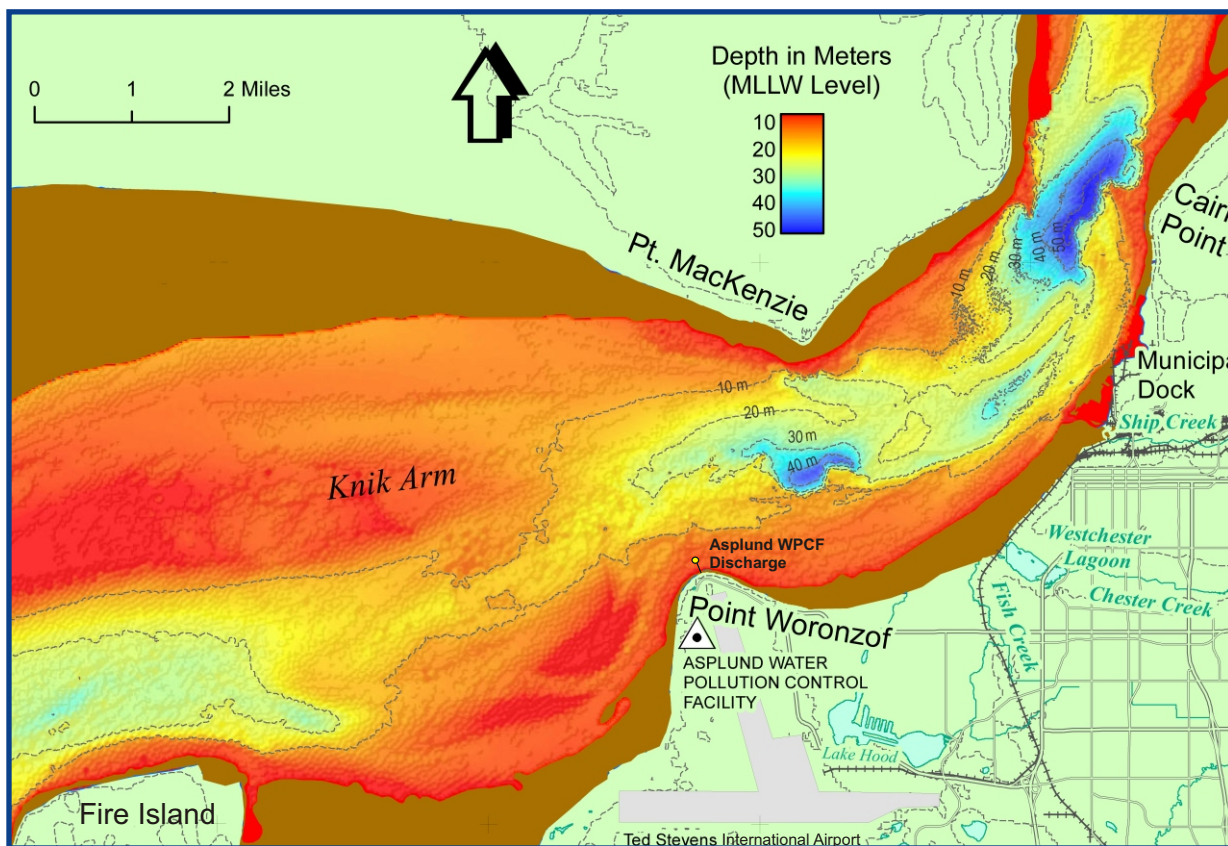


# MONITORING PROGRAM ANNUAL REPORT

## JANUARY - DECEMBER 2010

### ANCHORAGE WATER AND WASTEWATER UTILITY JOHN M. ASPLUND WATER POLLUTION CONTROL FACILITY AT POINT WORONZOF



Prepared for:



MUNICIPALITY OF ANCHORAGE  
Anchorage Water & Wastewater Utility  
Anchorage, Alaska

Prepared by:



**KINNETIC  
LABORATORIES  
INCORPORATED**

1102 WEST 7TH AVE., ANCHORAGE, AK 99501  
(907) 276-6178

February 2011



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**NPDES Permit AK-002255-1**

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**February 2011**



## **PREFACE**

This document is the Monitoring Program Annual Report required for submittal to the U.S. Environmental Protection Agency (EPA) by National Pollutant Discharge Elimination System (NPDES) Permit AK-002255-1 for discharge from the John M. Asplund Water Pollution Control Facility (WPCF), operated by the Anchorage Water and Wastewater Utility (AWWU) at Point Woronzof under authority of the Municipality of Anchorage (MOA). The NPDES permit incorporates provisions necessitated by the Clean Water Act (CWA) Section 301(h) variance from the requirements of secondary treatment.

The elements of the monitoring program are:

- Influent, Effluent, and Sludge Monitoring
  - In-Plant Sampling
  - Toxic Pollutant and Pesticide Sampling
  - Pretreatment Monitoring
  - Whole Effluent Toxicity Monitoring
- Receiving Water Quality Monitoring
  - Plume Dispersion
  - Intertidal Zone Bacteria
- Sediment and Bioaccumulation Monitoring
  - Sediment Analyses
  - Bioaccumulation Analyses

During 2010, the monitoring program consisted of sampling and analysis of the influent, effluent, and sludge twice for toxic pollutants and pesticides, one receiving water quality sampling and analysis effort, and quarterly whole effluent toxicity (WET) testing. In addition, AWWU conducted the required self-monitoring program for the influent, effluent, and sludge. The sediment and bioaccumulation components of the program were conducted during 2003 and 2004 and were required to be performed only once by the current NPDES permit.

This annual report provides information and data pertaining to the monitoring program performed to meet the requirements as set forth in the NPDES permit that became effective on 2 August 2000. The report covers the period of 1 January through 31 December 2010.



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## **APPENDICES (CD ATTACHED)**

### **APPENDIX A Toxic Pollutant and Pesticide Monitoring, July Sampling**

- A1 Oil & Grease, Cyanide, Metals, Priority Pollutants, Dioxin, Pesticides, and Hydrocarbons  
SGS North America, Test America, & Columbia Analytical Services
- A2 Asbestos  
Solar Environmental Services, Inc. and IATL
- A3 Enterococci  
AWWU Asplund Laboratory

### **APPENDIX B Toxic Pollutant and Pesticide Monitoring, August Sampling**

- B1 Oil & Grease, Cyanide, Metals, Priority Pollutants, Dioxin, Pesticides, and Hydrocarbons  
SGS North America, Inc. and Test America
- B2 Asbestos  
Solar Environmental Services, Inc. and IATL
- B3 Enterococci  
AWWU Asplund Laboratory

### **APPENDIX C Receiving Water Quality Monitoring**

- C1 Total Suspended Solids and Cyanide  
Soil Control Lab
- C2 Aromatic Hydrocarbons and Color  
SGS North America, Inc.
- C3 Polycyclic Aromatic Hydrocarbons  
Texas A&M GERG
- C4 Trace Metals  
Battelle
- C5 Receiving Water & Intertidal Bacteria  
AWWU Asplund Laboratory and Spectra Laboratories
- C6 Total Residual Chlorine and Turbidity  
Kinnetic Laboratories, Inc.
- C7 Hydrographic Data Profiles  
Kinnetic Laboratories, Inc.
- C8 Field Data Sheets  
Kinnetic Laboratories, Inc.



## **SUMMARY**

### **PURPOSE**

This report is submitted to meet the requirements of the U.S. Environmental Protection Agency (EPA) and the Alaska Department of Environmental Conservation (ADEC) as outlined in the National Pollutant Discharge Elimination System (NPDES) Permit AK-002255-1 that was signed on 30 June 2000 and became effective on 2 August 2000. This permit authorizes discharge of effluent from the John M. Asplund Water Pollution Control Facility (Asplund WPCF). Wastewater from the Municipality of Anchorage is treated at this facility before discharge to the receiving waters of Knik Arm in Cook Inlet, Alaska. The NPDES permit incorporates the requirements necessitated by the Clean Water Act (CWA) 301(h) variance from secondary treatment and is in compliance with provisions of the Federal Water Pollution Control Act as amended by the CWA (33 U.S.C. §1251 et seq.) and the Water Quality Act of 1987.

### **HISTORY**

In September 1979, the AWWU submitted to the EPA a 301(h) secondary treatment variance application proposing an improved discharge which eliminated chlorination and required the addition of both a 610-meter (m) extension and a 305-m diffuser to the Asplund WPCF outfall. The outfall extension was intended to move the point of discharge beyond the negative influence of a gyre that was reported to exist off Point Woronzof on a flood tide and was presumed to carry effluent toward shore, causing bacterial contamination of the shoreline.

Further studies were subsequently undertaken to derive design criteria for the outfall improvements. The central issue was to evaluate outfall design alternatives and the chlorination/no chlorination option in relation to a system of eddies that occur on the flood tide. These studies were completed as an Amendment to the Wastewater Facilities Plan for Anchorage, Alaska (CH2M Hill et al., 1985). This amended plan recommended the use of the existing 245-m outfall with the addition of a three-nozzle diffuser. It was shown that chlorination would still be required to meet bacterial standards even with an extended outfall and diffuser. Because the same water quality standards could be met by chlorinating and installing an improved diffuser at the end of the existing outfall, there was no need to extend the outfall.

Concurrent with the studies to amend the facilities plan, a revised CWA 301(h) variance application was submitted to the EPA. After extensive EPA review, public comment, and hearings, the Final Permit Decision was issued by EPA and the five-year NPDES permit became effective 16 October 1985 (EPA, 1985a). As required by this permit, a multi-port diffuser was installed in August 1987 prior to the second year of receiving water sampling. Fourteen years of monitoring were performed under the initial NPDES permit.

The AWWU submitted an application to renew the CWA 301(h) variance from secondary treatment in 1990. A more recent application was submitted in 1998 with additional information provided to EPA in 1999. A draft NPDES permit that incorporated the 301(h) variance was issued in 1999 for public comment. The renewed permit was signed by EPA on 30 June 2000 to become effective on 2 August 2000 for five years. The permit was administratively extended in August 2005 pending a permit renewal decision from EPA. The most recent application for a reauthorization of the NPDES permit and CWA 301(h) variance was submitted in January 2005 and is currently being reviewed by EPA.

## **RECEIVING WATER ENVIRONMENT**

The Asplund WPCF discharges into the Knik Arm of Cook Inlet, a unique body of estuarine water with extremely high tidal fluctuations (over 39 feet [12 meters] with a mean range of 7.98 meters [m] at Anchorage; NOAA/NOS, 2010). These fluctuations produce extensive tidal flats, swift tidal currents of 4 - 6 knots, and intense mixing within Cook Inlet. The continual input of sediments, combined with the re-suspension of bottom sediments due to high bottom shear stress with each tidal cycle, results in naturally high suspended sediment concentrations of up to 2500 milligrams/liter (mg/L) in Knik Arm. This sediment originates from riverine and glacial melt waters flowing into Cook Inlet and Knik Arm from the Knik, Matanuska, and Susitna Rivers.

Large temperature extremes occur between summer and winter. In the winter, ice can reach thicknesses of 1 - 2 m and consists of broken pieces due to the large tides and currents. An important consideration to this monitoring study is the large volume of saline water that enters Cook Inlet that is completely vertically mixed with the riverine and glacial inputs by tidal turbulence which allows this water body to be very effective in wastewater dilution and assimilation.

## **MONITORING OBJECTIVES**

The monitoring that was conducted during 2010 consisted of two main components: (1) in-plant monitoring of influent, effluent, and sludge, including whole effluent toxicity testing; and (2) receiving water quality monitoring in the vicinity of the discharge and mixing zone, and at a control site across Knik Arm. Objectives of the 2010 program as outlined in the permit are:

### **2010 MONITORING OBJECTIVES**

#### **Influent, Effluent, and Sludge Monitoring**

- Determine compliance with the NPDES permit and State of Alaska water quality criteria
- Determine effectiveness of the industrial pretreatment program
- Aid in assessing the water quality at discharge point
- Characterize toxic substances
- Monitor plant performance
- Determine compliance with the regulatory criteria of Section 301(h) of the CWA
- Provide data for evaluating re-issuance of the NPDES permit

#### **Receiving Water Quality Monitoring**

- Determine compliance with the NPDES permit and State of Alaska water quality criteria
- Aid in assessing the water quality of the receiving water
- Determine compliance with the regulatory criteria of Section 301(h) of the CWA
- Determine the level of bacterial concentrations in nearshore waters
- Provide data for evaluating re-issuance of the NPDES permit

## **MONITORING RESULTS**

As part of its self-monitoring program, AWWU conducted daily, weekly, and monthly sampling of influent, effluent, and sludge, depending on the parameter measured. In addition, monitoring

for toxic pollutants and pesticides was conducted twice during 2010, once in July and once in August. Whole effluent toxicity testing was conducted quarterly, while receiving water quality monitoring was performed once in July. The following summarizes results of this year's monitoring based on the permit requirements:

## **2010 MONITORING RESULTS**

### **Influent, Effluent, and Sludge**

- The influent, effluent, and sludge monitoring showed, that with no exceptions, the Asplund WPCF met the NPDES permit requirements and complied with all applicable AWQS. AWWU's self-monitoring of TRC, pH, fecal coliform bacteria, BOD<sub>5</sub>, and TSS showed compliance with all permit effluent limitations throughout 2010.
- AWWU's self-monitoring of TRC and pH showed that the permit limit for daily maximum TRC levels in the effluent was never exceeded and pH was within permit limits throughout 2010.
- The permit limit for the monthly maximum geometric mean of 850 fecal coliform colonies per 100 mL by most probable number (FC MPN/100 mL) technique was not exceeded in 2010. The fecal coliform monthly criteria "that not more than 10 % of the effluent samples shall exceed 2600 FC MPN/100 mL" was also not exceeded in any month during 2010.
- AWWU's self-monitoring of TSS and BOD<sub>5</sub> showed compliance with both regulatory and permit effluent limitations. TSS and BOD<sub>5</sub> were well within the daily, weekly, and monthly criteria for the entire year. Average monthly removals for BOD<sub>5</sub> and TSS of greater than 30 % are required by the amendment to the CWA (40 CFR Part 125; Final Rule, 8/9/94). The removal rate for both TSS and BOD<sub>5</sub> met the 30 % minimum removal requirement for all months during 2010. Annual removals were 79 % for TSS and 48 % for BOD<sub>5</sub> which indicate an exceptional level of primary treatment.
- Total aromatic hydrocarbons (TAH), total aqueous hydrocarbons (TAqH), and total ammonia concentrations in the effluent were all found to be below their maximum allowable effluent concentrations (MAECs) throughout 2010.
- Concentrations of metals and cyanide in the effluent never exceeded their MAECs at any time during any of the 2010 monitoring events.
- Concentrations of toxic pollutants and pesticides, including metals and cyanide, in the influent and effluent were all within the established range or lower than values from a national study of secondary treatment plants (EPA 1982a).
- Toxic pollutant sludge concentrations were found to be very low compared to the limits established under 40 CFR Part 503 and most were either not-detected or within the established range or lower than values from a national study of secondary treatment plants. Most metals fell at or below the typical concentrations and all metals were below 95<sup>th</sup> percentile worst case values (EPA 1985c).
- Whole effluent toxicity testing conducted quarterly met all permit limitations for toxicity for all tested species and all monitoring events in 2010.

## Water Quality

- Little variation among stations was observed for most hydrographic parameters indicating that the receiving water environment is uniform and well mixed near the outfall.
- To test the hypothesis that the water quality at the ZID boundary was not degraded with respect to the water quality at the nearfield and control stations, statistical comparisons were employed. Conventional parameters such as salinity and temperature did show significant differences between sites, however, these were not ascribed to the outfall but were due to river influences at the control stations. No statistically significant differences were seen for dissolved oxygen, pH, turbidity, TRC, TSS, color, or fecal coliform.
- Fecal coliform concentrations in receiving water and intertidal samples were found to be low at most locations. AWQS criteria of a median of not more than 14 FC MPN/100 mL, a geometric mean of not more than 20 FC MPN/100 mL, and of not more than 10 % of the samples exceeding 40 FC MPN/100 mL were met at all receiving water locations. Intertidal stations exceeded the criteria of not more than 10 % of samples exceeding 40 FC MPN/100 mL as a result of two elevated samples of unknown source at one location.
- Supplemental receiving water quality samples obtained as part of the plume dispersion monitoring indicated that dissolved metals were all below the AWQS at all locations on the ZID boundary and outside of the ZID including the control stations. No statistically significant differences between the outfall and control station groupings were seen for any dissolved metal. Total metals were elevated compared to the dissolved as a result of the naturally high suspended sediment load. Statistically significant differences were seen for arsenic, cadmium, and copper due to higher ambient TSS at the control stations.
- All cyanide concentrations in the receiving waters were found to be below the receiving water quality criterion limit of 1.0 µg/L and no statistically significant differences were detected between concentrations at the control and outfall stations.
- Supplemental receiving water samples also demonstrated that TAH and TAqH met the AWQS at all locations. No statistically significant differences were detected between the control and outfall stations for TAH. TAqH did exhibit significant differences due to higher levels seen at the within mixing zone station, but levels were still within AWQS.
- Turbidity and color met the AWQS at all stations. TRC was at or below the detection limit of 10 µg/L at all locations including those located within the ZID, as compared to the AWQS of 7.5 µg/L for chronic, 13.0 µg/L for acute marine water use, and 100 µg/L as ADEC's practical quantitation limit for regulatory purposes. Based on the highest maximum daily effluent TRC concentration of 880 µg/L seen during 2010 and a 180:1 effluent dilution credit, it is expected that TRC in the receiving water would be <5 µg/L before reaching the ZID boundary and would always meet all AWQS for TRC.

## CONCLUSIONS

Results from the past year of the monitoring program confirm previous studies, data in the 301(h) variance renewal application, and the decision by the EPA to reissue the NPDES permit with 301(h) variance. The Asplund WPCF is operating within regulatory requirements with no exceptions seen in 2010 and is showing no measurable impacts to the marine environment.



## **1.0 INTRODUCTION**

### **1.1 REGULATORY/ENVIRONMENTAL BACKGROUND**

The monitoring program is designed to meet the requirements of the NPDES Permit AK-002255-1 which authorizes discharge of municipal effluent into the Knik Arm of Cook Inlet receiving waters from the John M. Asplund WPCF, operated by the AWWU under authority of MOA (Figure 1). The NPDES permit, which became effective on 2 August 2000, incorporates the requirements necessitated by the CWA 301(h) secondary treatment variance and is in compliance with provisions of the Federal Water Pollution Control Act as amended by the Clean Water Act (CWA 33 U.S.C. §1251 et seq.) and the Water Quality Act of 1987, P.L. 100-4.

#### **1.1.1 Regulatory Background**

In 1972, while the Asplund WPCF and outfall were being built for the MOA, the Federal Water Pollution Control Act (FWPCA) was amended to establish two phases of effluent limitations applicable to all Publicly Owned Treatment Works (POTWs). Under Section 301(b), POTWs were required to achieve secondary treatment of effluent by 1 July 1977 and the "best practicable waste treatment technology" by July 1983.

Congress again amended the FWPCA in 1977. Section 301(h) was added, providing that the Administrator of the EPA, upon application from a POTW and with the concurrence of the State, might issue an NPDES permit modifying the requirements of Section 301(b). On 15 June 1979, EPA promulgated the regulations regarding the issuance of this variance from secondary treatment to an applicant discharging into certain ocean and estuarine waters and demonstrating compliance with the 301(h) criteria.

In September 1979, the AWWU submitted to the EPA a 301(h) variance application proposing an improved discharge which eliminated chlorination and required the addition of both an extension and diffuser to the Asplund WPCF outfall. Earlier studies had recommended the construction of a 610-m outfall extension and a 305-m diffuser. The proposed extension/diffuser reportedly could meet fecal coliform receiving water standards without chlorination and prevent shore contact of the wastewater plume.

As a parallel program, the AWWU undertook preparation of a wastewater master plan for the Anchorage area. The resultant Wastewater Facilities Plan for Anchorage, (Ott Water Engineers, Inc. et al., 1982) and the Environmental Impact Statement, City of Anchorage, Alaska, Wastewater Facilities (EPA and Jones & Stokes, 1982) were accepted by the EPA and ADEC.

Further studies were subsequently undertaken to derive design criteria for the outfall improvements. Significant efforts were included in this study to improve the reconnaissance level data upon which the outfall length and diffuser design were to be based and to evaluate bacterial standards applicable to Knik Arm. The central issue was to evaluate outfall design alternatives and the chlorination or no-chlorination option in relation to the presence of a system of eddies that occur to the east of Point Woronzof on the flood tide which might be capable of transporting the effluent shoreward. These latter studies were completed as an Amendment to the Wastewater Facilities Plan for Anchorage, Alaska (CH2M Hill et al., 1985). This amended plan recommended use of the existing 245-m outfall with the addition of a three-nozzle diffuser. It was shown that chlorination would be required to meet bacterial standards even with an

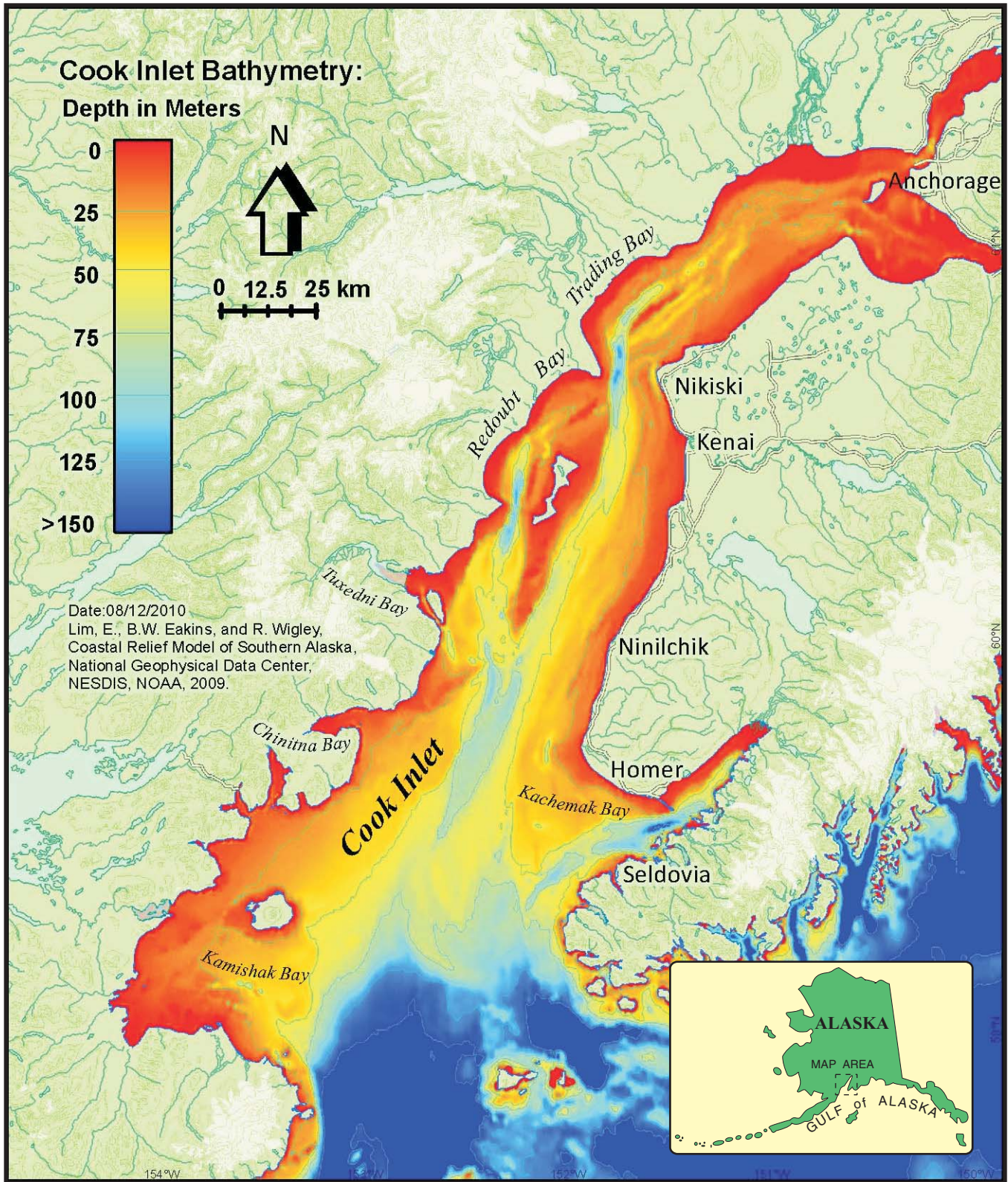


Figure 1. General Study Area and Bathymetry.

extended outfall and diffuser. Because the same standards could be met by use of chlorination and the existing outfall, there was no need to extend the outfall. With continued chlorination, all water quality standards were predicted to be met by the amended facilities plan.

Concurrent with the studies to amend the facilities plan, a revised application entitled Application for Modification of Secondary Treatment Requirements, Section 301(h), Clean Water Act was submitted to the EPA (CH2M Hill et al., 1984). The EPA Region 10 301(h) Review Team's Tentative Decision Document, entitled Analysis of the Section 301(h), Secondary Treatment Variance Application for the Asplund WPCF (EPA, 1985b), and a draft NPDES permit were made available for public comment on 17 January 1985. After comments and appropriate hearings, the Final Permit Decision (EPA, 1985a) was issued 13 September 1985, and the start date of the five-year NPDES Permit AK-002255-1 was listed as 16 October 1985. As required by this permit, a multi-port diffuser was installed at the Asplund WPCF outfall in the beginning of August 1987. This occurred prior to the 1987 summer water quality monitoring program. This original NPDES permit expired on 15 October 1990.

The AWWU submitted a renewal application for the permit in April 1990 which addressed amendments made to the 301(h) provisions by the Water Quality Act. That renewal application was not acted upon by the EPA and the facility continued to operate under an administrative extension of the 1985 permit until August 2000. In 1998 it was projected that the growth of Anchorage would result in the discharge limits contained in the 1985 permit being exceeded within a few years. Therefore, the AWWU prepared and submitted another renewal application which replaced the 1990 application in October 1998 (CH2M Hill, 1998).

In tandem with the renewal application, the AWWU conducted special studies and submitted a request for site-specific water quality criteria (SSWQC) to the ADEC for the Point Woronzof area of Cook Inlet in December 1998. This request for SSWQC was for turbidity and a suite of metals and was necessitated because the Alaska Water Quality Standards (AWQS) for marine waters could not be achieved for these waters as a result of the naturally high suspended sediment loads in Cook Inlet due to glacial inputs. The approach to the request was based on the EPA's Metals Policy that had been recently promulgated which recommends the use of only the dissolved fraction of metals as bioavailable and appropriate for the protection of aquatic life and associated beneficial uses of the water body. Following both agency and public review and comments, the SSWQC were incorporated into the AWQS as amended on 27 May 1999. The SSWQC for the Point Woronzof area included turbidity and the dissolved fraction of arsenic, cadmium, hexavalent chromium, copper, lead, mercury, nickel, selenium, silver, and zinc.

Following the promulgation of these new AWQS, a tentative decision to grant the AWWU its 301(h) variance was made by the EPA on 4 November 1999. The tentative decision, draft NPDES permit, and permit fact sheet were then made available for public review and comments. The State of Alaska's Division of Government Coordination issued its Final Consistency Determination for the action in February 2000. The current NPDES permit for the Asplund WPCF was signed by the EPA and went into effect 2 August 2000 for five years, and was then administratively extended in August 2005 pending permit renewal. The most recent application for a reauthorization of the NPDES permit with 301(h) variance was submitted in January 2005 and is still under review by the EPA.

The NPDES permit specifies the required monitoring program. The Monitoring Program Plan (Kinnetic Laboratories, Inc., 2000a), submitted to the EPA in October 2000, identified how the



AWWU plans to fulfill the requirements of this monitoring program. This annual report documents the progress and results of the monitoring program that was performed in 2010.

Since the issuance of the current NPDES permit, EPA has approved ADEC's proposed use of dissolved metals for the AWQS, approved all of the proposed SSWQC for Upper Cook Inlet in the vicinity of Pt. Woronzof, and removed Alaska from the National Toxic Rule (EPA, 2006). In September of 2009 EPA approved the 2009 revisions to the AWQS and the December 2008 State of Alaska Toxics Manual which lists numerical limits. Except for cadmium and mercury where the dissolved cadmium standard changed from 9.3 µg/L in the SSWQC to 8.8 µg/L in the AWQS and mercury which changed from 0.025 µg/L in the SSWQC to 0.94 µg/L in the AWQS, all other dissolved metals criteria are the same between the two standards.

### **1.1.2 Environmental Background**

The Asplund WPCF discharges to the receiving waters of Cook Inlet, Alaska. The discharge is located off Point Woronzof in Knik Arm of Upper Cook Inlet.

Cook Inlet is a major tidal estuary that is approximately 333 kilometers (km; 180 nautical miles) long and 93 - 148 km (50 - 80 nautical miles) wide at its lower end with a large assimilative capacity. Bathymetry indicates the Inlet is deep, generally 36.6 m (20 fathoms) north of the Forelands and about 164.6 m (90 fathoms) at the mouth (refer to Figure 1). Numerous rivers, including the major Knik, Matanuska, and Susitna River drainages, discharge into the Inlet. A detailed map of the Point Woronzof region indicates deep water (9.1 - 51.8 m) extending well past Anchorage up the Knik Arm (Figure 2).

Cook Inlet is a unique estuary, with perhaps the closest parallel being the Bay of Fundy between New Brunswick and Nova Scotia, Canada. The occurrence of tidal bores at the head, currents of 4 - 6 knots, suspended loads of up to 2500 mg/L, large temperature extremes, and moving pancake ice of up to one meter (m) thick make Cook Inlet unique. The high tidal ranges result from the geometry of the Inlet which has a natural resonance period close to the semi-diurnal tidal period. The resulting large tidal fluctuations and fast currents cause complete vertical mixing of the Inlet waters including any discharges into those waters.

Another important factor to this study is the large volume of saline water that enters Cook Inlet that is completely vertically mixed with the riverine inputs by tidal turbulence. This allows the water body to be very effective in wastewater dilution and assimilation.

The particle size distributions of the natural suspended sediments near Point Woronzof show that very large particles are suspended by the current-generated turbulence, with 50 percent of the load being in the size range of 65 - 250 microns. The settling of large particles is seen at slack tide, but due to the shifting currents, never settle completely. Settling rate tests of the suspended material show that 93% of the solids in an ambient water sample settle in twenty minutes,

Previous work has indicated that due to the extremely swift currents, no seabed accumulation of suspended sediments, either natural or from the discharge, occur in the vicinity of the outfall. In this location, the bottom is strictly coarse gravel and cobble because of these currents. However, areas of deposition do exist, such as to the east of Point Woronzof, where mudflats and beaches are found, and to the southwest of the Point. The area between Fire Island and the mainland is hard-packed sand with no deposition of silt or finer materials as a result of the high current

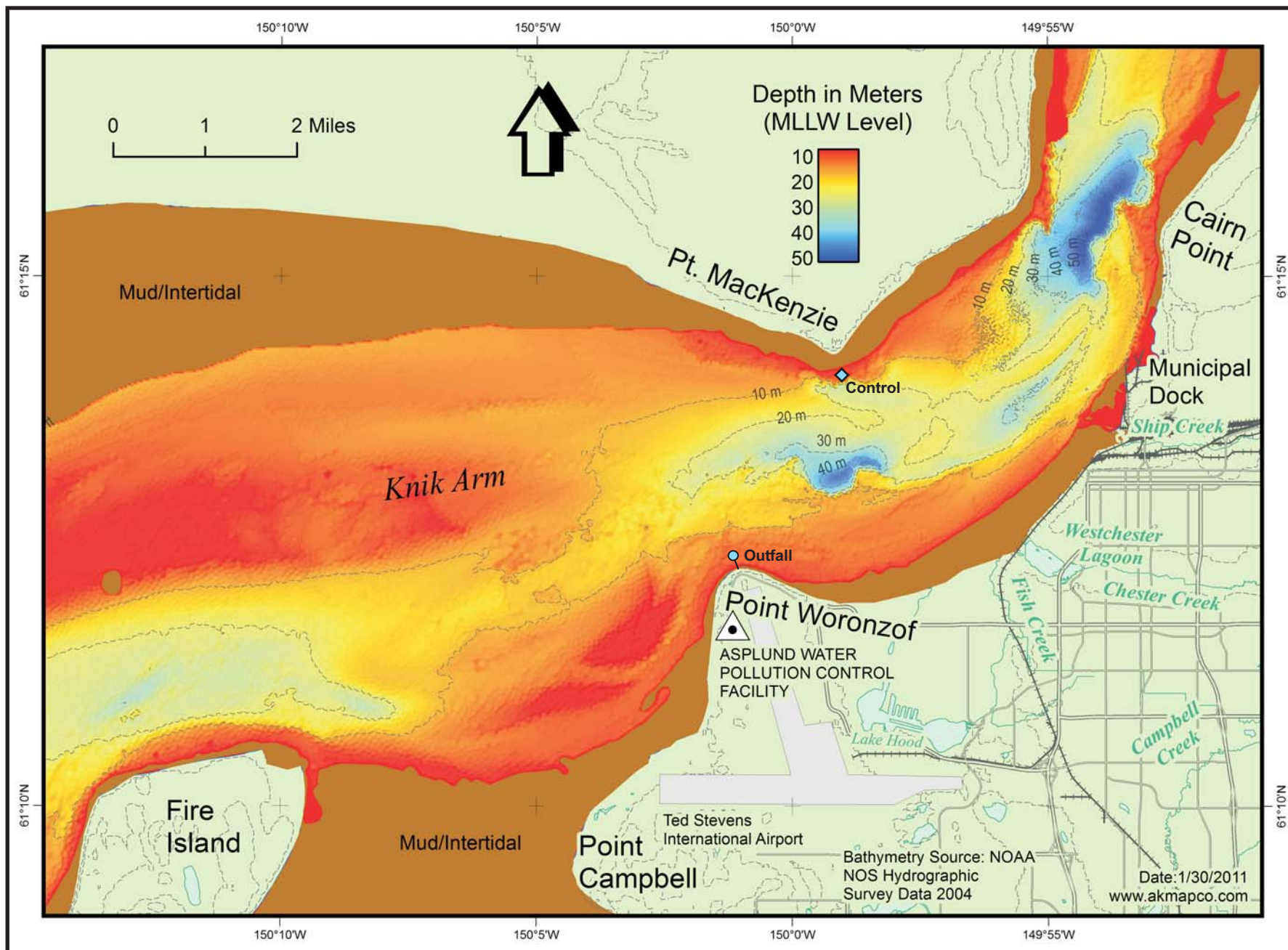


Figure 2. Asplund WPCF Outfall and Control Station Location.

energy. Of course, any suspended solids in these materials of effluent origin would actually dilute the much larger natural load in the receiving water (400 - 2,500 mg/L versus approximately 50 mg/L effluent). Studies have also shown that essentially no benthic biota are found on the scoured cobble/gravel bottom or on the rock beaches at both Point Woronzof and the control area. Similar sampling of soft bottom beaches and tidal flats showed very sparse abundances and very low diversities. Benthic and intertidal marine fauna populations are limited by the harsh physical environment of mud and silt, high turbulence and bottom scouring, large tide and strong currents, and extreme ice conditions.

Current trajectories in the immediate vicinity of the outfall are of concern because of flow separation zones on either side of Point Woronzof. Previous work has indicated that, on a flood tide, a clockwise eddy sometimes exists east of Point Woronzof. This eddy may result in the shoreward transport of wastes at certain stages of tide. A flow separation also exists to the west of Point Woronzof during ebb flow; however the effluent is not entrained shoreward in this area.

## **1.2 STUDY DESIGN**

### **1.2.1 Monitoring Objectives**

The monitoring program as described by NPDES Permit No. AK-002255-1 includes influent, effluent, and sludge monitoring at the Asplund WPCF; receiving water and sediment quality monitoring; biological and toxicological monitoring; and a toxics control program. The objectives of the overall monitoring program as outlined in the NPDES permit are to:

- Determine compliance with the NPDES permit
- Determine compliance with State water quality criteria
- Determine effectiveness of the industrial pretreatment program
- Aid in assessing the water quality at the discharge point
- Characterize toxic substances
- Monitor plant performance
- Determine compliance with the regulatory criteria of Section 301(h) of the CWA
- Determine the level of bacterial concentrations in nearshore waters
- Monitor for changes in sediment quality (organic enrichment, alteration of grain size distribution, and pollutant contamination) (note: not required or performed in 2010)
- Determine if pollutants from the discharge are accumulating in exposed biological organisms (note: not required or performed in 2010)
- Provide data for evaluating re-issuance of the NPDES permit

### **1.2.2 Program Description**

The elements of the monitoring program for the Asplund WPCF are:

- Influent, Effluent, and Sludge Monitoring, including
  - In-Plant Sampling
  - Toxic Pollutants and Pesticides (including Metals and Cyanide)
  - Pretreatment Monitoring
  - Whole Effluent Toxicity (WET) Testing
- Receiving Water Quality Monitoring, including

- Plume Dispersion and Water Quality
- Intertidal Bacteria
- Biological and Sediment Monitoring, including
  - Sediment Quality
  - Bioaccumulation

Table 1 provides an overview of the monitoring requirements as described by the permit. Detailed information regarding each program component is provided in Section 2.0, Methods.

### 1.2.3 Hypotheses

Hypotheses were formulated for the monitoring program as an unbiased approach in determining whether the Asplund WPCF was affecting the marine receiving water environment. The null (no effect) hypotheses tested for this year of monitoring are as follows:

*H<sub>o1</sub>: Applicable State and Federal effluent and receiving water standards were met by the Asplund WPCF discharge.*

*H<sub>o2</sub>: Water quality at the boundary of the ZID was not significantly changed with respect to nearfield or control stations.*

## 1.3 CONTRACTOR

The AWWU's designated contractor for the 2010 Asplund WPCF Monitoring Program was Kinnetic Laboratories, Inc. (KLI) of Anchorage, Alaska.

For influent, effluent, and sludge monitoring, aromatic hydrocarbon, pesticide, and volatile and semi-volatile priority pollutant analyses (gas chromatography/mass spectrometry scans), trace metals (total and dissolved) for the toxic pollutant, pesticide, and pretreatment monitoring were performed by SGS North America, Inc. of Anchorage, Alaska, Test America, and Columbia Analytical Services. WET testing was performed by ToxScan, Inc. of Watsonville, California. Asbestos analyses were performed by Solar Environmental Services, Inc. of Anchorage, Alaska, and IATL, Inc. In addition, AWWU's Asplund WPCF Laboratory performed the monthly in-plant analyses as part of its self-monitoring program and contracted the Part 503 sludge analyses to SGS North America, Inc. of Anchorage, Alaska.

KLI performed the receiving water sampling and analyses for turbidity and total residual chlorine (TRC). Analytical support for the receiving water sampling included: Battelle (Sequim, Washington) for trace metals; Soil Control Lab (Watsonville, California) for total suspended solids (TSS) and cyanide; SGS North America, Inc. for aromatic hydrocarbons and color; AWWU's Asplund WPCF Laboratory and Spectra Laboratories in (Tacoma, Washington) for bacteriology; and Texas A&M University's Geochemical and Environmental Research Group (GERG) (Texas) for polycyclic aromatic hydrocarbons (PAHs) analyses.

## 1.4 PERIOD OF REPORT

This report documents the progress and results of the monitoring program from 1 January through 31 December 2010 under the current NPDES permit.

**Table 1. Overall Monitoring Requirements.**

Parameter	Frequency	Sample Type	Remarks
In-Plant Sampling	See Table 2	See Table 2	See Table 2 - includes flow, TRC, DO, BOD <sub>5</sub> , TSS, temperature, pH, fecal coliform, total ammonia as nitrogen, enterococci bacteria, and oil and grease
Toxic Pollutants and Pesticides (including Metals and Cyanide)	2/year <sup>a</sup>	influent, 24-hr composite effluent, 24-hr composite sludge, 24-hr composite	See Table 2
Pretreatment Program	2/year <sup>a,b</sup>	influent, three 24-hr composite effluent, three 24-hr composite sludge, 24-hr composite	Includes metals and cyanide plus percent solids for sludge
Whole Effluent Toxicity (WET) Testing	4/year <sup>c</sup>	effluent, 24-hr composite	See Table 2
Receiving Water Quality	1/year <sup>d</sup>	receiving water	See Table 5
Intertidal Bacteria	1/year <sup>e</sup>	intertidal receiving water	Fecal coliform sampling at 8 intertidal stations
Sediment	Once during the fourth year of the permit <sup>e</sup>	grab samples of surficial (0-2 cm) sediment collected at intertidal and subtidal stations <sup>f</sup>	Includes total volatile solids (TVS), toxic pollutants and pesticides (including metals and cyanide), and sediment grain size distribution
Bioaccumulation	Once during the fourth year of the permit	grab samples of intertidal macroalgae ( <i>Vaucheria</i> spp.)  Note: Macroalgae was not available during 2003 or 2004. Therefore, in consultation with EPA and AWWU pacific cod ( <i>Gasdus macrocephalus</i> ) were collected and analyzed for this permit component in October 2004 <sup>g</sup>	Includes toxic pollutants and pesticides (including metals and cyanide)

<sup>a</sup> Sampling will be conducted twice per year: once in summer dry conditions and once in summer wet conditions.

<sup>b</sup> The first day of three consecutive days of sampling will be part of the Toxic Pollutant and Pesticides (metals and cyanide) sampling performed twice each year.

<sup>c</sup> WET testing will be performed on a quarterly basis.

<sup>d</sup> Sampling will be conducted once per year in summer dry conditions.

<sup>e</sup> Sampling will be conducted in conjunction with the receiving water sampling.

<sup>f</sup> Sampling will be performed at Intertidal Stations 1, 2, and Control (IT-1, IT-2, and IT-C); a subtidal station located at the ZID boundary, and a subtidal control station near Point MacKenzie (in a similar water depth as the ZID boundary).

<sup>g</sup> Sampling was to be performed in conjunction with the sediment analyses however algae was not available in sufficient quantities for sampling in 2003 or 2004. Pacific cod were collected and analyzed for this permit component in October 2004.



## 2.0 METHODS

### 2.1 INFLUENT, EFFLUENT, AND SLUDGE MONITORING

Influent, effluent, and sludge monitoring is outlined in Table 2. Routine daily, weekly, and monthly sampling of conventional pollutant parameters and flow rate were performed by AWWU. The less-frequently monitored parameters of enterococci bacteria, oil and grease, toxic pollutants and pesticides (including metals and cyanide), and Whole Effluent Toxicity (WET) testing were handled by KLI.

- ✓ determine compliance with the NPDES permit and State of Alaska water quality criteria
- ✓ determine effectiveness of the industrial pretreatment program
- ✓ aid in assessing the water quality at the discharge point
- ✓ characterize toxic substances
- ✓ help monitor plant performance
- ✓ determine compliance with the regulatory criteria of Section 301(h) of the CWA
- ✓ provide data for evaluating re-issuance of this permit

#### 2.1.1 In-Plant Monitoring

In-plant influent, effluent, and sludge sampling was performed by AWWU personnel as described in Table 2 and in a separate monitoring program plan prepared by AWWU (AWWU, 2000). Samples were obtained following the schedule of frequency required by the permit. Influent was sampled at a representative location in the influent headworks, upstream from the recycle streams. Effluent was sampled at a well-mixed point downstream from the chlorination input point in the final effluent line. Composite sludge samples were obtained from the sludge feed screw auger downstream of the addition of primary scum. Grab samples were obtained for total residual chlorine (TRC), dissolved oxygen (DO), temperature, pH, and fecal coliform. Composite samples were obtained for analysis of biochemical oxygen demand (BOD<sub>5</sub>), total suspended solids (TSS), and total ammonia as nitrogen.

#### 2.1.2 Toxic Pollutant and Pesticide Monitoring

As outlined in the permit, toxic pollutant and pesticide sampling was conducted twice this year, once during July 2010 (summer dry) and once during August 2010 (summer wet). Samples were collected as required by the permit and either analyzed by AWWU laboratory personnel or provided to KLI for shipment to the appropriate analytical laboratory. Plant influent was sampled as discrete grabs or by flow-proportional composite samplers (depending on the analysis method) at a representative location in the influent headworks upstream from the recycle streams. Effluent was sampled as discrete grabs or using flow-proportional samplers at a well-mixed point downstream from the chlorination injection point in the final effluent line. Influent and effluent samples were chilled as required during composite sampling. Composite sludge grab samples were obtained from the sludge feed screw auger.

Samples were composited for the analysis of pesticides, semi-volatile organics, metals, asbestos, and cyanide. Samples consisted of composites of flow-proportioned samples collected over a 24-hour (hr) period using two Teledyne ISCO Model 4700 Refrigerated Autosamplers. Grab samples for volatile organics analysis were collected every three hours during the 24-hr sampling period and composited by the laboratory prior to analysis. Grab samples were collected for analysis of total hydrocarbons as oil and grease and purgeable aromatic compounds.

**Table 2. Influent, Effluent, and Sludge Monitoring Requirements.**

Parameter	Sample Point <sup>a</sup>	Sample Frequency	Sample Type
Flow <sup>b</sup>	effluent	continuous	continuous
Total Residual Chlorine (TRC) <sup>b</sup>	effluent	continuous <u>or</u> every 2-4 hrs	grab
Dissolved Oxygen (DO) <sup>b</sup>	effluent	4/week	grab
Biochemical Oxygen Demand (BOD <sub>5</sub> ) <sup>b</sup>	influent and effluent	4/week	24-hr composite
Total Suspended Solids (TSS) <sup>b</sup>	influent and effluent	4/week	24-hr composite
Temperature <sup>b</sup>	influent and effluent	4/week	grab
pH <sup>b</sup>	influent and effluent	4/week	grab
Fecal Coliform Bacteria <sup>b</sup>	effluent	3/week	grab
Total Ammonia as N <sup>b</sup>	effluent	1/month	24-hr composite
Enterococci Bacteria <sup>b</sup>	effluent	2/year <sup>d</sup>	grab
Oil and Grease <sup>c</sup>	effluent	2/year <sup>d</sup>	grab
Toxic Pollutants and Pesticides (including Metals and Cyanide) <sup>e</sup>	influent, effluent, and sludge	2/year <sup>d</sup>	24-hr composite
WET <sup>f</sup>	effluent	4/year <sup>f</sup>	24-hr composite

<sup>a</sup> When both influent and effluent samples are required, samples will be collected during the same 24-hr period.

<sup>b</sup> AWWU will perform this monitoring component.

<sup>c</sup> KLI will perform this monitoring component.

<sup>d</sup> Twice per year sampling: once during summer in dry conditions and once in wet conditions.

<sup>e</sup> As part of the pretreatment program sampling requirements, arsenic, cadmium, chromium, copper, cyanide, lead, mercury, molybdenum, nickel, silver, and zinc in influent, effluent, and sludge will be sampled, along with percent solids (in sludge only). In 2010, these metals were analyzed and reported by as total recoverable metals and dissolved metals for influent and effluent and as total metals in mg/kg dry weight for sludge. Sampling will be as follows: Influent and effluent as three separate 24-hr composite samples taken on 3 consecutive days (Mon - Fri), the first day of which coincides with the twice yearly sampling (summer-dry and-wet conditions); sludge as one composite of eight grabs/day when influent and effluent samples are being taken. In addition, the other four metals from the toxic pollutant list will be analyzed in the summer wet/summer dry samples: beryllium, antimony, thallium, and selenium.

<sup>f</sup> WET requirements are summarized in the text (Section 2.1.4). Initial testing will be a screening period performed during three quarters, during which three species will be tested to determine the most sensitive species. Re-screening will be performed each year during one quarter (different than the previous year) to determine the species to use for continued testing. Accelerated testing requirements will be triggered if chronic toxicity is greater than 143 TUC (chronic toxicity units, TUC=100/NOEC).

Sludge samples were collected from the sludge feed screw auger every three hours over a 24-hr period and the eight samples representatively composited prior to analysis.

At time of collection, all samples were appropriately labeled using pre-prepared, project-specific sample labels as described in Section 2.5. Sample collection and shipment was documented using project-specific chain of custody forms as described in Section 2.5.

Toxic pollutants as defined by the permit are those substances listed in 40 Code of Federal Regulations (CFR) 401.15 (Table 3). This list involves 65 categories of pollutants, including asbestos, aromatic hydrocarbons, pesticides, metals, and polychlorinated biphenyls (PCBs). Pesticides as defined in the permit are demeton, guthion, malathion, mirex, methoxychlor, and parathion as listed in 40 CFR 125.58. Other pesticides which were tested for are included on the toxic pollutants list (40 CFR 401.15). The methods that were used to analyze these constituents for the program and for which KLI was responsible, as well as those performed by AWWU, are also provided in Table 3. Preservation and maximum holding time information for each of these methods is provided in Table 4. All samples were collected in the appropriate precleaned sample containers and preserved, if necessary, as described by the EPA or equivalent method. All sample containers were immediately placed on gel ice after sampling. Samples remained chilled as required during shipment to the analytical laboratory.

### **2.1.3 Pretreatment Monitoring**

The pretreatment monitoring program as outlined in Table 1 and Table 2 was performed by the AWWU. This monitoring was performed twice in 2010 in conjunction with the summer dry and wet sampling. As part of the pretreatment program sampling requirements, arsenic, cadmium, chromium, copper, cyanide, lead, mercury, nickel, silver, and zinc in influent, effluent, and sludge were sampled, along with percent solids (in sludge only). These samples were analyzed by SGS North America Inc. as total recoverable metals and dissolved metals for influent and effluent and as total recoverable metals in dry weight for sludge. Sampling was conducted as follows: Influent and effluent as three separate 24-hr composite samples taken on 3 consecutive days (Monday - Thursday), the first day of which coincided with the twice-yearly toxic pollutant and pesticide sampling (summer dry and summer wet, respectively). The sludge sampling consisted of a single composite of eight grabs/day when influent and effluent samples were being taken. A detailed study plan describing this monitoring was provided previously (AWWU, 2000).

### **2.1.4 Whole Effluent Toxicity Testing**

As outlined in the permit, the WET testing must be performed on a quarterly basis on 24-hr flow-composited effluent samples. Effluent was sampled by discrete flow-proportional samplers at a well-mixed point downstream from the chlorination injection point in the final effluent line. Effluent samples were collected in the appropriate precleaned sample containers as described in the bioassay method, chilled, and shipped immediately to the toxicity laboratory for testing. Samples were appropriately labeled at the time of collection using pre-prepared, project-specific sample labels as described in Section 2.5. Sample collection and shipment were documented using project-specific chain of custody forms. Sample containers were immediately placed on gel ice after sampling and remained chilled during shipment to the toxicity laboratory.

**Table 3. Methods<sup>a</sup> for the Analysis of Toxic Pollutants and Pesticides for Influent, Effluent, and Sludge Monitoring.**

Volatile Organic Compounds	Semi-Volatile Organic Compounds	Pesticides and PCBs	Inorganic Compounds
<b>EPA 624 (Inf/Eff)</b> <b>SW 8260B (Sludge)</b> Benzene Chlorinated benzenes Dichlorobenzenes Ethylbenzene Toluene Xylenes <sup>b</sup>	<b>EPA 625 (Inf/Eff)</b> <b>SW 8270C (Sludge)</b> Acenaphthene Benzidine Chloralkyl ethers Chlorinated ethanes Chlorinated naphthalenes Chlorinated phenols 2-chlorophenol DDT & metabolites Dichlorobenzenes Dichlorobenzidine 2,4-dichlorophenol 2,4-dimethylphenol Dinitrotoluene Diphenylhydrazine Fluoranthene Haloethers Heptachlor & metabolites Hexachlorobutadiene Hexachlorocyclopentadiene Hexachloroethane Isophorone Naphthalene Nitrobenzene Nitrophenols Nitrosamines Polycyclic aromatic hydrocarbons (PAHs) Pentachlorophenol Phenol Phthalate esters	<b>EPA 614 (Inf/Eff)</b> <b>SW 8141A (Sludge)</b> Demeton Malathion Parathion Guthion <sup>b</sup>	<b>EPA 100.1/EPA 100.2 (Inf/Eff)</b> <b>Polarized Light Microscopy (PLM; Sludge)</b> Asbestos
<b>EPA 624 (Inf/Eff)</b> <b>SW 8260B (Sludge)</b> Acrolein <sup>b</sup> Acrylonitrile <sup>b</sup> Benzene Carbon tetrachloride Chloralkyl ethers Chloroform Chlorinated benzenes Chlorinated ethanes 1,2-dichloroethane Dichloroethylenes Dichloropropane Dichloropropene 1,1,1-trichloroethane Ethylbenzene Halomethanes Methylene chloride Bromoform Dichlorobromomethane Toluene Tetrachloroethylene Trichloroethylene Vinyl chloride	<b>EPA 608 (Inf/Eff)</b> <b>SW 8081A Pesticides and SW 8082 PCBs (Sludge)</b> Aldrin/Dieldrin Chlordane (technical Mixture & metabolites) DDT & metabolites Endosulfan & metabolites Endrin & metabolites Heptachlor metabolites Hexachlorocyclohexane Polychlorinated biphenyls (PCBs) Toxaphene Mirex <sup>b</sup> Methoxychlor <sup>b</sup>	<b>EPA 200.8 (Inf/Eff)</b> <b>SW 6020 (Sludge)</b> Antimony Arsenic Beryllium Cadmium Chromium Copper Lead Molybdenum Nickel Silver Selenium Thallium Zinc  <b>EPA 245.1 (Inf/Eff)</b> <b>SW7471A/B (Sludge)</b> Mercury  <b>SM 4500-CN-E (Inf/Eff)</b> <b>EPA 9010B Mod (Sludge)</b> Cyanide	
	<b>SW 8280A (Inf/Eff/Sludge)</b> 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD)		

Inf Influent

Eff Effluent

<sup>a</sup> "EPA" refers to the EPA document *Methods for Chemical Analysis of Water and Wastes*, revised March 1983, Document No. EPA-600/4-79-020 or 40 CFR 136; "SW" refers to the EPA Manual SW 846, *Test Methods for Evaluating Solid Waste*. 3rd Ed., 1986.

<sup>b</sup> Included with expanded method analyte list.

**Table 4. Preservation and Analytical Procedures for Influent, Effluent, and Sludge.**

<b>Parameter</b>	<b>Sample Type</b>	<b>Preservation</b>	<b>Maximum Holding Time</b>	<b>Method<sup>a</sup></b>
Temperature	Inf/Eff	None required	Analyze immediately	SM 2550B
pH	Inf/Eff	None required	Analyze immediately	SM 4500-H <sup>+</sup> B
BOD <sub>5</sub>	Inf/Eff	Cool, ≤6°C	48 hours	SM 5210B
Total Residual Chlorine	Eff	Fill completely	Analyze immediately	Hach 8167
DO Electrode	Eff	None required	Analyze immediately	SM 4500-O G
Suspended Solids	Inf/Eff	Cool, ≤6°C	7 days	EPA 160.2
Total Solids	Sludge	Cool, ≤6°C	7 days	SM 2540G
Enterococci	Inf/Eff	Cool, ≤6°C, Na <sub>2</sub> S <sub>2</sub> O <sub>3</sub> in effluent	24 hours	SM 9230C
Asbestos	Inf/Eff	Cool, ≤6°C, dark	Filter within 48 hours of receipt at lab	EPA 100.1/100.2
	Sludge	Cool, ≤6°C	28 days	Polarized Light Microscopy (PLM)
Fecal Coliform Bacteria	Eff	Cool, ≤6°C 0.0008% Na <sub>2</sub> S <sub>2</sub> O <sub>3</sub>	6 hours	SM 9221E
Total Ammonia as N	Eff	Cool, ≤6°C, H <sub>2</sub> SO <sub>4</sub> to pH <2	28 days	Hach 8038
Total Hydrocarbons as Oil and Grease	Inf/Eff	Cool, ≤6°C, dark HCl to pH<2	28 days	EPA 1664 HEM <sup>b</sup>
Volatile Organics	Inf/Eff	Cool, ≤6°C, dark, HCL to pH<2 L- Ascorbic Acid in effluent	14 days	EPA 624
	Sludge	Cool, ≤6°C	14 days	SW 8260B
Dioxins	Inf/Eff	Cool, ≤6°C	30 days until extraction/45 days after extraction	SW 8280A
	Sludge	Cool, ≤6°C	30 days until extraction/45 days after extraction	SW 8280A
Semi-Volatile Organics	Inf/Eff	Cool, ≤6°C, dark L- Ascorbic Acid in effluent	7 days until extraction/40 days after extraction	EPA 625
	Sludge	Cool, ≤6°C	14 days until extraction/40 days after extraction	SW 8270C
Pesticides & PCBs	Inf/Eff	Cool, ≤6°C, L- Ascorbic Acid in effluent	7 days until extraction/40 days after extraction	EPA 614 and EPA 608
	Sludge	Cool, ≤6°C	14 days until extraction/40 days after extraction	SW 8141A/8081A SW 8082



Initial WET testing was performed as a screening period over the course of three quarters during each of which three toxicity tests were performed; each with one vertebrate and two invertebrate species. These screening tests were performed during the third and fourth quarters of 2000 and the first quarter of 2001. Screening included the vertebrate *Atherinops affinis* (topsmelt) for survival and growth; an invertebrate bivalve species (either *Mytilus* spp. [mussel; survival and growth] or *Crassostrea gigas* [oyster; larval development]; and an invertebrate echinoderm species fertilization test (*Strongylocentrotus purpuratus* [purple urchin] or *Dendraster excentricus* [sand dollar]). Once the initial screening period was completed, the single most sensitive species (bivalve) was used for subsequent toxicity testing until re-screening was performed again. As required by the permit, re-screening must be performed each year during one quarter (different than the previous year) to determine the most sensitive species to use for continued testing. Re-screening was performed in the second quarter of 2002 and the third quarter of 2003, with bivalves again found to be the most sensitive species. Re-screening that was performed from 2004 through 2010 found the purple sea urchin to be the most sensitive species.

Accelerated testing requirements will be triggered if chronic toxicity is greater than 143 TUC (chronic toxicity units,  $TUC=100/\text{No Observed Effect Concentration [NOEC]}$ ). Accelerated testing will include the implementation of the initial investigation Toxicity Reduction Evaluation (TRE) workplan along with at least one additional toxicity test. If the investigation indicates the source of toxicity (e.g., a plant upset), and no toxicity greater than 143 TUC is observed in this additional test, the normal schedule of testing will be re-instated. If toxicity greater than 143 TUC is observed, then accelerated testing will continue with six more tests performed on a biweekly basis over a 12-week period. Testing will commence within two weeks of receipt of the sample results of the exceedance. If no toxicity greater than 143 TUC is observed in these tests, then the normal schedule of testing will be re-instated. If toxicity greater than 143 TUC is observed in any of the six tests, then a TRE will be initiated within 15 days of receipt of the sample results of the exceedance. A Toxicity Identification Evaluation (TIE) may also be initiated as part of the overall TRE process, and if this is initiated during the accelerated testing period, the accelerated testing may be terminated or used as necessary in performing the TIE.

Toxicity testing was performed as described in Short-Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Marine and Estuarine Organisms (EPA, 1988) and the 'West Coast Marine Methods Manual', First Edition (EPA, 1995) as required by the permit. The presence of chronic toxicity was estimated as described by these references. Quality assurance for the toxicity testing included the testing of a series of five dilutions and a control, including the concentration of the effluent at the edge of the ZID (0.70 %) as well as two dilutions above and two dilutions below 0.70 %. Reference toxicants were tested concurrently with the effluent testing, using the same procedures. If the effluent tests did not meet all the acceptability criteria as specified in the referenced methods, then the effluent was re-sampled and re-tested as soon as possible. Control and dilution water was natural or synthetic seawater as called for by the referenced methods. If the dilution water was different from the culture water, a second control using culture water will be run.

As part of the WET testing, an initial investigation TRE plan was prepared and submitted to EPA under separate cover (Kinnetic Laboratories, Inc., 2000b). This plan describes the events that will occur should chronic toxicity be detected. As required by the permit and the manual Toxicity Reduction Evaluation Guidance for Municipal Wastewater Treatment Plants (EPA, 1999b), a preliminary TRE will be initiated within 15 days of the receipt of sample results of the

permit exceedance. A more detailed TRE workplan will subsequently be developed to more fully investigate and identify the cause of the toxicity, identify and provide a schedule of the actions that AWWU will use to mitigate the impact of the discharge, and to prevent the recurrence of the toxicity. As noted above, the TIE may be initiated as part of the overall TRE process during the accelerated testing schedule.

### **2.1.5 Part 503 Sludge Monitoring**

Operations at the Asplund WPCF include a sludge incinerator that is subject to regulation under 40 CFR Part 503 - Standards for the Use or Disposal of Sewage Sludge. The current NPDES permit requires sludge monitoring twice per year, once during summer dry conditions and once during summer wet conditions as noted earlier. There are no Part 503 monitoring requirements included in the reissued NPDES permit because EPA Region 10's current policy is to remove these requirements from NPDES permits with the intention of writing "sludge only" permits in the future. However, the Part 503 regulations are "self-implementing" in that the facility is required to meet the monitoring requirements in the regulation whether they are specifically included in a permit or not. Therefore, monitoring at the Asplund WPCF includes Part 503 monitoring of sludge. Monitoring frequencies required by 40 CFR Part 503 for incineration are once per 60 days for arsenic, cadmium, chromium, lead, and nickel. Frequency required for mercury is at least once per year. Frequency for beryllium is not specified. AWWU has chosen to also test for mercury and beryllium once per 60 days, more frequently than required, so as to be consistent with the testing frequency for the other metals. Allowable limits are site-specific and were re-calculated per Part 503 regulation in May 2008 by CH2M Hill based on 2007 data (CH2M Hill, 2008). While methods for this monitoring component have been described elsewhere (AWWU, 2000) and results of the monitoring have been provided under separate reporting requirements to EPA, the data are also included in this report.

## **2.2 RECEIVING WATER QUALITY MONITORING**

### **2.2.1 Water Quality Sampling**

As required by the permit, water quality must be monitored annually during the summer in dry weather conditions (Table 1). Sampling was performed at non-fixed stations during consecutive ebb and flood tides at the outfall station and a single flood tide at the control station. Station locations were determined by following the track of

- |  |
|--|
| <ul style="list-style-type: none"><li>✓ determine compliance with the NPDES permit and State of Alaska water quality criteria</li><li>✓ aid in assessing the water quality at the discharge point</li><li>✓ determine compliance with the regulatory criteria of Section 301(h) for the CWA</li><li>✓ determine the level of bacterial contamination in nearshore waters</li><li>✓ provide data for evaluation of permit re-issuance</li></ul> |
|--|

drogues released above the diffuser at the outfall station and at the control station located north across Knik Arm from Point Woronzof, directly off Point MacKenzie in a similar water depth as the outfall. Three drogue tracks on each tide were performed at each location. Four stations were sampled on each drogue track released at the outfall as follows:

- above the diffuser
- as close to the ZID boundary as practicable
- at least one nearfield station along the drogues path near the ZID
- in the shallow subtidal area before the drogue grounds or along the drogues path at a farfield location.



As noted in the permit, the ZID is defined as the water column above the area delineated by the sector of a circle with the center located over the outfall, 30 meters (m) shoreward of the diffuser, 650 m in radius, and with a 220° angle (Figure 3).

The plume location was determined by following a holey-sock current drogue (Figure 4). The drogue consisted of a six-foot cylindrical nylon tube ballasted at the bottom with a two-pound weight and lead line and attached at the top with a bridle to a spherical float. This float was attached to the tracking spar via a connecting line. These cylindrical or spherical designs that enclose a parcel of water have been found to more accurately follow the ambient current patterns than other drogue designs such as the window shade design (Sombardier and Niiler, 1994).

Sampling was performed by positioning the vessel over the diffuser (or control station) for the first sampling station of the drogue track. The drogue was released at approximately the same time and followed until navigation information indicated that the ZID had been reached, at approximately 650 m from the outfall, at which time the ZID boundary station was sampled. The third and fourth stations along each drogue track were sampled as the drogue traveled through the channel in Knik Arm or as it slowed in shallow water prior to grounding. Navigation was accomplished using a differential global positioning system (DGPS). If DGPS coordinates were unavailable, standard GPS coordinates were recorded and noted on the appropriate log.

Samples were collected for the parameters outlined in Table 5. The surface waters of all stations were sampled for fecal coliform, color, turbidity, and TRC. Surface samples were collected by grabbing directly into the appropriate sample bottles at sample depth (15 - 30 centimeters [cm]). Mid- and bottom depth turbidity samples were collected at all stations using Niskin<sup>®</sup> bottles. Mid- and bottom depths were determined at each station using the survey vessel's fathometer. Samples were collected as simultaneously as possible at all three target depths. Hydrographic profiles of temperature, salinity, DO, and pH were collected at all stations using a Seabird SEACAT<sup>®</sup> SBE-19 CTD (conductivity, temperature, and depth) recorder. This instrument was also equipped with a DO, pH, and optical backscatter (turbidity) sensors to allow profiles of these parameters to be recorded. Samples for the analysis of total and dissolved metals, TSS, polycyclic aromatic hydrocarbons (PAH), and total aromatic hydrocarbons (TAH) were collected from surface waters at the first three stations (diffuser, ZID boundary, and channel) at low tide along the first flood drogue track at both the outfall and control stations. These samples were collected as grab samples directly into the appropriate sample containers. A single replicate sample for each parameter or a single profile was collected at each station except for quality control samples, which are described in Section 4.2.

Samples were analyzed following the methods provided in Table 6. Samples were appropriately labeled at time of collection using pre-prepared, project-specific sample labels as described in Section 2.5 and prepared for shipment to the laboratory. Preservation and maximum holding time information for each of these methods is also provided in Table 6. All sample containers were immediately placed on gel ice after sampling. Samples remained chilled as required during shipment to the analytical laboratory.

### **2.2.2 Intertidal Bacterial Sampling**

As part of the receiving water quality monitoring effort, intertidal sampling for fecal coliform bacteria was also performed at eight stations shown in Table 7 and depicted in Figure 3. Two

**Figure 3. Asplund WPCF Outfall, ZID, and Locations of Intertidal Bacteriological Sampling.**

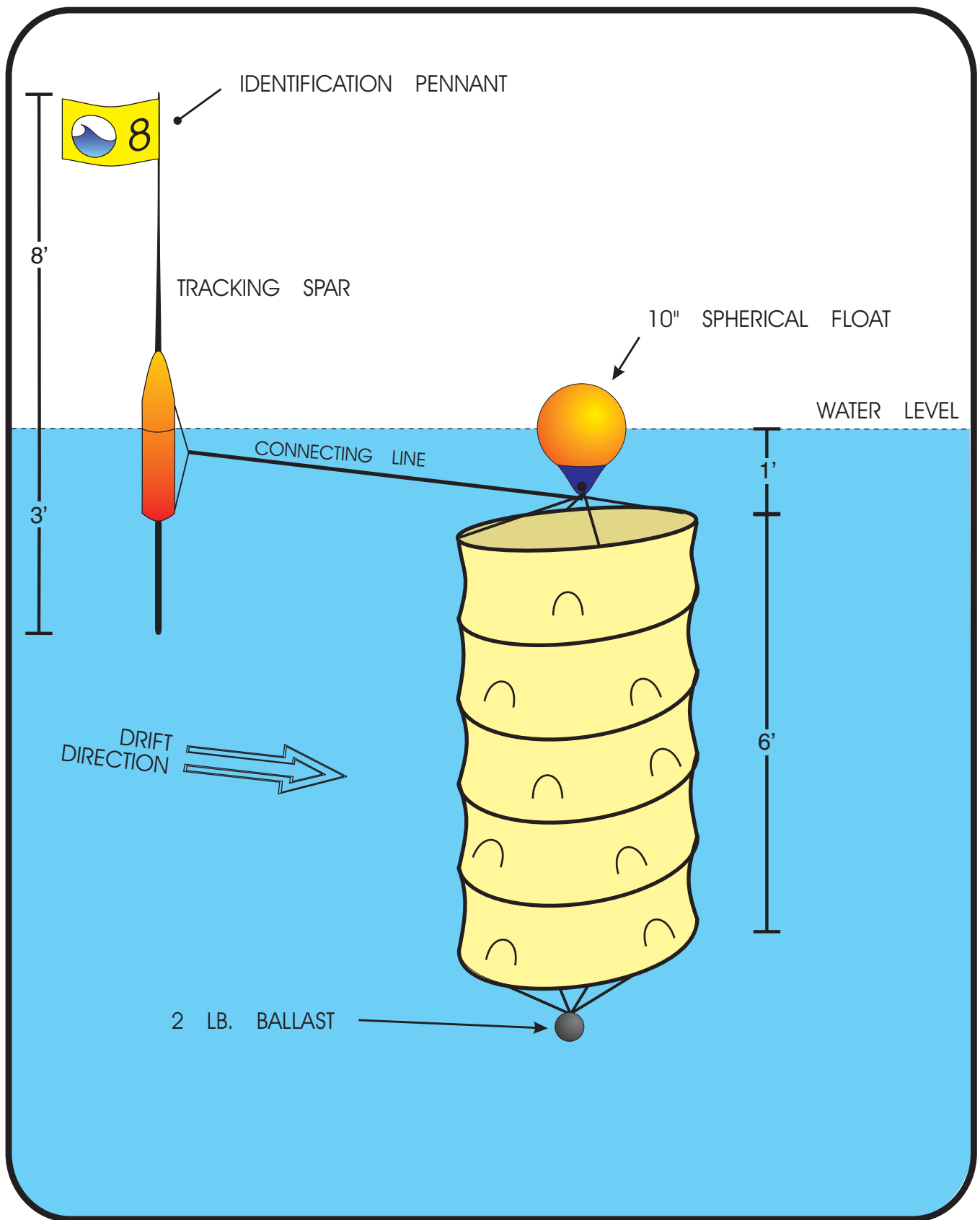


Figure 4. Holey-Sock Drogue, Flotation, and Marker Buoy.

**Table 5. Receiving Water Quality Monitoring Requirements.**

Parameter	Sampling Depth		
	Surface (above 0.5 m)	Surface, Mid-, and Bottom	Profile (1- to 3-m intervals)
Fecal Coliform	all stations <sup>a</sup> , within the 15-30 cm layer		
Color	all stations, within the 15-30 cm layer		
Total Residual Chlorine (TRC)			
Field Observations: presence or absence of floating solids, visible foam (other than trace), oil wastes, and/or sheen	all stations where surface samples are collected		
Total Aqueous Hydrocarbons (TAqH)	first three stations along the first flood drogue track at both the outfall and control locations		
Total Aromatic Hydrocarbons (TAH)			
Metals and Cyanide <sup>b</sup>			
Total Suspended Solids (TSS)			
Turbidity		all stations	
pH			all stations
Temperature			
Dissolved Oxygen (DO)			
Salinity			

<sup>a</sup> Non-fixed stations were sampled following the track of drogues released at the diffuser (outfall station) or at a fixed station having the same depth due north across Knik Arm from Point Woronzof near Point MacKenzie (control station). Three drogue tracks were made during each of a consecutive flood and ebb tide at the outfall station. Stations included the following along each outfall drogue track: above the diffuser; as close to the ZID boundary as practicable; one near-field station in the channel of Knik Arm; and a far-field station along the drogue path or in the shallow subtidal area before the drogue grounds. Three drogue tracks were also made during a flood tide at the control station in conjunction with or as soon as practicable as the sampling at the outfall station.

<sup>b</sup> Metals include arsenic, cadmium, chromium, copper, lead, mercury, nickel, silver, and zinc; these were analyzed and reported as both total recoverable and dissolved metals.

**Table 6. Methods, Preservation, and Maximum Holding Times for the Analysis of Receiving Water Quality Samples.**

Parameter	Method <sup>a</sup>	Preservation	Maximum Holding Time
Fecal Coliform	SM 9221E	Cool, 6°C, dark, (0.0008% Na <sub>2</sub> S <sub>2</sub> O <sub>3</sub> in presence of chlorine)	8 hours (6 hours max transport, 2 hours once received by lab)
Color	SM 2120B	Cool, 6°C, dark	48 hours
Total Residual Chlorine (TRC)	SM 4500-Cl G	None	Analyze immediately
Turbidity	SM 2130B	Cool, 6°C, dark	48 hours
Total Aqueous Hydrocarbons (TAqH)	EPA 602 plus xylenes	Cool, 6°C, HCl to pH<2, L- Ascorbic Acid in presence of chlorine	14 days
	EPA 610	Cool, 6°C, dark, L- Ascorbic Acid in presence of chlorine	7 days until extraction/ 40 days after extraction
Total Aromatic Hydrocarbons (TAH)	EPA 602	Cool, 6°C, HCl to pH<2 L- Ascorbic Acid in presence of chlorine	14 days
Metals (Total Recoverable and Dissolved)	See note <sup>b</sup>	Cool, 6°C, HNO <sub>3</sub> to pH <2 (after filtration for dissolved)	90 days – Hg 180 days – all others
Cyanide	EPA 335.2	NaOH, 6°C	14 days
Total Suspended Solids (TSS)	EPA 160.2	Cool, 6°C	7 days
Dissolved Oxygen (DO)	SM 4500-O G (electrode)	None	<i>in situ</i>
pH	SM 4500-H <sup>+</sup> B	None	<i>in situ</i>
Temperature	SM 2550B <sup>c</sup>	None	<i>in situ</i>
Salinity	SM 2520B <sup>c</sup>	None	<i>in situ</i>

<sup>a</sup> "EPA" refers to the EPA document *Methods for Chemical Analysis of Water and Wastes*, revised March 1983, Document No. EPA-600/4-79-020, or 40 CFR 136. "SM" refers to *Standard Methods for the Examination of Water and Wastewater*, 20th ed., 1998.

<sup>b</sup> Dissolved metals were filtered before acidification; total recoverable metals were digested by ASTM Method D4309-91. Cadmium, chromium, copper, nickel, lead, silver and zinc were subject to pre-concentration by chelation following EPA Method 1640 prior to analysis by inductively coupled plasma mass spectroscopy. These metals, along with antimony, beryllium, selenium, and thallium, were analyzed as total recoverable and dissolved metals as appropriate for ICP/MS (EPA Method 1638). Mercury was analyzed using cold vapor atomic fluorescence following EPA Method 1631. Arsenic was determined by flame ionization atomic spectroscopy (SW846 Method 7062).

<sup>c</sup> Modified for *in situ* measurements collected with the CTD.

**Table 7. Approximate Locations of Intertidal Bacteria Sampling Stations.**

Station	Station Location Relative to Diffuser	Latitude (N)	Longitude (W)
IT-1	2000 m east	61° 12' 10"	149° 58' 55"
IT-2	1200 m east	61° 12' 11"	149° 59' 50"
IT-3	750 m east	61° 12' 15"	150° 00' 20"
IT-4	250 m east	61° 12' 19"	150° 00' 52"
IT-5	250 m southwest	61° 12' 15"	150° 01' 10"
IT-6	750 m southwest	61° 12' 02"	150° 01' 28"
IT-7	2000 m southwest	61° 11' 22"	150° 02' 02"
IT-C	Across Knik Arm from the diffuser	61° 14' 26"	150° 01' 09"

replicate water samples were collected from each station near high slack water when the water depths were between 1 to 3 feet (ft). Additional quality control samples were collected as described in Section 4.2. Surface samples were collected by grabbing from 15 - 30 cm depths directly into the appropriate container. Samples were analyzed using the same procedures described previously and in Table 6.

In addition to the required intertidal samples, two replicated fecal coliform samples were also collected once during the water quality monitoring effort from three area streams that empty into Knik Arm: Ship, Chester, and Fish Creeks. Samples were analyzed using the same procedures described previously and in Table 6.

At time of collection, all fecal coliform samples were appropriately labeled using pre-prepared, project-specific sample labels as described in Section 2.5. All samples were collected in the appropriate pre-cleaned sample containers, dechlorinated when necessary, and preserved as described by the method. Samples were placed on gel ice immediately after sampling and remained chilled during transport to the laboratory. Field notes, including navigational and sampling information, were recorded on project-specific field logs. As required by the permit, field observations taken at each station included the presence or absence of floating solids, visible foam in other than trace amounts, oily wastes, or sheen. Weather observations were also recorded. All field documentation was reviewed by the field leader at the completion of the survey for accuracy and completeness. Sample collection and shipment was documented using project-specific chain of custody forms as described in Section 2.5.

### **2.2.3 Vessel Support**

The *NORTH FORTY*, a 26-ft KLI-owned survey vessel, was used for drogue tracking and water sampling in 2010. In addition, a 15-ft Zodiac<sup>®</sup> was used to retrieve grounded drogues and conduct intertidal bacteria sampling. The Zodiac<sup>®</sup> was also used to transport samples with short holding times (i.e., bacterial and turbidity samples) ashore during the sampling effort.

## **2.3 SEDIMENT AND BIOACCUMULATION MONITORING**

As stipulated in the NPDES permit, sediment and bioaccumulation monitoring was to be performed during the fourth year after the effective date of the permit. Accordingly, the intertidal and subtidal sediment sampling was performed in conjunction with the 2003 receiving water monitoring sampling and the bioaccumulation sampling was performed during 2004.

## **2.4 LABORATORY ANALYSIS**

Laboratory analyses of all samples for this program followed preservation and analysis procedures described by EPA-accepted protocols as referenced in this document (Table 4 and Table 6). These procedures are fully described by the referenced documents and/or 40 CFR 136.

## **2.5 DOCUMENTATION PROCEDURES**

All field and sampling data were recorded on appropriate pre-printed project-specific field data forms. Field data forms included drogue tracking forms, water sampling log forms, sample identification/chain of custody forms, and sample labels. These forms were tailored to the monitoring program to facilitate accurate and complete documentation of field activities. The field task leader was responsible for review and approval of all field documentation; this was completed as soon as possible after sampling.

Sampling logs included specific information such as station identification, sample identification numbers, navigational data, sampling or photographic observations, sampling depths, and collection date and time. Drogue tracking logs included station identification information along with navigational data to allow the track of each drogue to be later determined and plotted. Pre-printed labels included such information as station designation, analysis type, date of collection, sampling personnel, and a pre-assigned sample identification number to uniquely identify each sample. Field duplicate and field blank quality control (QC) samples were labeled as were regular environmental samples so as to be blind to the laboratory analysts.

Sample identification and integrity was ensured by a rigidly-enforced chain of custody program. Sample identification/chain of custody (COC) forms provided specific information concerning the identification, handling, and shipment of samples.

Pertinent information from the sample label was transferred onto the COC, along with other information as required. COC forms were completed, signed by field personnel, and copied if needed. In some cases, where photocopying was not convenient or possible, two-part carbonless forms were used. The original of each COC form was packed with the samples in coolers for shipment to the laboratory. The field task leader retained a copy of each form for the field records and for tracking purposes should a shipment become lost or delayed. Upon receipt of the samples at the analytical laboratory, the laboratory sample custodian signed the samples in by checking all sample labels against the COC information and noting any discrepancies as well as sample condition (e.g., sample temperature, containers leaking or damaged during shipment). Internal sample tracking procedures at the laboratory were initiated immediately upon receipt of samples as described by each laboratory's standard operating procedures.





### 3.0 RESULTS

#### 3.1 INFLUENT, EFFLUENT, AND SLUDGE MONITORING

##### 3.1.1 Monthly Discharge Monitoring Data

Results of AWWU's daily, weekly, and monthly sampling of influent and effluent for non-metals are presented as monthly summaries in Table 8. Averages are based on the 12-month period from January through December 2010.

The removal of BOD<sub>5</sub> as determined by subtracting the effluent from the influent divided by the influent  $[(\text{Inf}-\text{Eff})/\text{Inf} \times 100]$  averaged 48 %, and removal of TSS averaged 79 % for the 12-month reporting period. On a monthly average basis, the removal of BOD<sub>5</sub> ranged from 43 to 53%. These averages exceed the minimum values required by the amendments to the CWA (40 CFR Part 125.60; Final Rule, 8/9/94), whereby dischargers with 301(h) variances are required to remove 30 % of BOD<sub>5</sub> and 30 % of the suspended solids on a monthly basis. The highest monthly average effluent BOD<sub>5</sub> was 159 mg/L, substantially less than the permit limitation of 240 mg/L. All of the BOD<sub>5</sub> values (daily, weekly, and monthly averages) reported for the calendar year 2010 met the permit limitations. Total suspended solids concentrations in the effluent were low and typical of those seen historically at the Asplund WPCF with the highest monthly average effluent concentration of 61 mg/L and average of 53 mg/L compared to the permit limit of 170 mg/L. Weekly average and daily maximum TSS also met permit requirements for all sampling events in 2010. On a monthly average basis, the removal of TSS ranged from 77 to 81 % with all values within the CWA requirements of 30% removal.

The highest mean monthly fecal coliform bacteria count was 49 FC MPN/100 mL seen in September 2010. All of the months in 2010 met the permit limitation of 850 FC MPN/100 mL based on a geometric mean of at least five samples, which ranged from 7 to 49 FC MPN/100 mL, well below the permit limitation. The criterion of not more than 10 % of the samples analyzed should exceed 2,600 FC MPN/100 mL was also met for every month in 2010. Better plant performance was seen in 2010 in terms of effective chlorination resulting in lower fecal coliform bacteria concentrations than in some prior years.

The TRC daily maximum concentration did not exceed the permit-required limitation of 1.2 mg/L for the entire year, with a maximum daily value of 0.88 mg/L and a monthly maximum daily range of 0.35 to 0.88 mg/L. The monthly averages of TRC concentrations ranged from 0.22 to 0.43 mg/L, with an overall annual average of 0.29 mg/L. The permit requirement that effluent pH remain between 6.5 and 8.5 standard units was always met, exhibiting a minimum and maximum range of 6.5 to 7.7 pH units for the year.

Although other parameters such as DO, temperature, and ammonia do not have permit limitations, ranges were typical of those seen historically. DO in the effluent exhibited monthly averages ranging from 2.3 to 3.6 mg/L, with a yearly average of 3.0 mg/L which was similar to that seen in 2008 and 2009 where the yearly averages were 2.9 and 2.8 mg/L, respectively. Temperature showed yearly averages of 12.8 and 13.0 °C in the influent and effluent, respectively. Monthly values for total ammonia in the effluent ranged from 14.6 to 26.7 mg/L, with a yearly average of 22.9 mg/L, similar to that seen historically. Average Plant flow for the year was 26.8 million gallons per day (mgd) which is very similar to the average flow rate seen over the past five years.

 **Table 8. Discharge Monitoring Data for Influent and Effluent Non-Metals.**

Month	Average EFF Flow Rate (MGD)	Temperature Average (°C)		pH Minimum/ Maximum (pH) <sup>a</sup>		TRC Average (mg/L)		DO Average (mg/L)		BOD <sub>5</sub> Average (mg/L)			Total Susp. Solids Average (mg/L)			Fecal Coliform Average (MPN/100 mL)		Total Ammonia Average (mg/L)	
		INF	EFF	INF	EFF	INF	EFF	INF	EFF	INF	EFF	REM (%)	INF	EFF	REM (%)	INF	EFF	INF	EFF
01/10	25.6	10.9	11.0	7.0/7.9	7.0/7.3	NT	0.33	NT	2.9	244	132	46	228	51	78	NT	10	NT	23.7
02/10	25.4	10.2	10.9	7.1/8.0	6.9/7.6	NT	0.33	NT	3.4	237	126	47	229	52	77	NT	20	NT	22.2
03/10	25.9	10.3	10.7	6.8/8.0	6.8/7.4	NT	0.29	NT	3.4	253	132	48	243	50	79	NT	38	NT	14.6
04/10	28.5	10.0	10.6	6.8/7.9	6.8/7.5	NT	0.43	NT	3.3	216	109	50	218	47	79	NT	12	NT	21.6
05/10	27.6	11.6	12.0	7.2/8.0	6.8/7.4	NT	0.34	NT	3.0	235	135	43	232	53	77	NT	11	NT	22.1
06/10	27.5	13.6	13.9	6.6/7.8	6.5/7.7	NT	0.33	NT	3.6	279	159	43	271	55	80	NT	7	NT	24.2
07/10	29.2	14.8	15.0	6.7/7.3	6.6/7.1	NT	0.22	NT	2.3	278	157	44	275	56	80	NT	11	NT	26.4
08/10	30.2	15.0	15.2	7.0/7.6	6.7/7.1	NT	0.24	NT	3.0	280	132	53	289	61	79	NT	25	NT	24.4
09/10	27.9	15.2	15.4	7.1/7.6	7.0/7.4	NT	0.24	NT	2.9	250	123	51	261	53	80	NT	49	NT	18.1
10/10	25.1	15.0	15.0	6.9/7.8	6.8/7.6	NT	0.27	NT	3.0	278	140	50	285	57	80	NT	8	NT	25.5
11/10	24.7	13.9	13.9	7.0/7.9	7.0/7.7	NT	0.23	NT	2.5	245	125	49	253	47	81	NT	10	NT	26.7
12/10	24.0	12.9	12.9	7.2/7.9	6.9/7.7	NT	0.24	NT	2.9	243	129	47	242	56	77	NT	9	NT	24.8
<b>Average</b>	26.8	12.8	13.0	6.6/8.0	6.5/7.7	---	0.29	---	3.0	253	133	48	252	53	79	---	18	---	22.9

<sup>a</sup> Monthly or Yearly (minimum-maximum)

NT Not tested (tested in effluent only)

REM Percent Removal

--- Not applicable

### 3.1.2 Toxic Pollutants and Pesticides Analyses

Toxic pollutant and pesticide monitoring for influent, effluent, and sludge was conducted on 13 - 14 July 2010 for summer-dry weather and 16 - 17 August 2010 for the summer-wet sampling. Sampling was performed over a 24-hr period by AWWU personnel.

Results of the toxic pollutant and pesticide analyses are provided in Table 9 (July 2010) and Table 10 (August 2010). For semi-volatile organic compounds (EPA Methods 625/8270C), volatile organic compounds (EPA Methods 624/8260B), and pesticides (EPA Methods 608/8081A/8082 and 614/8141A), only those pollutants that were detected in the influent, effluent, or sludge are listed. Refer to Appendices A and B for the laboratory reports and a complete listing of pollutants analyzed. Pollutants found in the influent were usually detected in the effluent and were also often present in the sludge, and vice versa. In general, pollutant concentrations were very low and many of the concentrations reported for the two samplings were so low that they had to be estimated and below method reporting limits (MRLs).

Percent removal values shown in these tables were computed from influent and effluent concentrations. Percent removal was only calculated for compounds where a concentration in the influent and/or effluent was reported at a level above the MRL. Compounds with estimated concentrations (denoted with a "J" qualifier) were not used for percent removal calculations unless a detected concentration above MRL was reported for that compound in the other type of sample (influent or effluent). The reported MRL was used for calculations where necessary (where a compound was reported as non-detect [ND]). Where laboratory duplicate analyses were performed for a parameter, a percent removal is provided for each duplicate. For summed values, such as BETX, the MRL was used for values reported as ND.

Calculation of percent removal for some contaminants may not truly be representative of treatment plant efficiency due to several factors that influence the removal values. Most notable is the fact that influent and effluent autosamplers do not produce parallel samples over the same required 24-hour time period due to the approximately 6-hour hydraulic residence time of the wastewater flowing through the treatment process prior to being discharged as final effluent. Also affecting the calculation of percent removal is the addition of more than 1 million gallons of fresh water from the city's drinking water supply and/or on-site well water to the treatment process after the influent autosampler. Additionally, incinerator scrubber water, filtrate from the belt filter presses, scum concentrator return water, and in-plant wash down water are added back into the treatment process which only impacts the effluent composite sample. Often the percent removal calculation is performed on data derived at or below the MRL. In some cases where a contaminant is not detected above the MRL, any comparison of that estimated value to a value that was detected above the MRL is statistically insignificant despite the requirement to report it. As such, calculation of negative removals is possible in spite of all evidence to support an efficient and effective treatment process.

The types and concentrations of measured volatile and semi-volatile organic compounds varied somewhat between the two sampling periods. Compounds that were detected in either the influent or effluent during at least one of the sampling events included: acetone, chloroform, chloromethane, 1,4-dichlorobenzene, methylene chloride, toluene, total xylenes, bis(2-ethylhexyl) phthalate, diethyl phthalate, 4-methylphenol, phenol, and 3&4-methylphenol (p&m cresol). Many of these compounds were estimated values that were only detected below their reporting limits and therefore were qualified with a J but were generally seen in both the influent

**Table 9. Toxic Pollutants and Pesticides in the Influent, Effluent, and Sludge, Sampled 13 and 14 July 2010.**

<b>Pollutant</b>	<b>Influent<sup>a</sup> (µg/L)</b>	<b>Effluent<sup>a</sup> (µg/L)</b>	<b>Sludge<sup>a</sup> (µg/g)</b>	<b>Percent Removal</b>
<b>VOLATILE ORGANICS– detected substances only</b>				
Acetone	97 B	130 B	7.210	34
Benzene	ND (1.8)	ND (1.8)	ND (0.0783)	---
Chloroform	2.0 J	3.5 J	ND (0.0783)	---
Chloromethane	ND (0.89)	0.95 J	ND (0.392)	---
Ethylbenzene	ND (2.2)	ND (2.2)	ND (0.0783)	---
Toluene	10	9.8	0.932	2
Methylene Chloride <sup>f</sup>	4.8 J,B	6.0 B	ND (0.392)	-25
1,4- Dichlorobenzene	0.91 J	0.73 J	0.133	---
Xylenes (total)	7.8 J	4.7 J	0.282	---
<b>SEMI-VOLATILE ORGANICS – detected substances only</b>				
Bis(2-ethylhexyl) phthalate	ND (54)	14	31 D	74
Butyl Benzyl Phthalate	ND (54)	ND (11)	6.6 D	---
Fluoranthene	ND (54)	ND (11)	2.8 D	---
4- Methylphenol	180 D	99	95 D	45
Phenol	180 D	ND (11)	ND (6.2)	94
Pyrene	ND (54)	ND (11)	2.3 D	---
<b>HYDROCARBONS</b>				
Oil & Grease (EPA 1664-HEM)	47000	35500	NT	24
Total Aromatic Hydrocarbons as BETX from EPA Method 624	21.8	18.5	1.37	15

**Table 9. Toxic Pollutants and Pesticides in the Influent, Effluent, and Sludge, Sampled 13 and 14 July 2010. (continued)**

<b>Pollutant</b>	<b>Influent<sup>a</sup> (µg/L)</b>	<b>Effluent<sup>a</sup> (µg/L)</b>	<b>Sludge<sup>a</sup> (µg/g)</b>	<b>Percent Removal</b>
<b>DISSOLVED METALS</b>				
Antimony	ND(1.0)	ND(1.0)	NT	---
Arsenic	ND(5.0)	ND(5.0)	NT	---
Beryllium	ND(0.40)	ND(0.40)	NT	---
Cadmium	ND(0.50)	ND(0.50)	NT	---
Chromium	ND(2.0)	ND(2.0)	NT	---
Copper	24.8	48.0	NT	-94
Lead	0.760	0.964	NT	-27
Mercury	0.243	0.302	NT	-24
Molybdenum	ND(10.0)	ND(10.0)	NT	---
Nickel	3.31	3.73	NT	-13
Selenium	ND(5.0)	ND(5.0)	NT	---
Silver	ND(1.0)	ND(1.0)	NT	---
Thallium	ND(1.0)	ND(1.0)	NT	---
Zinc	33.3	75.3	NT	-126
<b>TOTAL METALS</b>				
Antimony	ND(1.0)	ND(1.0)	1.68	---
Arsenic	ND(5.0)	ND(5.0)	5.53	---
Beryllium	ND(0.40)	ND(0.40)	ND(0.314)	---
Cadmium	3.67	0.660	1.26	82
Chromium	3.09	ND(2.0)	13.6	35
Copper	61.7	82.9	207	-34
Lead	4.02	2.38	19.3	41
Mercury	ND(0.20)	ND(0.20)	NT	---
Molybdenum	ND(10.0)	ND(10.0)	5.91	---
Nickel	5.12	3.62	12.0	29
Selenium	ND(5.0)	ND(5.0)	2.95	---
Silver	1.38	ND(1.0)	6.78	28
Thallium	ND(1.0)	ND(1.0)	ND(0.0627)	---
Zinc	175	118	610	33

**Table 9. Toxic Pollutants and Pesticides in the Influent, Effluent, and Sludge, Sampled 13 and 14 July 2010. (continued)**

<b>Pollutant</b>	<b>Influent<sup>a</sup> (µg/L)</b>	<b>Effluent<sup>a</sup> (µg/L)</b>	<b>Sludge<sup>a</sup> (µg/g)</b>	<b>Percent Removal</b>
<b>PESTICIDES – detected substances only</b>				
2,4'-DDD	ND(0.053)	0.012	0.027 D	77
2,4'-DDE	ND(0.053)	ND(0.011)	0.047 D	---
4,4'-DDT	ND(0.092)	0.086 P	180 P,D	7
<b>ENTEROCOCCI BACTERIA</b>				
Enterococci <sup>b</sup>	NT	1011.2/4352	NT	---
<b>OTHER COMPONENTS</b>				
Asbestos <sup>c</sup>	82.20	14.80	ND	82
Cyanide	ND(5.0)	ND(5.0)	ND(0.67)	---
Dioxin (2,3,7,8-TCDD)	ND(0.00103)	ND(0.00103)	ND(0.0010314)	---

*a* Detection limits or reporting limits are included in parentheses for non-detected (ND) values

*b* Enterococci reported in CFU/100 ml; two replicates- (sample and duplicate).

*c* Asbestos reported in million fibers/L (influent and effluent) and present or none detected (sludge).

P More than 40% RPD between primary and confirmation column results. The higher of the two results is reported.

J Estimated value (below MRL but above MDL)

--- Not applicable (not calculated)

ND None detected

NT Not tested

D Reported result from a dilution.

B Compound found in lab blank and sample.

**Table 10. Toxic Pollutants and Pesticides in the Influent, Effluent, and Sludge, Sampled 16 and 17 August 2010. (continued)**

<b>Pollutant</b>	<b>Influent<sup>a</sup> (µg/L)</b>	<b>Effluent<sup>a</sup> (µg/L)</b>	<b>Sludge<sup>a</sup> (µg/g)</b>	<b>Percent Removal</b>
<b>VOLATILE ORGANICS– detected substances only</b>				
Acetone <sup>f</sup>	220	180	0.068	18
Benzene	ND (1.8)	ND (1.8)	0.00047	---
Chloroform <sup>f</sup>	2.0 J	3.0 J	0.00096 J	-50
1,4-Dichlorobenzene	0.78 J	0.67 J	0.017	-14
Ethylbenzene	ND (2.2)	ND (2.2)	0.0055	---
Methylene chloride <sup>f</sup>	15 B	7.2 B	0.0065 B	-100
Tetrachloroethene	ND (2.7)	ND (2.7)	0.0052	---
Toluene	5.3	6.3	0.130	-19
1,2,3-Trichlorobenzene	NT	NT	0.0062 B	---
1,2,4-Trichlorobenzene	NT	NT	0.0072 B	---
Trichlorofluoromethane	NT	NT	0.0013 J	---
Xylenes (total)	8.6 J	5.7 J	0.030	---
<b>SEMI-VOLATILE ORGANICS– detected substances only</b>				
Diethyl phthalate	ND (20.0)	7.04	ND (21.9)	65
3&4-Methylphenol (p&m Cresol)	221 RL3	84.0	141 RL3	62
Phenol	336 RL3	18.0	ND (21.9)	95
<b>HYDROCARBONS</b>				
Oil & Grease (EPA 1664-HEM)	98100	49100	NT	50
Total Aromatic Hydrocarbons as BETX from EPA Method 624	17.9	16.0	0.166	11

**Table 10. Toxic Pollutants and Pesticides in the Influent, Effluent, and Sludge, Sampled 16 and 17 August 2010.**

<b>Pollutant</b>	<b>Influent<sup>a</sup> (µg/L)</b>	<b>Effluent<sup>a</sup> (µg/L)</b>	<b>Sludge<sup>a</sup> (µg/g)</b>	<b>Percent Removal</b>
<b>DISSOLVED METALS</b>				
Antimony	ND (1.0)	ND (1.0)	NT	---
Arsenic	ND (5.0)	ND (5.0)	NT	---
Beryllium	ND (0.40)	ND (0.40)	NT	---
Cadmium	ND (0.50)	ND (0.50)	NT	---
Chromium	ND (2.0)	ND (2.0)	NT	---
Copper	10.3	13.3	NT	-29
Lead	0.315	0.356	NT	-13
Mercury	ND (0.20)	ND (0.20)	NT	---
Molybdenum	ND (10.0)	ND (10.0)	NT	---
Nickel	3.78	3.85	NT	-2
Selenium	ND (5.0)	ND (5.0)	NT	---
Silver	ND (1.0)	ND (1.0)	NT	---
Thallium	ND (1.0)	ND (1.0)	NT	---
Zinc	22.9	28.0	NT	-22
<b>TOTAL METALS</b>				
Antimony	ND (1.0)	ND (1.0)	1.54	---
Arsenic	ND (5.0)	ND (5.0)	4.73	---
Beryllium	ND (0.40)	ND (0.40)	ND (0.329)	---
Cadmium	ND (0.50)	ND (0.50)	1.72	---
Chromium	4.16	ND (2.0)	15.1	----
Copper	67.7	41.9	273	38
Lead	3.19	1.81	18.6	43
Mercury	ND (0.20)	ND (0.20)	0.833	---
Molybdenum	ND (10.0)	ND (10.0)	5.27	---
Nickel	5.95	3.96	14.5	33
Selenium	ND (5.0)	ND (5.0)	ND (1.64)	---
Silver	1.20	ND (1.0)	4.02	---
Thallium	ND (1.0)	ND (1.0)	ND (0.0657)	---
Zinc	178	94.9	653	47



**Table 10. Toxic Pollutants and Pesticides in the Influent, Effluent, and Sludge, Sampled 16 and 17 August 2010.**

Pollutant	Influent <sup>a</sup> (µg/L)	Effluent <sup>a,b</sup> (µg/L)	Sludge <sup>a</sup> (µg/g)	Percent Removal
<b>PESTICIDES – detected substances only</b>				
4,4'-DDT	0.074 P,D	0.094 P,D	0.270	-27
<b>ENTEROCOCCI BACTERIA</b>				
Enterococci <sup>c</sup>	NT	>2419.6/>2419.6	NT	---
<b>OTHER COMPONENTS</b>				
Asbestos <sup>d</sup>	ND (<4.11)	ND (<37.00)	ND	---
Cyanide	ND (5.0)	5.0	ND(0.68)	---
Dioxin (2,3,7,8-TCDD)	ND(0.000000941)	ND(0.000000704)	ND(0.000000147)	---

*a* Detection or reporting limits are included where possible in parentheses for non-detected (ND) values

*b* Duplicate field sample analysis or duplicate laboratory analysis provided (value/duplicate value)

*c* Enterococci reported in CFU/100 mL

*d* Asbestos reported in million fibers/L (influent and effluent) and present or none detected (sludge)

*e* Result from August 503 metals sampling.

*f* Also present in trip blank.

--- Not applicable (not calculated)

D Reported result from a dilution.

J Estimated value (below MRL but above MDL)

B Method blank contamination.

P More than 40 % RPD between primary and confirmation columns results. [The higher of the two results is reported.]

ND None detected

NT Not tested or lab did not match analyte lists and cannot retrieve data.

RL3 Reporting limit raised, high levels non-target analytes.

and effluent samples. Trace levels of acetone were seen in the laboratory method blank in the July sampling and methylene chloride was also seen in both the trip blank and laboratory method blanks for both the July and August sampling efforts at concentrations similar to that seen in the influent and effluent. Chloroform was also seen in the August trip blank. Organic compounds that were seen in the sludge during the July 2010 sampling effort included acetone, toluene, 1,4-dichlorobenzene, xylenes, bis(2-ethylhexyl) phthalate, butyl benzyl phthalate, fluoranthene, 4-methylphenol, and pyrene. In August 2010, of those organic compounds that were detected in either the influent or effluent, acetone, chloroform, 1,4-dichlorobenzene, methylene chloride, toluene, 3&4-methylphenol, and total xylenes were also seen in the sludge. Several of the sludge concentrations were estimated and qualified with a J as they fell below MRLs.

Oil and grease concentrations measured in the influent and effluent in 2010 using EPA 1664A HEM were similar to those seen in 2009 although slightly elevated to those seen over the prior four years that ranged from 14.9 to 32.0 mg/L with effluent concentrations of 35.5 and 49.1 mg/L during the July and August sampling, respectively. Oil and grease effluent values from 2005 to 2008 ranged from 15-32 mg/L. Effluent BETX values as measured by EPA 624 were 18.5 and 16.0 µg/L in the July and August 2010 samplings, respectively. Refer to Sections 5.1 and 5.2 for further discussion of the significance of the total hydrocarbon values.

The AWQS have site-specific criteria for Knik Arm of Cook Inlet and the Point Woronzof area and also include State wide criteria that are based on dissolved metals. These AWQS were utilized to determine the MAEC (defined as the value specified as the receiving water limit and/or the permit limit multiplied by the initial dilution of 142:1 for conservative substances and 180:1 for non-conservative substances (TRC, ammonia, cyanide, TAH, and TAqH) after taking into account the natural background concentration). Both total and dissolved concentrations of metals in the effluent were then compared to the MAECs.

Total recoverable metals concentrations in both the influent and effluent were found to be low. Influent and effluent concentrations of total antimony, arsenic, beryllium, molybdenum, mercury, selenium, and thallium were all below detection limits during both sampling periods. Other total recoverable metals such as, chromium, copper, lead, nickel, silver, and zinc were seen in the influent or effluent during both sampling events and cadmium during one sampling event, but at very low levels when compared to their respective MAECs.

Dissolved metals concentrations were also found to be low. Dissolved antimony, arsenic, beryllium, cadmium, chromium, molybdenum, selenium, silver, and thallium were all below MRLs for both influent and effluent for both sampling periods. Dissolved copper, lead, nickel and zinc were above detection limits in either influent or effluent during both of the sampling events and mercury during the July sampling event. The concentration for dissolved copper in effluent was found to be the highest of any of the metals with respect to its MAEC of 317 µg/L, with a level of 48 µg/L during the July sampling. While dissolved copper was found to be the highest metal detected in the effluent with respect to its MAEC, it was still nearly an order of magnitude less than the MAEC.

Few pesticides were detected in the influent, effluent, or sludge during the July or August 2010 sampling events. During the July sampling, 2,4'-DDD and 4,4'-DDT was detected in the effluent and sludge and 2,4'-DDE in the sludge only. During the August sampling, 4,4'-DDT was detected in the influent, effluent, and sludge samples. The source of these pesticides is unknown since DDTs have not been manufactured, used, or sold in the U.S. for over 20 years. No PCBs

were detected in either the July or August sampling events. For a complete list of the various chlorinated organic and pesticide analytes, refer to Appendices A1 and B1.

The permit calls for the analysis of enterococci bacteria in effluent twice per year in conjunction with the summer dry and summer wet sampling. The enterococci in the effluent was reported as 1,011.2 and 4,352 most probable number (MPN)/100 mL in the two samples analyzed for the July 2010 sampling and >2,419.6 MPN/100 mL for two replicate samples taken during the August 2010 sampling event.

Asbestos was detected at low levels in influent and effluent during the July event but was not detected in the influent, effluent, or sludge during the August sampling event. The concentration of cyanide in the influent and effluent was below the detection limit of 5.0 µg/L during July 2010 sampling event. Cyanide was seen in the effluent during the August sampling event at 5.0 µg/L. The effluent cyanide concentration was still well below the MAEC of 181 µg/L. Dioxins (2,3,7,8-TCDD) were tested in influent, effluent, and sludge during both sampling events, but none were detected.

### **3.1.3 Pretreatment Monitoring Data**

As part of the NPDES permit, AWWU is to conduct pretreatment monitoring twice per year in conjunction with the toxic pollutant and pesticide analyses. This monitoring includes three consecutive days of 24-hr composite sampling of the influent and effluent and one day of sludge sampling. Pretreatment analyses include cyanide and a suite of metals that are analyzed as both total and dissolved. Results of the pretreatment monitoring are presented in Table 11.

Collection of samples for trace metals analysis, as part of the toxic pollutant and pesticide sampling events in July and August 2010, coincided with the first day of the pretreatment monitoring for the Asplund WPCF during 2010, as discussed in Section 3.1.2. Individual metals concentrations for the 3-day pretreatment sampling event were generally found to be very similar with little variation between sampling days, particularly for the effluent.

Of all the metals in the effluent, total copper and zinc concentrations were the highest. However, concentrations of these metals were still well below their respective MAECs. For example, dissolved copper in the effluent was reported at concentrations of 32.6 - 48.0 µg/L during the three days of pretreatment sampling in July 2010 and a range of 12.2 - 13.3 µg/L during the August sampling effort as compared to the MAEC of 317 µg/L. Total copper in the effluent was found to range from 41.9 to 82.9 µg/L for the six pretreatment samples compared to the MAEC of 317 µg/L. Influent values were generally more variable than those seen in effluent, as would be expected. Dissolved zinc in the effluent ranged from 28 - 99.3 µg/L during both pretreatment samplings, while total zinc ranged from 94.9 - 130 µg/L during these samplings as compared to an MAEC of 11,249 µg/L. Dissolved and total mercury were below reporting limits of 0.2 µg/L in all six effluent pretreatment samples, as compared to the MAEC of 2.73 µg/L. Other metals were also found to be substantially less than their respective MAECs. Total cyanide concentrations in the effluent ranged from <5 - 7.7 µg/L as compared to a MAEC of 181 µg/L.

### **3.1.4 Whole Effluent Toxicity Testing Results**

Quarterly WET testing was conducted on 24-hr flow composite effluent samples as required under the permit during all four quarters of calendar year 2010. Echinoderm fertilization tests

**Table 11. Pretreatment Monitoring Data for Influent and Effluent Metals and Cyanide.**  
Concentrations are in µg/L.

Parameter	July 2010						August 2010					
	Influent			Effluent			Influent			Effluent		
Sample Date	14	15	16	14	15	16	17	18	19	17	18	19
<b>Dissolved Metals</b>												
Antimony	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00
Arsenic	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00
Beryllium*	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400
Cadmium	<0.500	<0.500	<0.500	<0.500	<0.500	<0.500	<0.500	<0.500	<0.500	<0.500	<0.500	<0.500
Chromium	<2.00	<2.00	<2.00	<2.00	<2.00	<2.00	<2.00	<2.00	<2.00	<2.00	<2.00	<2.00
Copper	24.8	26.4	23.5	48.0	32.6	46.7	10.3	4.68	10.5	13.3	12.7	12.2
Lead	0.760	0.915	1.79	0.964	0.993	1.93	0.315	<0.200	0.291	0.356	0.280	0.225
Mercury	0.243	<0.200	<0.200	0.302	<0.200	<0.200	<0.200	<0.200	<0.200	<0.200	<0.200	<0.200
Molybdenum	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0
Nickel	3.31	3.73	3.57	3.73	3.89	3.73	3.78	4.15	3.65	3.85	3.82	4.23
Selenium	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00
Silver	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00
Thallium	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00
Zinc	33.3	28.4	34.3	75.3	70.9	85.3	22.9	16.6	14.1	28.0	33.4	99.3
<b>Total Metals and Cyanide</b>												
Antimony	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00
Arsenic	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00
Beryllium*	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400	<0.400
Cadmium	3.67	<0.500	<0.500	0.660	<0.500	<0.500	<0.500	0.836	0.931	<0.500	<0.500	<0.500
Chromium	3.09	3.00	4.85	<2.00	<2.00	<2.00	4.16	3.79	4.74	<2.00	<2.00	<2.00
Copper	61.7	58.0	69.5	82.9	49.2	61.4	67.7	83.6	64.7	41.9	48.4	45.0
Cyanide	<5.0	<5.0	<5.0	<5.0	<5.0	<5.0	<5.0	<5.0	<5.0	5.0	6.8	7.7
Lead	4.02	3.71	8.02	2.38	1.96	3.79	3.19	5.04	4.00	1.81	2.02	1.89
Mercury	<0.200	<0.200	<0.200	<0.200	<0.200	<0.200	<0.200	<0.200	<0.200	<0.200	<0.200	<0.200
Molybdenum	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0	<10.0
Nickel	5.12	6.02	7.20	3.62	4.47	4.29	5.95	6.37	6.10	3.96	4.22	4.40
Selenium	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00	<5.00
Silver	1.38	<1.00	2.19	<1.00	<1.00	<1.00	1.20	1.55	1.37	<1.00	<1.00	<1.00
Thallium	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00	<1.00
Zinc	175	159	183	118	101	130	178	220	1520	94.9	106	103

\* Not required by permit for "Pretreatment" monitoring  
Values reported as "<" (less than), are reporting limits

were performed using the purple sea urchin, *Strongylocentrotus purpuratus*, during the second, third, and fourth quarters of 2010. Use of this test during the second, third, and fourth quarters was based on the screening test results from the third quarter of 2009 and first quarter of 2010 which had determined the sea urchin to be the most sensitive species tested (see Section 2.1.4). Annual re-screening for the most sensitive species in 2010 was performed during the first quarter. Based on interpretation of this and previous years detailed laboratory results, the laboratory recommended continuing with the sea urchin as the most sensitive species for 2011 until the annual 3-species rescreening is performed again during 2011.

Results of all the tests performed in 2010 are summarized below and presented in Table 12 as the lowest observed effect concentration (LOEC), no observed effect concentration (NOEC), and in chronic toxicity units (TUc), where  $TUc = 100/NOEC$ . Detailed results in the form of descriptive laboratory reports that present all data in tabular form along with statistical analyses, QA/QC information, and reference toxicant test results have previously been submitted to ADEC and EPA with Asplund WPCF's monthly discharge monitoring reports (DMRs) and are not duplicated here in this report.

**Table 12. Summary of WET Test Data from 2010.**

Toxicity Test	LOEC (%)	NOEC (%)	TUc
<b>1<sup>st</sup> Quarter 2010</b>			
Bivalve (survival)	> 2.8	2.8	≤ 35.7
Bivalve (development)	> 2.8	2.8	≤ 35.7
Topsmelt (survival)	> 2.8	2.8	≤ 35.7
Topsmelt (growth)	> 2.8	2.8	≤ 35.7
Echinoderm (fertilization)	2.8	1.4	71.4
<b>2<sup>nd</sup> Quarter 2010</b>			
Echinoderm (fertilization)	1.4	0.7	142.8
<b>3<sup>rd</sup> Quarter 2010</b>			
Echinoderm (fertilization)	1.4	0.7	142.8
<b>4<sup>th</sup> Quarter 2010</b>			
Echinoderm (fertilization)	> 2.8	2.8	≤ 35.7

The three-species WET testing was performed during the first quarter of 2010. The test included: the bivalve larvae, *Mytilus galloprovincialis*, survival and development; topsmelt, *Atherinops affinis*, survival and growth; and echinoderm, *Strongylocentrotus purpuratus* fertilization. The first quarter sampling event was conducted between 24 and 29 January, 2010.

Results of the bivalve test showed that no concentration of effluent that was tested produced any toxicity to the test organisms. The NOECs for both survival and development were 2.8 % effluent and both LOECs were > 2.8 % effluent. Chronic toxicity units were ≤ 35.7 TUc for both survival and development in the bivalves. Both reference toxicant tests were within laboratory control chart limits and indicated typical sensitivity of the test population.

The topsmelt bioassay showed no significant decrease in survival or growth at any concentration compared with seawater-only controls. The LOECs for survival and growth were  $> 2.8 \%$ , and the NOECs were both  $2.8 \%$ . The effluent therefore showed  $\leq 35.7$  TUC in both the survival and growth endpoints. The concurrent reference toxicant test results were within laboratory control chart limits and indicated typical sensitivity of the test population. All test acceptability criteria (TAC) were met in both the effluent bioassay and the reference toxicant bioassay.

Results of the echinoderm fertilization test showed that effluent that was tested produced a toxic response to the test organisms at the highest test concentration. The LOEC for fertilization was therefore  $2.8 \%$  and the NOEC was  $1.4 \%$ , with a TUC of 71.4, meeting the allowable permit limit of not exceeding 143 TUC. Based on the results of the three-species testing and past years' results, it was recommended to continue to use the urchin as the most sensitive species for the toxicity testing to be conducted in the subsequent quarters of 2010 and until the three-species comparison is repeated in 2011.

Second quarter 2010 echinoderm fertilization testing was performed on a single 24-hr composite sample collected on 20-21 May 2010. Results from this sample showed no significant decrease in egg fertilization at any effluent sample concentration with an LOEC of  $1.4 \%$  and a NOEC of  $0.7 \%$  effluent. The TUC was 142.8, which was within the permit maximum allowable limit of 143 TUC. All TAC were met in both the effluent bioassay and the reference toxicant bioassay.

The WET testing for the third quarter with echinoderms was performed on a sample collected 18-19 July 2010. Results of the echinoderm fertilization test conducted during the third quarter showed a significant decrease in egg fertilization occurred at the  $2.8 \%$  and  $1.4 \%$  effluent concentrations when compared to the seawater-only controls. The LOEC for fertilization was therefore  $1.4 \%$  and the NOEC was  $0.7 \%$ , with a TUC of 142.8, meeting the allowable permit limit and the bioassay met all TAC.

The WET testing for the fourth quarter with echinoderms was performed on samples collected 25-26 October, 2010. Results of the echinoderm fertilization test conducted during the fourth quarter showed that no significant decrease in egg fertilization occurred at any effluent concentration when compared to the seawater-only controls. The LOEC for fertilization was therefore  $> 2.8 \%$  and the NOEC was  $2.8 \%$ , with a TUC of  $\leq 35.7$ , meeting the allowable permit limit. All TAC were met in both the effluent bioassay and the reference toxicant bioassay.

### **3.1.5 Part 503 Sludge Monitoring Data**

The AWWU operates a sludge incinerator at the Asplund WPCF for which the NPDES permit requires sludge monitoring twice per year as part of the "Toxic Pollutants and Pesticides/Pretreatment" sampling requirements. As described in Section 2.1.5, AWWU performed Part 503 sludge monitoring with a minimum frequency of once every 60 day period (6 times/year). These data will be submitted along with other incinerator operational information to EPA by 19 February 2011. This submittal will take the form of a separate report; however, for completeness and for comparison purposes, this information has been included here as well.

Results of the sludge monitoring for metals for the year are presented in Table 13. Metals concentrations were extremely low compared to allowable limits and were usually very similar to those seen historically. In 2009, levels of chromium and zinc were found to be high during the

**Table 13. Part 503 Discharge Monitoring Data for Sludge Metals.** Concentrations are in mg/kg dry weight. All metals are reported as total metals.

Parameter <sup>1,6</sup>	Arsenic	Beryllium <sup>3,5</sup>	Cadmium	Chromium	Lead	Nickel	Mercury <sup>4,5</sup>
Site Specific Limit 2010 <sup>2</sup>	99	2014	88	1937	5045	6727	129
02/10/10	2.23	ND	1.01	7.94	14.20	7.52	2.140
03/31/10	3.38	ND	2.53	16.20	19.60	12.00	0.403
05/20/10	9.21	ND	1.35	11.00	24.80	8.81	1.130
07/13/10	5.53	ND (0.314)	1.26	13.60	19.3	12.0	NT <sup>7</sup>
08/16/10	4.73	ND (0.329)	1.72	15.10	18.60	14.5	0.833
09/09/10	5.13	ND	1.77	14.80	24.60	13.90	0.451
12/14/10	3.86	ND	1.03	9.05	12.60	7.86	0.834
MINIMUM	2.23	ND (0.314)	1.01	7.94	12.60	7.52	0.403
MAXIMUM	9.21	ND(0.329)	2.53	16.20	24.80	14.5	2.140
AVERAGE <sup>8</sup>	4.87	ND	1.52	12.53	19.10	10.94	0.96

1) *Test Methods for Evaluating Solid Waste, Physical/Chemical Methods, SW-846, 3rd Edition*, EPA. Samples for total metals analysis, with the exception of mercury, were prepared (acid digestion) according to EPA method 3050, SW-846.

2) Site-specific sludge limits calculated by CH2MHill May 2008, based primarily on October 2007 Asplund Incinerator Source Test.

3) Beryllium emissions shall not exceed 10 grams per day. With a control efficiency of 0.9998 at the maximum sludge feed rate, a sludge concentration of 500 milligrams per dry kilogram of sludge will not result in a violation of the limit.

4) Mercury emissions shall not exceed 3,200 grams per day. With a control efficiency of 0.0 at the maximum sludge feed rate, a sludge concentration of 9.63 milligrams per dry kilogram of sludge will not result in a violation of the limit.

5) Monitoring frequencies required by 40 CFR Part 503 for incineration are once per 60 days for arsenic, cadmium, chromium, lead, and nickel. Mercury is at least once per year. Frequency for beryllium is not specified. AWWU has chosen to test mercury and beryllium more frequently than required to be consistent with the other metals.

6) Samples were analyzed by SGS - EPA 6020, Mercury by EPA 7471A and/or 7471B methods. Parentheses contain PQL values.

7) NT = not tested.

8) Average computed using detection limit where concentration is ND (non-detect).

June sampling event, but this was not seen during 2010. The only metal that had historically been elevated for some sampling events was arsenic. In 2010, the highest concentration of arsenic in the sludge was 9.21 mg/kg compared to the allowable limit of 99 mg/kg. As mentioned above, no actual limits exist in the current NPDES permit. Allowable limits are site-specific and were calculated by the permittee per Part 503 regulations (CH2M Hill 2008). EPA plans to issue sludge only permits in the future; in the interim, 40 CFR Part 503 regulations are “self-implementing”.

### **3.2 RECEIVING WATER QUALITY MONITORING RESULTS**

Water quality sampling of the receiving water was conducted on 13 - 14 July 2010, concurrent with the summer dry sampling. Sampling results are contained in the following subsections.

#### **3.2.1 Plume Dispersion Sampling**

##### **Drogue Tracking Results**

Drogues were released on 13 July 2010 at the ZID station for the ebb and flood tidal cycles and on 14 July 2010 at the control station for the flood tidal cycle. Three drogues were deployed during each tidal cycle and four stations sampled along each drogue track.

##### **ZID Site**

The Point Woronzof ebb drogue drop and tracking cycles were performed during the morning and early afternoon of 13 July 2010. The predicted tidal range during ebb stage was 36.2 feet (Figure 5 and Table 14; NOAA Tides and Currents, 2010). A composite of the ebb drogue deployments is depicted in Figure 6 and are very similar to that seen in previous years.

The ebb drogues traveled from approximately 3.7 to 4.3 nautical miles, all three traveling in a west southwesterly direction. No eddies were observed during these drogue tracks, nor did any of the drogues become grounded during their tracks. The first ebb (E1) drogue was released at 09:15 Alaska Daylight Time (ADT), about 25 minutes after high tide. This drogue tracked southwest but north of the shoal that was evident at low tide. The drogue traveled at an average speed of 148 centimeters per second (cm/s) over its entire track of approximately 4.3 nautical miles. The second ebb drogue (E2) was released at 11:13 ADT and also tracked north of the shoal, traveling west southwest with an average speed of 142 cm/s over the entire track, traveling approximately 3.7 nautical miles. The third drogue (E3) was released at 13:17 ADT four and a half hours after high slack. The third drogue followed a similar path to the E1 and E2 drogues traveling in a west southwesterly direction. This drogue traveled approximately 3.9 nautical miles at 113 cm/s over its entire track. The relationship of drogue tracks with respect to the tide are shown in Figure 5.

Flood drogue tracks are depicted in Figure 7. The tidal range during flood stage was 35.0 feet (Figure 5 and Table 14; NOAA, 2010). The first flood drogue (F1) was deployed on 13 July at 16:00 ADT at low slack water at the outfall. This drogue traveled easterly along the shoreline on the east side of Point Woronzof for approximately 1.1 nautical mile at an average speed of 38 cm/s before looping back towards shore and entering the shallow intertidal area where it grounded and was retrieved. The fact that this drogue looped back along the shore is evidence of a small eddy in the lee of Point Woronzof. The second flood drogue (F2) was deployed at 17:48



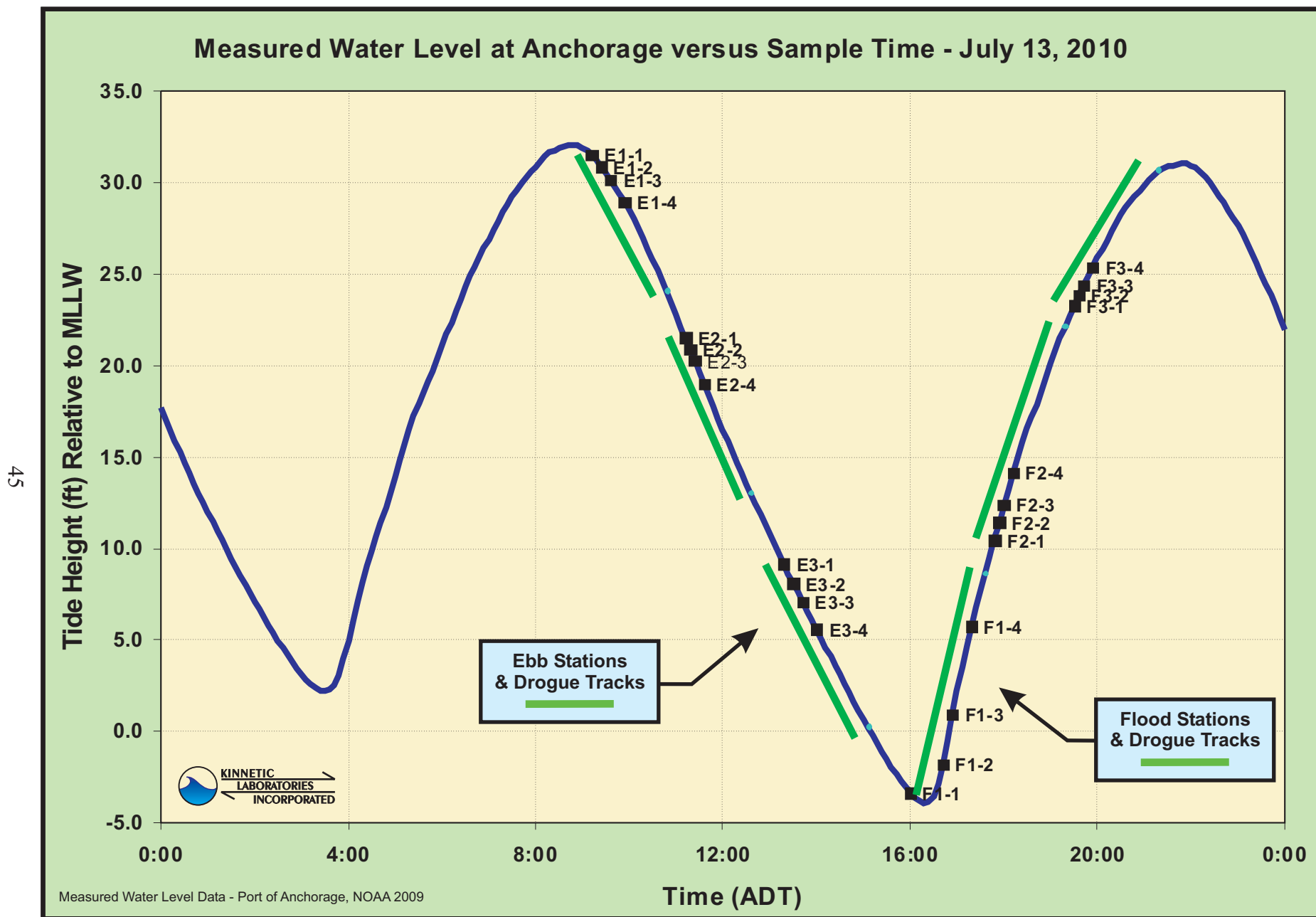


Figure 5. Tidal Information for Receiving Water Sampling, Ebb and Flood Tides.

**Table 14. 2010 Drogue Tracking Information.**

DATE	STATION	TIDAL INFORMATION				DROGUE NO.	RELEASE TIME AFTER SLACK (HOURS:MINUTES)	DROGUE SPEED AVERAGE (CM/S)
		Slack Water (Alaska Daylight Time <sup>a</sup> ; Stage)		Direction	Range (Feet) <sup>b</sup>			
13 July 2010	ZID	08:50	HIGH	EBB	36.2	E1	00:25	148
13 July 2010	ZID	08:50	HIGH	EBB	36.2	E2	02:23	142
13 July 2010	ZID	08:50	HIGH	EBB	36.2	E3	04:27	113
13 July 2010	ZID	16:02	LOW	FLOOD	35.0	F1	-00:02	38
13 July 2010	ZID	16:02	LOW	FLOOD	35.0	F2	01:46	68
13 July 2010	ZID	16:02	LOW	FLOOD	35.0	F3	03:28	118
14 July 2010	CONTROL	16:44	LOW	FLOOD	35.7	C1	00:44	193
14 July 2010	CONTROL	16:44	LOW	FLOOD	35.7	C2	02:06	156
14 July 2010	CONTROL	16:44	LOW	FLOOD	35.7	C3	03:50	216

<sup>a</sup> NOAA Tides and Currents 2010. (Port of Anchorage, Anchorage)

<sup>b</sup> Observed water level variations during tide.

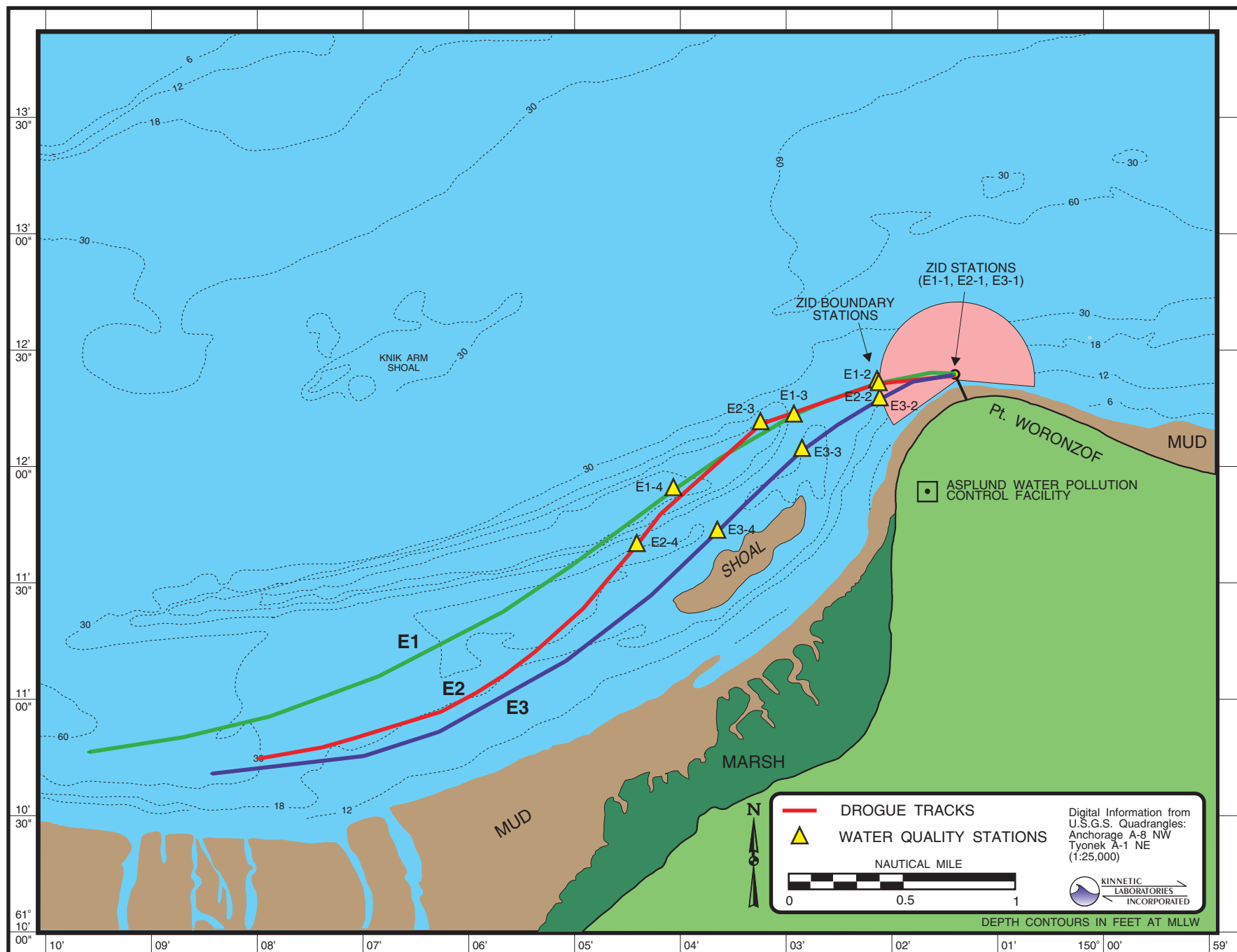


Figure 6. Summary of Ebb Drogue Tracks and Receiving Water Sampling Locations at Point Woronzof, 13 July 2010.

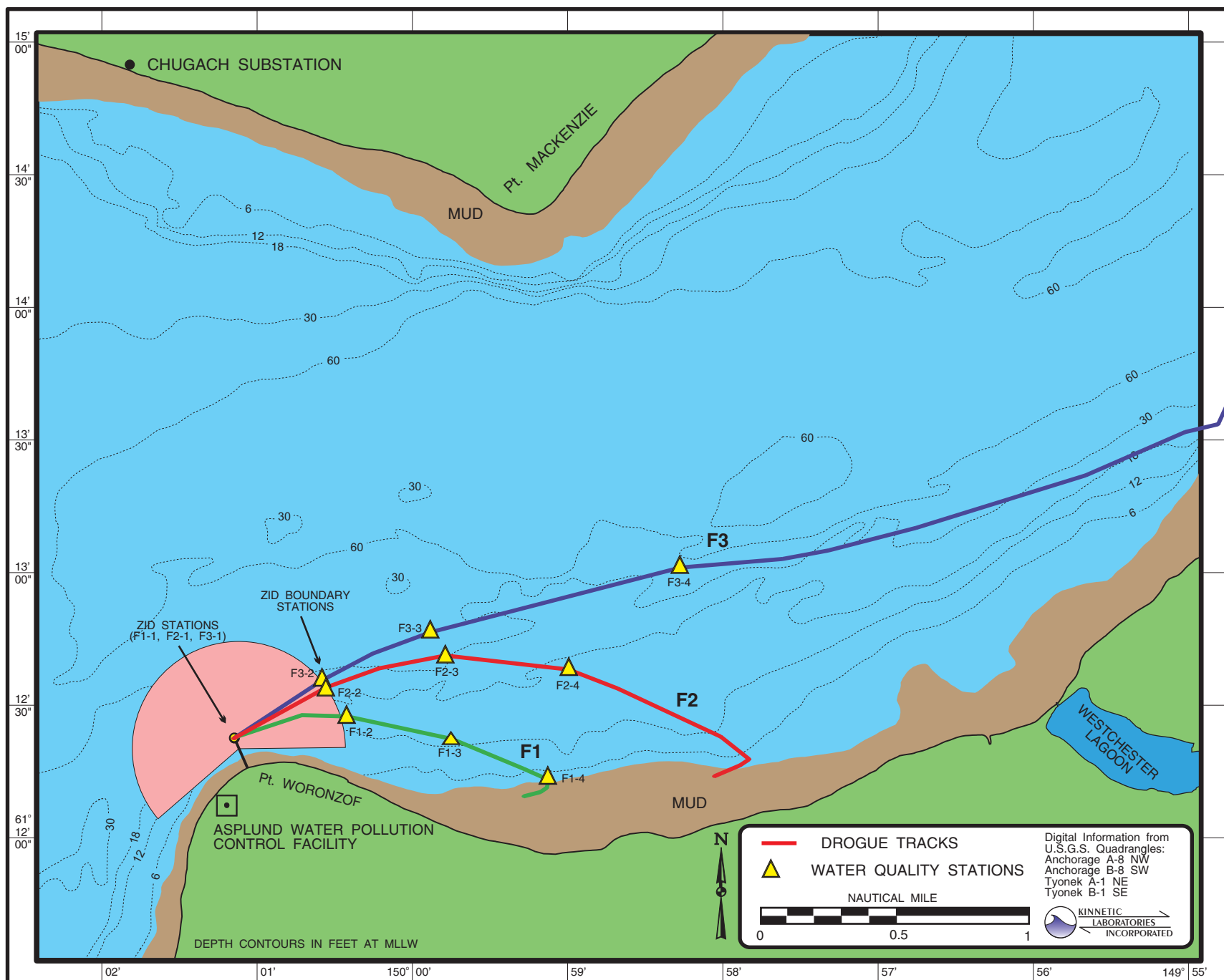


Figure 7. Summary of Flood Drogue Tracks and Receiving Water Sampling Locations at Point Woronzof, 13 July 2010.

ADT, approximately 1¼ hours after low slack. This drogue was transported initially to the northeast then southeasterly and further offshore than the first drogue and was tracked for about 1.9 nautical miles at an average speed of 68 cm/s before it looped back towards shore and was recovered. The third flood drogue (F3) was deployed at 19:30 ADT, almost 3½ hours after low slack water, and tracked for almost two hours. The third drogue traveled in a northeast direction as had the second drogue for more than a mile, then moved further out from the shoreline, where it continued moving northeast in the central Knik Arm Channel with an average speed of 118 cm/s. This drogue was tracked for approximately 4.1 nautical miles and was recovered 0.5 nautical miles offshore and to the west of the Port of Anchorage.

### **Control Site**

The Point MacKenzie control drogues were deployed and tracked on 14 July 2010. The predicted tidal range during the flood tide was 35.7 ft. Tidal information is provided in Figure 8 and Table 14 (NOAA, 2010). A composite of the three drogue trajectories at the control site is presented in Figure 9 which is very similar to prior years. The relationship of drogue tracks with respect to the tide and when sampling took place are shown in Figure 8.

The three control drogues had somewhat different tracks with the first drogue tracking closest to shore and the last drogue tracking furthest offshore in the center portion of the Knik Arm Channel. The first drogue was released at 17:28 ADT, 45 minutes after low tide, and traveled to the east and then to the northeast parallel to the shoreline. This drogue traveled 3.7 nautical miles with an average speed of 193 cm/s over the entire track before being retrieved north of Port MacKenzie 0.5 nautical miles from shore. The second drogue (C2) was released at 18:50 ADT, 2 hours and 6 minutes into the flood tidal cycle, and tracked for about one hour. This drogue had an average speed of 156 cm/s over the entire track and moved towards the northeast in mid-channel before encountering a counterclockwise eddy that brought it back towards shore near Port MacKenzie. The second drogue eventually became entangled with the support pilings at Port MacKenzie where it was recovered after traveling 3.8 nautical miles. The third control drogue (C3) was released at 20:34 ADT, approximately 4 hours after low slack water. The drogue traveled in a manner similar to the first and second drogues and slightly further offshore, moving northeast into the central channel with an average speed of 216 cm/s, traveling 3.9 nautical miles in all.

### **Summary of Receiving Water Quality Data**

The summer water quality sampling for all analysis types was conducted concurrently with the drogue tracking studies on 13 - 14 July 2010. As discussed previously, three drogues were released at the ZID for both ebb and flood tides and three were released at the control site for the flood tide. Water samples and CTD measurements were obtained at four stations along each drogue's track prior to their being retrieved. In the current NPDES permit, the ZID boundary is located 650 m distance from the outfall diffuser. To accomplish the ZID site sampling station, the vessel was positioned directly upcurrent from the diffuser and allowed to drift down across it. Upon reaching the outfall diffuser, the drogue was dropped and the within-ZID station sampled. The distance from the outfall diffuser was monitored with the DGPS, and upon reaching 650 m distance from the diffuser, the ZID-boundary station was sampled. The third and fourth stations were then sampled along the drogue's path. Due to high current speeds, anchoring the vessel and sampling at each station was not practical or desirable for this type of sampling due to the large wire angles of sampling gear and quickness needed in sampling.

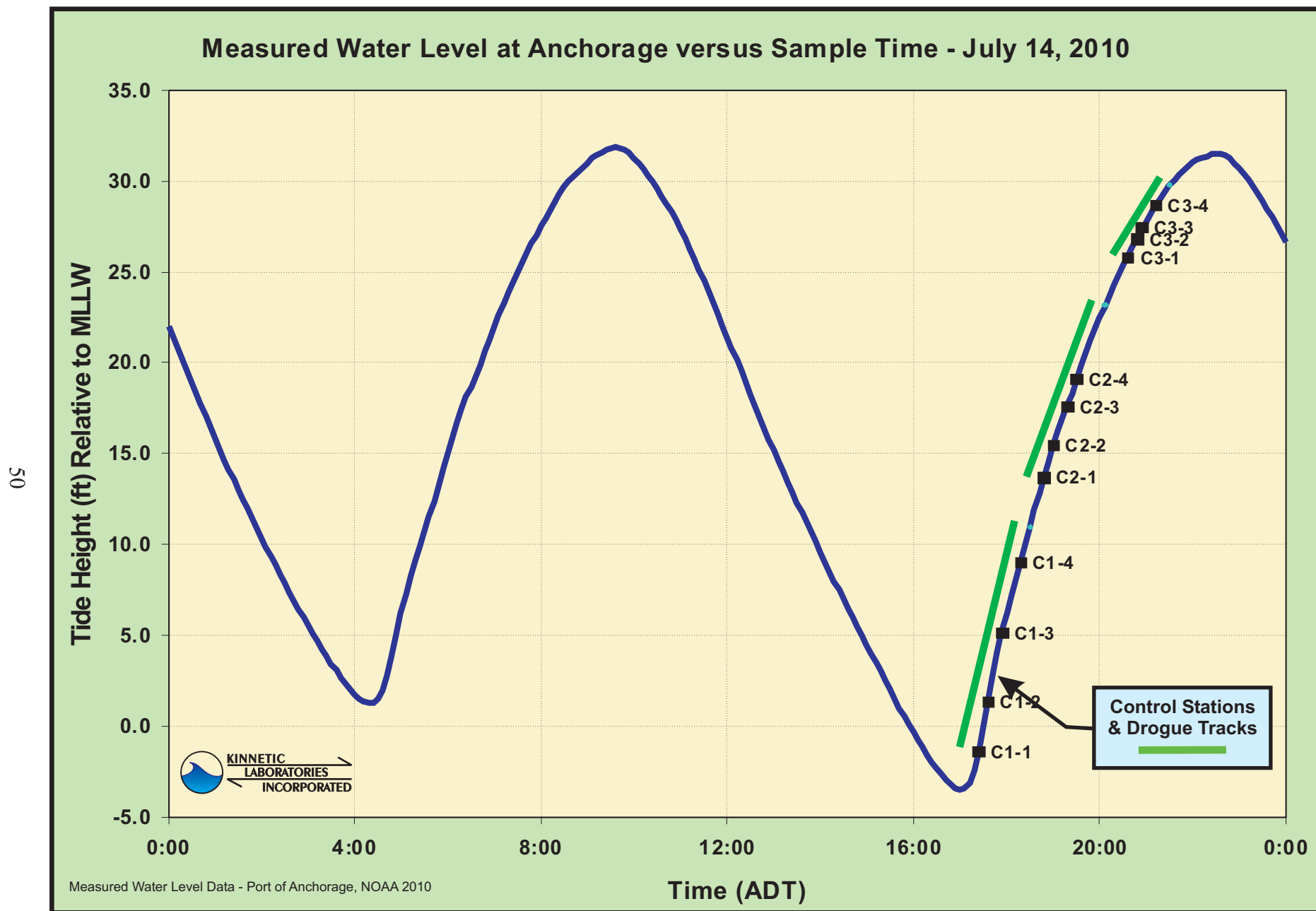


Figure 8. Tidal Information for Receiving Water Sampling, Control Tide.

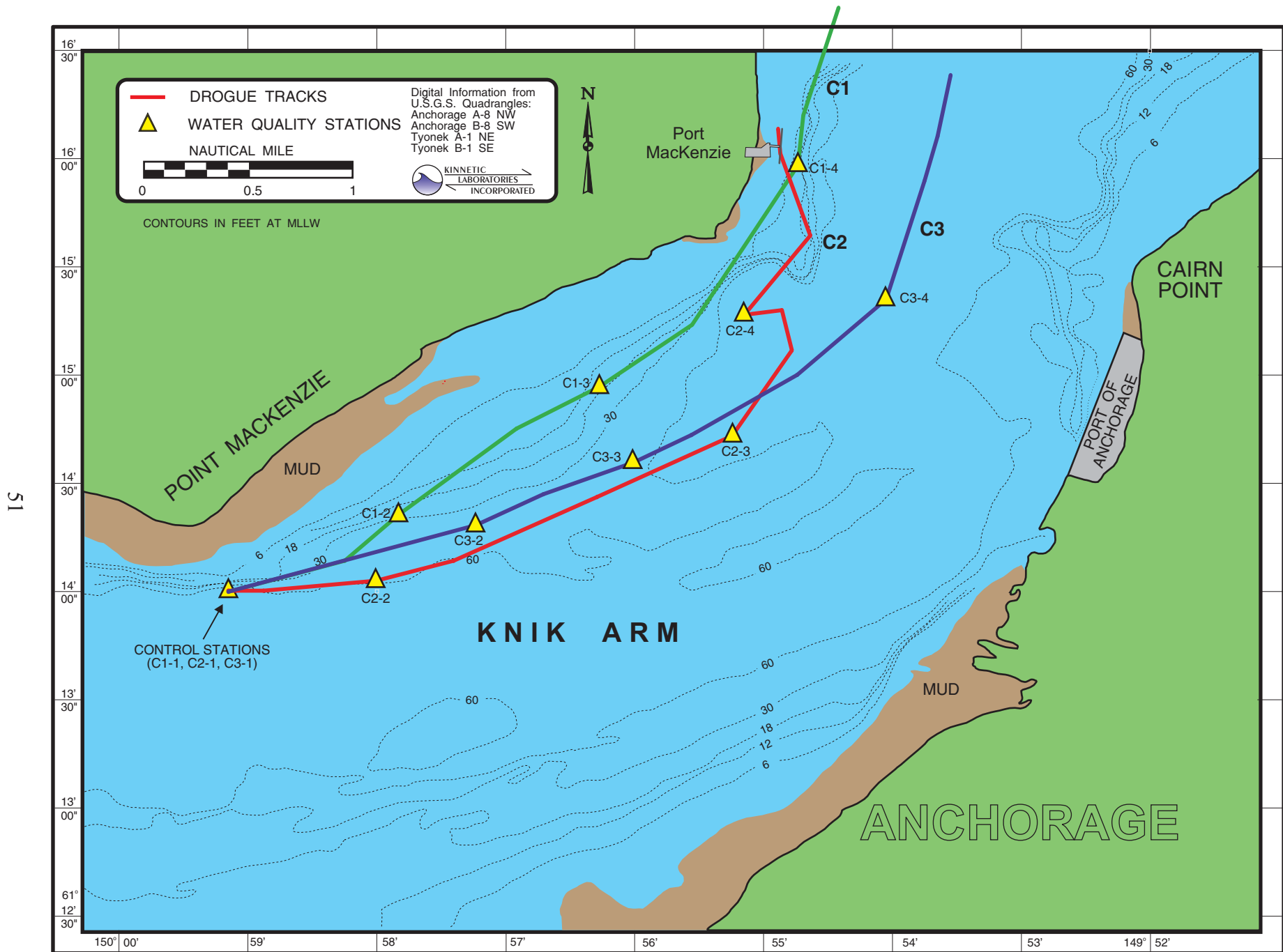


Figure 9. Summary of Control Drogue Tracks and Receiving Water Sampling Locations at Point MacKenzie, 14 July 2010.

In the field, samples were coded on their labels by location and depth to provide an easy summary of the water quality measurements obtained. The station designation is represented by: drogue drop location (C=control, E=ebb, and F=flood), the first number represents the drogue number, and the second number represents the station along the drogue's path. The final character represents surface (S), mid-depth (M), or bottom (B) sample. The waters of the inlet are extremely well-mixed both vertically and horizontally, as indicated by the CTD data. During the survey, water temperatures were in a narrow range with a minimum of 12.99 °C and a maximum of 13.93 °C (Table 15). Salinities were found to vary from a minimum of 8.10 parts per thousand practical salinity units (psu) to a maximum of 14.70 psu. Salinities were generally found to increase slightly during the flood and decrease on the ebb, as is typical for estuaries. As has been seen often times in the past, the control stations were found to be slightly warmer and less saline due to a greater influence from river runoff from the Matanuska and Knik Rivers. Values for pH ranged from 8.01 to 8.15 with little to no vertical stratification and no noticeable difference between the outfall and control sites. Values for DO collected in-situ by the CTD ranged from 9.09 to 9.90 mg/L with most concentrations being at or near 100 percent saturation (UNESCO, 1973).

Representative hydrographic profiles of water quality are presented for a ZID boundary station during ebb tide, Station E1-2, and a typical control station, Station C1-3 (Figure 10). The water column was found to be usually well mixed from the surface to the bottom at all stations, although some stations exhibited some salinity stratification as seen at Station E1-2. This stratification was not attributed to the outfall but was due to freshwater influences from local river inputs. Refer to Appendix C7 for CTD profile plots and detailed data from each water quality station.

Surface samples were obtained at each station for the analysis of color, TRC, fecal coliform bacteria, and turbidity. Color values were found to range < 5 to 5 color units on the platinum-cobalt scale. In 2007, the highest color values ever observed over the 20-year history of the program were seen. These high values were believed to be the result of insufficient suspended sediment removal by the laboratory as high levels were seen at both the outfall and control sites and were not the result of AWWU's discharge. Since 2008, to circumvent the sediment removal problem, samples were allowed to settle and decanted prior to submission for analysis.

During 2010, all measured receiving water TRC concentrations were below the MDL of 0.010 mg/L. The average TRC concentrations during the effluent sampling on 13 July was 0.12 mg/L, which is much lower than the MAEC based on the AWQS. It should be noted that the lowest MDL that was achievable by analysts due to seawater matrix interferences for TRC analysis in 2010 is between the AWQS 1-hr average acute limit of 0.013 mg/L and the 4-day chronic limit of 0.0075 mg/L. Also, the MDL that was achieved is substantially less than the 0.10 mg/L limit that ADEC considers reasonable for regulatory purposes. This year the ion selective electrode method (SM4500 Cl-G) was used for the receiving water sampling to reduce interferences from common oxidizing agents, temperature, turbidity, or color; but all TRC methods are subject to positive interferences in estuarine or marine waters.

Fecal coliform values this year were quite low and ranged from < 2 to 50 FC MPN/100 mL. Many of the fecal coliform concentrations were reported  $\leq 2$ ; this included 15 out of 36 samples at the ebb and flood stations combined and 8 out of 16 samples at the control stations. The overall median for fecal coliform for all of the outfall stations (both ebb and flood) was 4 FC MPN/100 mL; the median at the control stations was 3 FC MPN/100 mL. Turbidity values for



**Table 15. Hydrographic and Water Quality Data, 13 and 14 July 2010.**

Station Number	Time (ADT)	Latitude (North)	Longitude (West)	Depth (M)	Temp <sup>b</sup> (°C)	Salinity <sup>b</sup> (‰)	pH <sup>b</sup> (units)	D.O. <sup>b</sup> (mg/L)	Turbidity (NTU)	Color (units)	TRC (mg/L)	Fecal Coliform <sup>a</sup>
<b>JULY 13</b>												
E1-1S	0915	61° 12.347'	150° 01.275'	0.5	13.22	11.40	8.05	9.50	309	ND (5.0)	ND (0.010)	<2
-1M				4.5	13.24	12.30	8.09	9.47	575			
-1B				9.0	13.27	14.02	8.07	9.48	432			
E1-2S	0925	61° 12.305'	150° 02.001'	0.5	13.24	11.80	8.07	9.48	382	ND (5.0)	ND (0.010)	<2
-2M				6.0	13.28	13.36	8.11	9.48	415			
-2B				12.0	13.30	14.62	8.09	9.46	350			
E1-3S	0936	61° 12.171'	150° 02.787'	0.5	13.26	11.70	8.06	9.46	202	ND (5.0)	ND (0.010)	2
-3M				7.5	13.28	13.91	8.11	9.46	328			
-3B				14.5	13.31	14.70	8.10	9.48	376			
E1-4S	0952	61° 11.853'	150° 03.922'	0.5	13.29	13.02	8.06	9.41	215/221	ND (5.0)	ND (0.010)	5
-4M				7.5	13.34	14.41	8.11	9.40	354			
-4B				14.5	13.32	14.68	8.11	9.49	370			
E2-1S	1113	61° 12.337'	150° 01.275'	0.5	13.31	12.18	8.09	9.44	454	ND (5.0)	ND (0.010)	2
-1M				4.0	13.30	12.20	8.11	9.46	462			
-1B				8.0	13.30	12.21	8.11	9.50	464			
E2-2S	1116	61° 12.302'	150° 02.005'	0.5	13.30	12.22	8.08	9.44	456	ND (5.0)	ND (0.010)	2
-2M				6.0	13.30	12.23	8.11	9.49	433			
-2B				12.0	13.30	12.23	8.11	9.52	478			
E2-3S	1124	61° 12.131'	150° 03.101'	0.5	13.30	12.36	8.06	9.43	468	ND (5.0)	ND (0.010)	<2
-3M				7.5	13.29	13.36	8.11	9.51	488/489			
-3B				14.5	13.30	12.37	8.12	9.54	480			

**Table 15. Hydrographic and Water Quality Data, 13 and 14 July 2010. (continued)**

Station Number	Time (ADT)	Latitude (North)	Longitude (West)	Depth (M)	Temp <sup>b</sup> (°C)	Salinity <sup>b</sup> (‰)	pH <sup>b</sup> (units)	D.O. <sup>b</sup> (mg/L)	Turbidity (NTU)	Color (units)	TRC (mg/L)	Fecal Coliform <sup>a</sup>
E2-4S	1138	61° 11.617'	150° 04.272'	0.5	13.30	12.46	8.08	9.42	444	ND (5.0)	ND (0.010)	2
-4M				5.5	13.30	12.47	8.11	9.46	452			
-4B				10.5	13.30	12.48	8.11	9.50	450			
E3-1S	1317	61° 12.340'	150° 01.277'	0.5	13.29	10.93	8.11	9.54	485	ND (5.0)	ND (0.010)	5
-1M				2.0	13.29	10.93	8.11	9.56	494			
-1B				3.5	13.28	10.93	8.11	9.56	497			
E3-2S	1326	61° 12.235'	150° 01.982'	0.5	13.29	10.94	8.10	9.53	476	ND/ND (5.0)	ND (0.010)	8
-2M				3.0	13.29	10.94	8.11	9.54	487			
-2B				5.5	13.29	10.94	8.11	9.58	509/516			
E3-3S	1343	61° 12.020'	150° 02.709'	0.5	13.39	10.97	8.07	9.47	492	ND (5.0)	ND (0.010)	5
-3M				4.0	13.29	10.98	8.11	9.55	511			
-3B				7.5	13.29	10.98	8.11	9.58	513			
E3-4S	1402	61° 11.676'	150° 03.509'	0.5	13.34	10.98	8.09	9.52	248	ND (5.0)	ND (0.010)	<2
-4M				4.5	13.26	11.06	8.12	9.56	539			
-4B				8.5	13.26	11.07	8.12	9.59	577/587			
F1-1S	1600	61° 12.348'	150° 01.276'	0.5	13.20	9.62	8.12	9.67	260	ND (5.0)	ND (0.010)	13 / 50
-1M				1.0	13.18	9.62	8.13	9.67	525			
-1B				1.5	13.18	9.61	8.12	9.68	560			
F1-2S	1640	61° 12.425'	150° 00.555'	0.5	13.27	9.88	8.09	9.62	440	ND (5.0)	ND (0.010)	2 / 8
-2M				1.5	13.27	9.88	8.10	9.62	415			
-2B				2.5	13.27	9.88	8.10	9.64	475			
F1-3S	1650	61° 12.340'	149° 59.877'	0.5	13.30	9.94	8.11	9.63	320	ND (5.0)	ND (0.010)	5 / 8
-3M				1.0	13.29	9.94	8.11	9.64	410			
-3B				2.0	13.29	9.94	8.11	9.64	420			

**Table 15. Hydrographic and Water Quality Data, 13 and 14 July 2010. (continued)**

Station Number	Time (ADT)	Latitude (North)	Longitude (West)	Depth (M)	Temp <sup>b</sup> (°C)	Salinity <sup>b</sup> (‰)	pH <sup>b</sup> (units)	D.O. <sup>b</sup> (mg/L)	Turbidity (NTU)	Color (units)	TRC (mg/L)	Fecal Coliform <sup>a</sup>
F1-4S	1714	61° 12.187'	149° 59.254'	0.5	13.37	9.96	8.12	9.57	190/195	ND (5.0)	ND (0.010)	2 / 4
-4M				1.5	13.34	10.00	8.12	9.58	325			
-4B				2.5	13.33	10.00	8.12	9.59	370			
F2-1S	1748	61° 12.344'	150° 01.277'	0.5	13.40	11.17	8.11	9.49	550	ND (5.0)	ND (0.010)	6 / 8
-1M				3.5	13.40	11.18	8.12	9.51	560			
-1B				6.5	13.39	11.16	8.13	9.56	575			
F2-2S	1755	61° 12.537'	150° 00.677'	0.5	13.41	11.19	8.11	9.34	490	ND (5.0)	ND (0.010)	8 / 7
-2M				4.5	13.40	11.14	8.13	9.46	545			
-2B				8.5	13.39	11.08	8.13	9.49	580			
F2-3S	1802	61° 12.656'	149° 59.911'	0.5	13.40	10.99	8.10	9.50	310/285	ND (5.0)	ND (0.010)	8 / 4
-3M				4.5	13.38	11.04	8.12	9.55	540			
-3B				8.5	13.37	10.93	8.13	9.57	570			
F2-4S	1814	61° 12.534'	149° 58.802'	0.5	13.47	10.95	8.09	9.50	195	ND/ND(5.0)	ND (0.010)	5 / 4
-4M				5.0	13.39	11.09	8.12	9.53	590			
-4B				9.5	13.38	10.86	8.14	9.60	610			
F3-1S	1930	61° 12.343'	150° 01.284'	0.5	13.38	12.17	8.11	9.44	470	ND (5.0)	ND (0.010)	2 / 4
-1M				5.0	13.38	12.16	8.13	9.46	480			
-1B				9.5	13.38	12.18	8.14	9.49	495			
F3-2S	1935	61° 12.563'	150° 00.709'	0.5	13.39	12.18	8.10	9.42	445/460	ND (5.0)	ND (0.010)	2 / 2
-2M				6.5	13.38	12.16	8.14	9.46	490			
-2B				13.0	13.38	12.13	8.15	9.54	510			
F3-3S	1940	61° 12.741'	150° 00.016'	0.5	13.43	11.14	8.11	9.49	415	ND (5.0)	ND (0.010)	2 / 2
-3M				8.0	13.37	12.28	8.13	9.48	425			
-3B				15.5	13.36	12.30	8.14	9.55	450			

**Table 15. Hydrographic and Water Quality Data, 13 and 14 July 2010. (continued)**

Station Number	Time (ADT)	Latitude (North)	Longitude (West)	Depth (M)	Temp <sup>b</sup> (°C)	Salinity <sup>b</sup> (‰)	pH <sup>b</sup> (units)	D.O. <sup>b</sup> (mg/L)	Turbidity (NTU)	Color (units)	TRC (mg/L)	Fecal Coliform <sup>a</sup>
F3-4S	1956	61° 12.984'	149° 58.406'	0.5	13.38	12.13	8.04	9.43	430	ND/ND(5.0)	ND (0.010)	<2 / 4
-4M				10.5	13.33	12.48	8.13	9.50	450			
-4B				20.5	13.33	12.49	8.14	9.58	455			
<b>JULY 14</b>												
C1-1S	1728	61° 14.004'	149° 59.148'	0.5	13.47	8.56	8.09	9.70	61	ND (5)	ND (0.010)	8
-1M				2.0	13.36	9.10	8.11	9.61	510			
-1B				3.5	13.34	9.11	8.11	9.63	520			
C1-2S	1738	61° 14.361'	149° 57.828'	0.5	13.33	9.16	8.11	9.72	480	ND (5)	ND (0.010)	7
-2M				2.0	13.22	9.24	8.11	9.73	520			
-2B				4.0	13.21	9.24	8.11	9.73	565			
C1-3S	1755	61° 14.948'	149° 56.276'	0.5	13.45	9.15	8.08	9.71	370	ND (5)	ND (0.010)	2
-3M				6.5	13.25	9.23	8.13	9.74	525			
-3B				11.0	13.34	9.36	8.13	9.73	565			
C1-4S	1816	61° 15.967'	149° 54.730'	0.5	12.99	8.10	8.06	9.84	600/575	ND (5)	ND (0.010)	11
-4M				8.5	13.07	8.75	8.14	9.83	560			
-4B				16.5	13.05	8.72	8.14	9.90	570			
C2-1S	1850	61° 14.006'	149° 59.153'	0.5	13.93	9.92	8.09	9.31	380	ND (5)	ND (0.010)	8
-1M				12.0	13.69	10.00	8.13	9.44	440			
-1B				23.5	13.44	10.06	8.14	9.48	530			
C2-2S(A)	1900	61° 14.053'	149° 58.009'	0.5	13.87	10.00	8.08	9.40	340	ND (5)	ND (0.010)	4
-2S(B)				0.5	13.90	10.02	8.11	9.39	420	ND (5)	ND (0.010)	7
-2S(C)				0.5	13.88	9.92	8.10	9.42	525	5	ND (0.010)	2
-2M				6.0	13.76	9.99	8.12	9.51	320			
-2B				12.0	13.56	10.09	8.12	9.61	325			

**Table 15. Hydrographic and Water Quality Data, 13 and 14 July 2010. (continued)**

Station Number	Time (ADT)	Latitude (North)	Longitude (West)	Depth (M)	Temp <sup>b</sup> (°C)	Salinity <sup>b</sup> (‰)	pH <sup>b</sup> (units)	D.O. <sup>b</sup> (mg/L)	Turbidity (NTU)	Color (units)	TRC (mg/L)	Fecal Coliform <sup>a</sup>
C2-3S	1915	61° 14.721'	149° 55.246'	0.5	13.92	9.95	8.10	9.59	310	ND (5)	ND (0.010)	2
-3M				7.5	13.65	10.00	8.12	9.63	450			
-3B				15.0	13.62	9.99	8.13	9.66	530			
C2-4S	1928	61° 15.278'	149° 55.148'	0.5	13.42	8.99	8.06	9.53	490	ND/ND(5.0)	ND (0.010)	2
-4M				20.0	13.50	9.67	8.12	9.49	470			
-4B				39.5	13.39	9.53	8.13	9.09	560			
C3-1S(A)	2034	61° 14.004'	149° 59.152'	0.5	13.57	10.56	8.09	9.46	420	ND (5)	ND (0.010)	<2
-1S(B)				---	---	---	---	---	445	ND (5)	ND (0.010)	11
-1S(C)				---	---	---	---	---	455	ND (5)	ND (0.010)	<2
-1M				6.0	13.53	11.27	8.11	9.46	445/450			
-1B				12.0	13.54	11.26	8.11	9.45	490			
C3-2S	2048	61° 14.308'	149° 57.231'	0.5	13.47	11.29	8.09	9.45	380	ND (5)	ND (0.010)	5
-2M				6.5	13.45	11.33	8.11	9.47	470			
-2B				12.5	13.51	11.29	8.12	9.47	540			
C3-3S	2056	61° 14.308'	149° 57.231'	---	---	---	---	---	280	ND (5)	ND (0.010)	2
-3M				---	---	---	---	---	535			
-3B				---	---	---	---	---	545			
C3-4S	2110	61° 15.349'	149° 54.050'	0.5	13.39	10.19	8.01	9.50	435	ND (5)	ND (0.010)	<2
-4M				31.0	13.45	11.36	8.11	9.45	470			
-4B				60.5	13.42	11.41	8.12	9.46	500/515			

<sup>a</sup> Fecal coliform reported as MPN/100 mL.

<sup>b</sup> Values from CTD for 0.5 m depth taken as close to surface as possible.

--- Samples not collected.

ND None detected (detection limit).

## Outfall Station

## Control Station

85

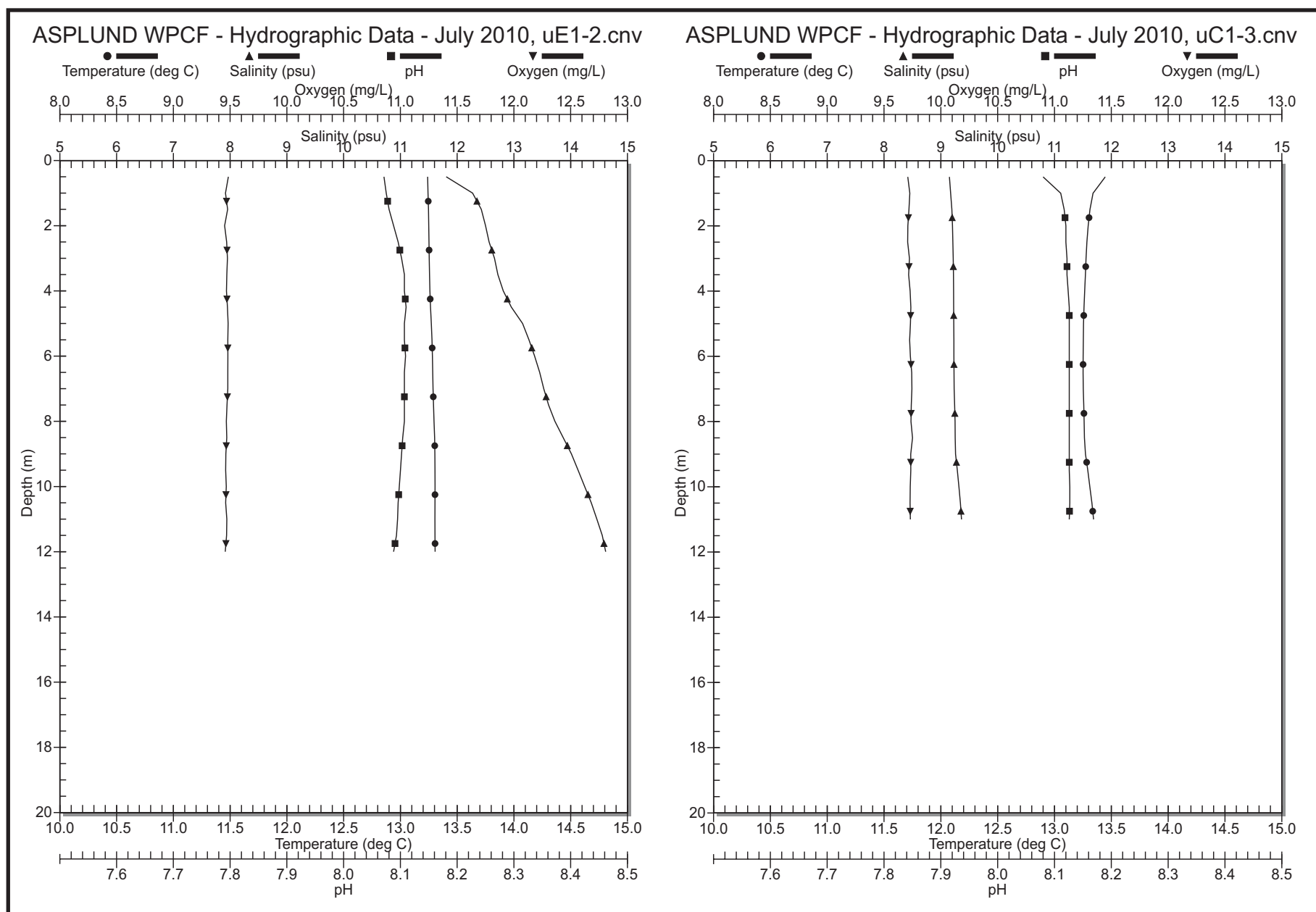


Figure 10. Sample Hydrographic Profiles from Outfall and Control Stations, July 2010.

water samples collected during the monitoring ranged from a low of 61 Nephelometric Turbidity Units (NTU) to a high of 610 NTU with lower values seen near the surface and near slack tide.

In addition to routine monitoring conducted at each receiving water quality station, supplemental surface samples were collected at the first three stations (diffuser, ZID boundary, and nearfield) along the first flood drogue trajectory at both the outfall and control sites. A sample of final effluent was also obtained at the same time for comparison. These supplemental samples were analyzed for BETX, polycyclic aromatic hydrocarbons (PAHs), dissolved and total recoverable trace metals, cyanide, and TSS.

The maximum dissolved arsenic, cadmium, copper, lead, nickel, silver, and zinc concentrations were seen at Station F1-1 located at low slack water above the diffuser, within the ZID (Table 16). The highest dissolved chromium concentration was seen at the outfall Station F1-3 and the highest dissolved mercury was seen at the control Station C1-2. The higher dissolved concentrations seen at Station F1-1 were probably a discharge related effect; however, all dissolved metals concentrations except copper were less than AWQS and SSWQC for the Point Woronzof area. Dissolved copper reported at 7.39 µg/L at Station F1-1 was higher than the AWQS of 3.1 µg/L, however this criterion does not apply to this station as it is located within the ZID mixing zone. All dissolved metals concentrations met AWQS at all ZID boundary, near field, and control stations.

Total metals concentrations were also quite variable with the highest levels for all metals except silver seen at the control site at Station C1-2 and most of the next highest levels seen at either Station C1-3 or F1-2. These results can be directly attributed to high ambient TSS levels at these stations. The TSS level at Station C1-2 (840 mg/L) was the highest of any station followed by Station F1-2 (690 and 698 mg/L) and Station C1-3 (580 mg/L). The differences between the outfall and control sites were the result of natural differences in TSS concentrations. Total suspended solid results ranged from 510 to 840 mg/L at the control stations compared to 280 to 698 mg/L at the outfall stations. The effluent sample had a TSS concentration of 57 mg/L.

Cyanide concentrations were found to be low at all receiving water stations including both the outfall and control locations. Cyanide concentrations ranged from a low 0.11 µg/L at C1-2 and F1-2 to a high of 0.19 µg/L at Station C1-1. Receiving water samples for cyanide were found to be well within the AWQS of 1 µg/L at all locations. The cyanide concentration in the effluent sample collected in conjunction with the receiving water sampling was 0.20 µg/L which is well below the MAEC of 181 µg/L.

Hydrocarbon analyses results are presented in Table 17. Total aromatic hydrocarbons (TAH) defined by the AWQS as BETX (EPA Method 602 samples from the concurrent summer dry sampling) was determined by summing benzene, ethylbenzene, toluene, and total xylenes. For values reported as ND, the reporting limit was used in the summation. Concentrations of TAH at the water quality stations were all ND ( $\leq 5.4$  µg/L) with the exception of Stations F1-1 and F1-3 which had concentrations of 6.5 µg/L and 5.8 µg/L, respectively. Concentrations of TAH were below the AWQS of 10 µg/L at all stations, including both outfall and control stations. With the exception of toluene at Stations F1-1 (2.1 µg/L) and F1-3 (1.37 µg/L), all BTEX constituents tested below method reporting limits across all stations. The effluent sample had a TAH concentration of 5.18 µg/L, which is also below the AWQS for the receiving water and significantly less than the MAEC of 1,810 µg/L.

**Table 16. Concentrations of Dissolved Metals, Total Recoverable Metals, Cyanide, and Total Suspended Solids in Receiving Water and Effluent Samples.** Values have not been blank corrected.

Station	Arsenic	Cadmium	Cyanide	Chromium	Copper	Mercury	Nickel	Lead	Silver	Zinc	TSS
	µg/L					ng/L	µg/L				mg/L
Dissolved Metals											
F1-1S (WITHIN ZID)	1.84	0.0822	NA	0.188	6.95	1.38	1.10	0.0266	0.00614	5.61	NA
F1-2S (ZID BOUNDARY)	1.58	0.0585	NA	0.188	1.36	0.383	0.842	0.00286	0.00231	0.464	NA
F1-3S (NEAR FIELD)	1.62	0.0536	NA	0.311	1.18	5.55	0.834	0.00389	0.00258	0.449	NA
C1-1S (CONTROL)	1.45	0.0462	NA	0.175	0.723	0.642	0.760	0.00178	<0.002	0.341	NA
C1-2S (CONTROL)	1.40	0.0512	NA	0.250	0.851	7.39	0.868	0.00248	0.00274	0.450	NA
C1-3S (CONTROL)	1.44	0.0514	NA	0.239	0.813	0.243	0.866	0.00334	0.00383	3.53	NA
EFFLUENT	1.50	0.121	NA	2.61	37.9	23.2	3.83	0.325	0.106	65.4	NA
DETECTION LIMIT	0.027	0.002	NA	0.055	0.023	0.10	0.026	0.001	0.002	0.042	NA
Total Metals											
F1-1S (WITHIN ZID) <sup>a</sup>	5.29	0.132	0.14 / 0.17	5.41	18.4	55.0	8.17	2.62	0.163	32.5	280
F1-2S (ZID BOUNDARY) <sup>a</sup>	8.60	0.153/0.149	0.11	11.6/13.2	24.1/24.6	52.0	13.9/14.4	7.21/7.47	0.102/0.103	37.4/38.5	690/698
F1-3S (NEAR FIELD) <sup>a</sup>	8.20/8.26	0.136	ND	11.4	22.9	41.8 / 40.8	13.3	6.79	0.0910	34.7	540
C1-1S (CONTROL)	7.21	0.120	0.19	10.2	19.9	44.8	12.3	5.42	0.0903	30.9	510
C1-2S (CONTROL)	10.8	0.175	0.11	16.6	33.5	82.4	19.7	9.88	0.145	50.7	840
C1-3S (CONTROL)	8.89	0.146	0.15	15.3	26.5	51.7	16.2	7.77	0.105	41.8	580
EFFLUENT	1.86	0.370	0.20	3.00	85.5	122	4.59	2.10	0.158	115	57
DETECTION LIMIT	0.027	0.002	0.10	0.055	0.023	0.10	0.026	0.001	0.002	0.042	0.41

<sup>a</sup> Field sample value/lab duplicate value (where applicable)

NA Not analyzed.



**Table 17. Supplemental Receiving Water and Effluent Hydrocarbon Analyses.**

PARAMETER	CONTROL FLOOD SAMPLES			ZID FLOOD SAMPLES			EFFLUENT <sup>a</sup>
	C1-1S	C1-2S	C1-3S	F1-1S	F1-2S	F1-3S	
<b>Volatile Organics (EPA 602) in µg/L with reporting limit in parenthesis</b>							
Benzene	< (0.400)	< (0.400)	< (0.400)	< (0.400)	< (0.400)	< (0.400)	< (1.8)
Toluene	< (1.00)	< (1.00)	< (1.00)	2.10	< (1.00)	1.37	9.8
Ethylbenzene	< (1.00)	< (1.00)	< (1.00)	< (1.00)	< (1.00)	< (1.00)	< (2.2)
Xylenes (Total)	< (3.00)	< (3.00)	< (3.00)	< (3.00)	< (3.00)	< (3.00)	4.7 J
TAH (as BETX)	5.4	5.4	5.4	6.5	5.4	5.8	18.5
<b>Polynuclear Aromatic Hydrocarbons (PAH) by GC/MS in µg/L</b>							
TPAH	0.044	0.052	0.068	0.648	0.113	0.082	5.18
<b>Total Aqueous Hydrocarbons (TAqH) in µg/L</b>							
TAqH <sup>b</sup>	5.45	5.45	5.47	7.15	5.51	5.88	23.7

<sup>a</sup> Effluent value from EPA 624 24 hour composite.

<sup>b</sup> Defined by the State of Alaska as BETX analytes plus PAH analytes from EPA Method 610 analysis; these calculated values include the full suite of PAH analyte values measured by from GERG.

< Below (detection limit).

All concentrations of individual PAHs were summed and reported as total PAHs (TPAH) in Table 17. TPAH concentrations were low with slightly higher concentrations seen at the outfall stations. The TPAH values ranged from 0.044 to 0.068 µg/L at the control stations and from 0.082 to 0.648 µg/L at the outfall stations. The TPAH concentration measured in the effluent sample was 5.18 µg/L. Total aqueous hydrocarbons (TAqH) as determined by the summation PAHs plus BETX were calculated for the six stations and effluent, with the contribution from BETX taken to be ≤5.4 µg/L for those stations with all NDs for the BETX contribution (Table 17). Concentrations of TAqH were below the AWQS of 15 µg/L at all stations, including both outfall and control stations. Control stations ranged in TAqH from 5.45 to 5.47 µg/L, while outfall station TAqH concentrations ranged from 5.51 to 7.15 µg/L. The concentration of TAqH in the effluent was estimated at 23.7 µg/L, compared to the MAEC of 2,715 µg/L.

### 3.2.2 Intertidal Zone and Stream Bacterial Sampling

Intertidal zone and stream bacteriological sampling was performed on 13 July 2010 (Table 18). Refer to Figure 3 for a map of the intertidal station locations and Figure 11 for stream locations. Intertidal sampling began approximately 2 hours prior to high tide at 20:10 ADT and was completed at 21:07 ADT. Two replicates were taken at all intertidal stations. Stream sampling was conducted from 12:40 to 13:16 ADT on 13 July 2010. In addition, replicate effluent samples were collected at the plant at 09:11 ADT on this date for fecal coliform analysis.

**Table 18. Summary of Bacterial Analyses, 13 July 2010.**

<b>Station and Replicate</b>	<b>Sample Time (ADT)</b>	<b>Fecal Coliform MPN/100 mL</b>
IT-1 Replicate 1	2107	8
IT-1 Replicate 2	2107	8
IT-2 Replicate 1	2102	2
IT-2 Replicate 2	2102	33
IT-3 Replicate 1	2052	<2
IT-3 Replicate 2	2052	5
IT-4 Replicate 1	2049	240
IT-4 Replicate 2	2049	46
IT-5 Replicate 1	2044	11
IT-5 Replicate 2	2044	17
IT-6 Replicate 1	2040	23
IT-6 Replicate 2	2040	17
IT-7 Replicate 1	2035	<2
IT-7 Replicate 2	2035	11
IT-C Replicate 1	2010	2
IT-C Replicate 2	2010	8
Plant Effluent Rep. 1	0911	8
Plant Effluent Rep. 2	0911	11
Fish Creek Rep. 1	1316	>2400
Fish Creek Rep. 2	1316	540
Chester Creek Rep.1	1301	<0.5
Chester Creek Rep.2	1301	49
Ship Creek Rep. 1	1240	49
Ship Creek Rep. 2	1240	79

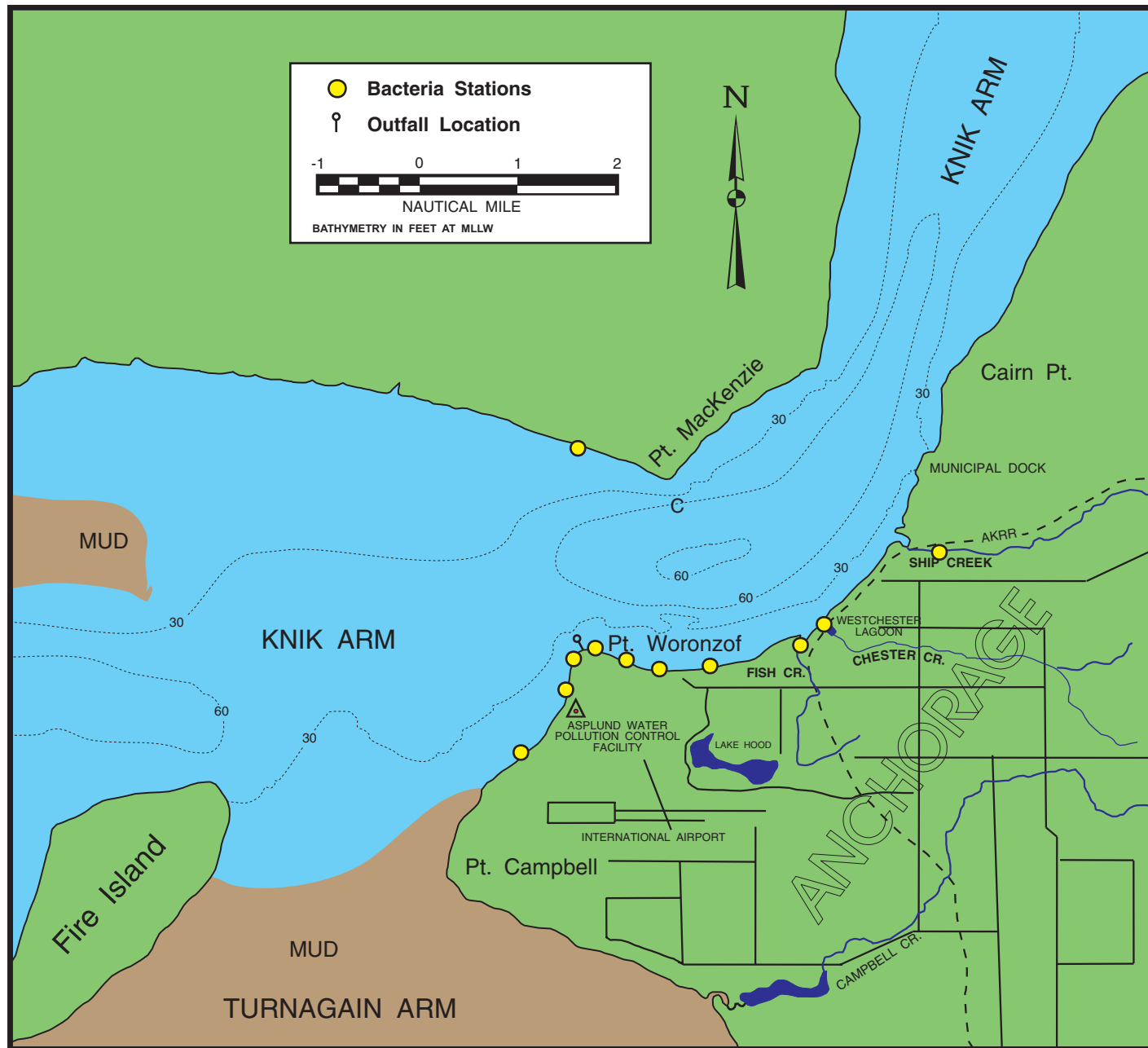


Figure 11. Stream and Intertidal Fecal Coliform Bacteria Sampling Locations.

Fecal coliform concentrations found in Fish, Chester, and Ship Creeks, that were sampled prior to the creeks entering Knik Arm, were higher than those seen in 2009 ranging from a low <0.5 FC MPN/100 mL in the first replicate at Chester Creek to a high of >2400 FC MPN/100 in the first replicate collected from Fish Creek. The replicate plant effluent samples taken on the same day showed fecal concentrations of 8 and 11 FC MPN/100 mL. Fecal coliform concentrations in the intertidal were variable this year and ranged from < 2 to 240 FC MPN/100 mL. The highest fecal concentrations (46 and 240 FC MPN/100 mL) were seen in the replicates at Station IT-4, 250 m east of the diffuser. The source of the elevated fecal coliform levels at this one site could not be determined since effluent samples taken that same day were found to be low and the intertidal area has been shown to be heavily used by both waterfowl and by hikers that access the beach at Pt. Woronzof and often use the area for walking their dogs.

## **4.0 QUALITY ASSURANCE/QUALITY CONTROL**

### **4.1 OBJECTIVES**

The monitoring program includes a comprehensive quality assurance/quality control (QA/QC) program that encompasses all aspects of the project, from initial sample collection and field observation recording through laboratory analysis and data analysis to reporting. The objectives of the QA/QC program were to fully document the field and laboratory data collected, to maintain and document data quality, and to ensure that the data collected are accurate, representative, and complete and are comparable with data collected through other EPA-regulated NPDES programs. The monitoring program was designed to allow the data to be assessed by the following parameters:

- Precision
- Accuracy
- Comparability
- Representativeness
- Completeness

Precision is a measure of agreement among repeated measurements of the same property which was assessed through duplicate and triplicate sampling and analysis. Accuracy is measure of the overall agreement of a measurement to a known value and includes a combination of random error (precision) and systematic error (bias) that are due to sampling and analytical operations. For this program these were assessed in field by comparing field instrumentation to known standards and in the laboratory by running standard reference material, performing blank spikes, matrix spikes, and comparing instrumentation to calibration standards. Comparability is a measure of the confidence with which one data set or method can be compared to another which was assured by utilizing standard EPA and other accepted sampling and laboratory protocols that could be traced back to known standards and using standard units of measure, such as navigational information that could be traced back to a know datum. Representativeness is the measure of the degree to which data accurately and precisely represent a characteristic of a population, parameter variations at a sampling point, or an environmental condition. This was assessed by determining sampling variability at a location by repeated sampling that then could be compared to laboratory variability. Completeness is a measure of the amount of valid data obtained that then can be compared to the amount of anticipated data as outlined in the project workplan.

These parameters were controlled by adhering to documented methods and procedures, by the analysis of quality control (QC) samples on a routine basis, the use of contract laboratories with existing QA/QC plans, accepted and defined data review and verification procedures, and a comprehensive sample documentation program. Throughout the program, KLI coordinated with the subcontracting laboratories to ensure that their in-house QA/QC programs were being implemented to meet the required standards.

Quality control activities in the field included adherence to documented procedures, including those in the program workplan, and the comprehensive documentation of sample collection and sample identification information. Sample integrity and identification were ensured by a rigidly-

enforced chain of custody program. The chain of custody procedure documents the handling of each sample from the time the sample was collected to the arrival of the sample at the laboratory.

Analytical methods in use throughout the program have been approved and documented by EPA. These methods were used as project-specific protocols to document and guide analytical procedures. Adherence to these documented procedures ensures that analytical results are properly obtained and reported.

## **4.2 FIELD QUALITY CONTROL**

Quality control activities in the field consisted of the following:

- adherence to documented procedures in the monitoring program workplan
- cross-checking of field identifications, measurements, and recording to ensure consistency, accuracy, and completeness of field logs.
- comprehensive documentation of field observations, sample collection, sample identification information, and navigation and drogue position information.

Sampling procedures proposed for this project have been successfully used for a number of years on the Asplund WPCF monitoring program. The use of documented and well-known procedures provides for greater likelihood of obtaining environmental samples uncontaminated by sampling procedures or apparatus. The use of project-specific field forms and data entry sheets also provide guidance to assure completeness of field data. Adherence to these procedures and use of these project documents helped ensure that data collected over the course of the project were complete, comparable, and accurate and that the study results are representative of conditions existing at the sampling sites.

### **4.2.1 Documentation**

For observations made in the field, cross-checking between personnel was used as the primary method of quality control. These included, for example, review of navigational information recorded on the drogue field log. As described in Section 2.5, sample documentation began in the field using pre-printed log forms, labels, chain of custody (COC) forms, and pre-determined sample identification numbers that were designed specifically for use on this project. This extensive field documentation provided a paper trail that exists for each sample or field observation and ensures credibility of the data. All field records were reviewed by the field crew leader as soon as possible after sampling was completed. After review and verification, field logs were filed at the KLI Anchorage office upon return from the survey.

Sample integrity and identification were ensured by the COC program. The chain of custody procedure documented the handling of a sample from the time the sample was collected to the receipt of the sample at the analytical laboratory. At the time of shipment, the field personnel kept a copy of the completed chain of custody form, and the original accompanied the samples to the laboratory.

### **4.2.2 Sample Handling**

Samples were frozen, chilled, and/or preserved as required by the appropriate methods in the field and until receipt at the laboratory. Samples were packed in coolers along with the

completed COC forms for shipment to analytical facilities as described in Section 2.0. Coolers were securely packed with ice packs as required and custody sealed with signed and dated fiber tape for shipment. Upon receipt by the laboratory the condition of the samples were noted on the COC form including; cooler temperature, broken or missing samples, etc.

#### **4.2.3 Navigation**

As described previously, navigation was accomplished with a DGPS system. The accuracy of the DGPS coordinates were verified by positioning the vessel over the diffuser during a low slack tide when the boil was evident and comparing DGPS readings with the known outfall location. Historical intertidal stations were re-acquired using a hand-held DGPS, distance and bearings, and visual sightings to temporary benchmarks and landmarks. All station information was entered on the appropriate field logs and reviewed by the field leader.

#### **4.2.4 Field Instrumentation**

Field equipment used for collection, measurement, and testing was subject to a strict program of control, calibration, adjustment, and maintenance. Care was taken to ensure that the instruments used for field measurements of temperature, salinity, DO, and pH were calibrated and checked with appropriate standards prior to and after each sampling event. The standards of calibration are in accordance with applicable criteria such as the U.S. Bureau of Standards, American Society of Testing and Materials (ASTM) Standards or National Institute of Standards and Technology (NIST), and follow the instrumentation manufacturer's recommended procedures.

Temperature calibration was ensured by pre-calibration at the factory with calibration checks of the electronic temperature sensor against a research grade NIST traceable thermometer reading taken from the same sample at the same time. The electronic sensor for salinity (conductivity) was also pre-calibrated at the factory and field checked against six ambient water samples which were collected for the analysis of salinity (SM 2520B) and compared to a certified Copenhagen Seawater Standard to verify the proper operation at the time of sampling. The DO probe was also pre-calibrated at the factory and field checked by comparison to saturated seawater. For pH, the probe was pre-calibrated using three known buffer solutions and checked in the field with a second field probe and three known buffer solutions.

#### **4.2.5 Sampling Variability**

Sampling variability was documented by sampling three replicates at one station for the water quality parameters. This included three replicate Niskin<sup>®</sup> bottle casts to obtain replicate turbidity samples and three replicate grabs at the surface for fecal coliform, color, and TRC analyses. In addition, triplicate casts of the CTD for pH, DO, temperature, and salinity were performed at one station in order to check reading variability from the probe's electronic sensors.

#### **4.2.6 Field Check Samples**

Field check samples include trip blanks for volatile organic analyses for EPA Methods 602 and 624, field blanks, field generated duplicates, standard reference materials (SRMs), or other samples of known concentration that may be sent to the laboratory. With the exception of the trip blanks which are initiated at the laboratory, field blanks and field duplicates samples were sent to the laboratory as blind samples to ensure unbiased reporting of results.

### **4.3 LABORATORY QUALITY CONTROL**

Analytical quality control for this project included the following:

- adherence to documented and approved procedures, including EPA, Standard Methods, etc., internal laboratory protocols, and respective laboratory QA/QC programs
- calibration of analytical instruments
- ability of each analytical laboratory to meet analytical precision, accuracy, limits of detection, and limits of quantification that meet EPA requirements
- use of quality control samples, internal standards, and surrogate solutions

The analytical laboratories used on this project operate under the quality assurance (QA) programs described in their QA management plans. These programs involve the participation of qualified and trained personnel; the use of standard operating procedures for analytical methodology and procedures; a rigorous system of documenting and validating measurements; maintenance and calibration of instruments; and the analysis of quality control samples for precision and accuracy tracking. The pertinent methods descriptions the laboratories are following are comprehensive and provide information concerning proper sample collection, receipt and login, processing, storage, and preservation; required apparatus and materials; analytical procedure; standardization and calibration techniques; quality control samples required; methods of calculating values and assessing data quality; and reporting and performance criteria.

#### **4.3.1 Documentation**

Documentation in the laboratory included signing the original COC forms, documenting sample condition upon receipt, and generating the internal documents that track samples through the laboratory (e.g., sample control logs, refrigerator logs, etc.). Any deviations from the prescribed methods or internal laboratory standard operating procedures (SOPs) were documented by the laboratory and included in a case narrative with the analysis report. Data affected by such deviations were appropriately qualified, as was any data that did not meet acceptable quality criteria. Typical data qualifiers included those denoting estimated concentrations (J) or not detected (ND or U), and method blank contamination (B).

#### **4.3.2 Calibration**

Calibration is an integral part of any instrumental analysis. Calibration requirements for each type of analysis to be used on this project are described in the appropriate methods. Typically, instrument calibration was performed daily or on a per batch basis as required by the method.

#### **4.3.3 Quality Control Procedures**

Internal laboratory quality control checks included the use of surrogate solutions and quality control samples such as procedural (or method) blanks, matrix spike/spike duplicates, standard reference materials (SRMs), method required QC check samples, and duplicates as specified in the EPA approved analytical procedures. In addition contract laboratories took part in EPA's annual DMRQA program to verify accuracy of their data. Surrogate compounds were spiked into samples as appropriate to assess individual sample matrix effects on sample analysis. This included QC samples such as procedural blanks and matrix spike samples. Surrogate compound



analyses were reported in percent recovery. Results from quality control samples allowed the laboratory to assess quality assurance parameters such as accuracy and precision of the data. Any data falling outside the acceptable criteria as defined in the methods were appropriately investigated by the laboratory, qualified, and described in the sample case narrative.

Method blanks are pure, organic- and/or metal-free reagent water that are run through the analysis process and used to verify that analyte concentrations are accurate and do not reflect contamination. Method blanks were analyzed as called for by each method, typically one per sample batch.

Laboratory accuracy was assessed by routine spiking of environmental samples with a standard addition as called for by the appropriate method. These sample matrix spikes and matrix spike duplicates (MS/MSD) were run on the organic analyses collected as part of both the in-Plant and receiving water monitoring components of the program. These spike samples are fortified with components of interest as required by the method following the initial analysis to check the ability of the method to recover acceptable levels and to determine accuracy of the data. Quality control charts are prepared and maintained by the laboratories where applicable to show the range of individual measurements encountered by following procedures such as those outlined EPA method guidance documents or in *Design of 301(h) Monitoring Programs for Municipal Wastewater Discharges to Marine Waters* (EPA, 1982b) and other guidance documents (e.g. EPA, 1994a and 1994b).

Trace metals analyses for the monitoring were supported through the use of certified standard reference materials (SRMs), which are quality control reference materials with known metals values that are obtained from the National Bureau of Standards and other sources. These SRMs were analyzed by the laboratory at the same time as the program samples in order to ensure laboratory accuracy. Results of the analyses of SRMs should fall within acceptable limits and can be expressed as percent recovery.

For receiving water quality samples, analytical and instrument variability was checked by laboratory splitting of one larger-volume field sample per sampling event into triplicates and analyzing the subsamples for the various water quality parameters. The individual measurements and concentration ranges were reported for each parameter of each split. In addition, duplicate analyses of samples split in the laboratory were used as a means to assess laboratory precision.

For other water quality parameters, the following summary of QA/QC procedures apply:

- Fecal Coliform Bacteria: *Escherichia coli* was used as a positive control for each analytical run. *Pseudomonas aeruginosa* was used as a negative control, and buffered dilution water was used as a blank. In addition, 10 % of the samples were run in duplicate.
- Enterococci Bacteria: *Streptococcus faecalis* was used as a positive control for each analytical run. *Escherichia coli* was used as a negative control, and buffered dilution water was used as a blank.

- Color: Fresh color standards were made prior to the beginning of the program. Samples were allowed to settle and decanted in the field and either pre-treated with paper filtration or centrifuged to remove turbidity and reported as "true color".
- Turbidity: The instrument was calibrated with a 20.0 NTU standard provided by the manufacturer. Due to the high turbidity in Cook Inlet, all samples were run at either the 10X or 100X scale on the nephelometer to ensure that the measured turbidities were within the range of the instrumentation. In addition, select samples were run in duplicate.
- Total Residual Chlorine: TRC was quantified with an ion selective electrode probe (SM4500 Cl-G) which requires a blank, blank spikes, and a series of laboratory calibration standards. To account for seawater matrix interference issues, the method blank and calibration standards were prepared with Cook Inlet background seawater.
- Salinity: Sensor is factory calibrated at a minimum of once per year and field checked with either a refractometer or salinometer.

#### **4.3.4 Method Detection Limits**

Depending on each laboratory's adopted terminology, the method detection limits (MDLs), practical quantitation limits (PQLs), or method reporting limits (MRLs) for the various analytes were determined using the appropriate method as described in the protocols. These MDLs, PQLs, and MRLs were reported with the data (see appendices) and are included in summary data tables as appropriate. Concentrations below the PQL, or MRL were typically qualified with the "ND" code for non-detect or "J" when reported as an estimated value.

#### **4.4 DATA REVIEW AND VALIDATION**

Data were verified by performing comparisons of final data against the original documentation, including the workplan, field logs and data sheets, and analytical reports. Any discrepancies were fully documented in the program files and described where necessary in the annual report. Data were validated according to accuracy, precision, and completeness for both the field sample collection and analytical laboratory components of the program. Qualitative evaluation and statistical procedures were used to check the quality of the field and chemical data as appropriate. The primary goals of these review and validation procedures are to ensure that the data:

- are representative of conditions in the study area
- are accurate
- demonstrate the required level of precision
- are comparable with data from other NPDES programs
- are acceptable for use as a tool to evaluate permit compliance
- allow independent technical appraisal of the program's ability to meet the monitoring program objectives.

Analytical data were subjected to review upon receipt from the laboratory following guidelines such as those published in *U.S. EPA Contract Laboratory Program National Functional Guidelines for Inorganic Data Review* (EPA 1994a), or *U.S. EPA Contract Laboratory Program National Functional Guidelines for Organic Data Review* (EPA 1994b). Items reviewed during

data validation included sample holding times, results for laboratory method blanks, matrix spike/spike duplicates (MS/SD), check standards or SRMs, field and laboratory duplicates, field and trip blanks, report completeness, and laboratory performance (i.e., ability to achieve method detection limits and adherence to QA/QC criteria established for this program). Items failing to meet such validation and review procedures were noted and corrected, if possible. Items that could not be corrected and fell outside of acceptable limits (e.g., a sample analyzed outside holding time) have been noted in data tables and in the appendices of this annual report if they occurred.

## **4.5 QUALITY ASSURANCE/QUALITY CONTROL RESULTS**

### **4.5.1 Field Instrumentation and Sampling Quality Control Results**

For influent, effluent, and sludge monitoring, field-generated duplicate influent and/or effluent samples were collected for the analysis of pesticides and enterococci bacteria during the June 2010 sampling. Results for these duplicate analyses are provided in Table 9 and the appendices, and were found to be within acceptance limits. Results from duplicate field samples collected for certain parameters during the receiving water sampling such as volatile organics, metals, cyanide, turbidity, and TSS are reported in the appropriate tables (Table 15, Table 16, and Table 17), and were found to be within acceptance limits.

Field blanks were collected for several parameters during each sampling event by pouring HPLC-grade deionized (DI) water into the appropriate sampling containers with the correct preservative. Trip blanks consisted of DI blank samples prepared at the laboratory that went through the same shipping and handling procedures as all the other sample containers of each analytical type; these remained unopened in the field.

The field blanks, trip blanks and method blanks analyzed using EPA Method 602 showed no measurable levels of the target compounds during the receiving water monitoring (Appendix C2). The trip blank analyzed in conjunction with the EPA 624 analyses for July showed trace levels of methylene chloride. The field generated blank prepared with the Asplund laboratory DI water analyzed in conjunction with the EPA 624 analyses for July also showed trace levels of methylene chloride. The associated EPA 624 method blank was a non-detect for all tested compounds (Appendix A1). The source of contamination in the trip and field generated blanks is unknown, although methylene chloride is a common laboratory contaminant that often shows up in both field samples and laboratory blanks. Methylene chloride was detected in the influent and effluent samples for both July and August sampling events. The method blank analyzed in August using EPA Method 625 showed no measurable levels of the target compounds.

Sampling variability for water quality parameters (fecal coliform, color, turbidity, and TRC) was determined by analyzing three discrete surface samples taken at Station C2-2S (Table 19). Where appropriate, the mean, standard deviation, and coefficient of variation are included in Table 19 to provide a measure of variability for parameters. The coefficient of variation for the various sample types was found to be 0 % for color, 0 % for TRC and 2.4 % for turbidity.

**Table 19. Sampling and Laboratory Variability for Water Quality Samples, 13 and 14 July 2010.**

Station	Subsample Designation	Fecal Coliform* (MPN/100 mL)	Color (units)	Turbidity (NTU)	TRC <sup>a</sup> (mg/L)
<b>SAMPLING VARIABILITY</b>					
C2-2S	A	4 [1.0-15]	<5	320	<0.010
[C2-3S (Turbidity only)]	B	7 [2.0-21]	<5	325	<0.010
	C	2 [1.0-10]	5	310	<0.010
Mean	---	4.3	5	318	<0.010
Standard Deviation	---	2.5	0	7.6	0
Coefficient of Variation (%)	---	58.1	0	2.4	0
<b>LABORATORY VARIABILITY</b>					
C3-1S	A	<2 [----]	<5	420	<0.010
	B	11 [4.0-29]	<5	445	<0.010
	C	<2 [----]	<5	455	<0.010
Mean	---	---	<5	440	<0.010
Standard Deviation	---	---	0	18.0	0
Coefficient of Variation (%)	---	---	0	4.1	0
E1-4S	A	NA	NA	215	NA
	B	NA	NA	221	NA
Relative % Difference	---	---	---	2.75	---
E2-3M	A	NA	NA	488	NA
	B	NA	NA	489	NA
Relative % Difference	---	---	---	0.2	---
E3-2S	A	NA	<5	NA	NA
	B	NA	<5	NA	NA
Relative % Difference	---	---	0	---	---
E3-2B	A	NA	NA	509	NA
	B	NA	NA	516	NA
Relative % Difference	---	---	---	1.36	---
E3-4B	A	NA	NA	577	NA
	B	NA	NA	587	NA
Relative % Difference	---	---	---	1.72	---
F1-1S	A	13	NA	NA	NA
	B	50	NA	NA	NA
Relative % Difference	---	118	---	---	---
F1-2S	A	2	NA	NA	NA
	B	8	NA	NA	NA
Relative % Difference	---	120	---	---	---
F1-3S	A	5	NA	NA	NA
	B	8	NA	NA	NA
Relative % Difference	---	46	---	---	---
F1-4S	A	2	NA	190	NA
	B	4	NA	195	NA
Relative % Difference	---	67	---	2.6	---

**Table 19. Sampling and Laboratory Variability for Water Quality Samples, 13 and 14 July 2010. (continued)**

Station	Subsample Designation	Fecal Coliform* (MPN/100 mL)	Color (units)	Turbidity (NTU)	TRC <sup>a</sup> (mg/L)
F2-1S	A	6	NA	NA	NA
	B	8	NA	NA	NA
	Relative % Difference	29	---	---	---
F2-2S	A	8	NA	NA	NA
	B	7	NA	NA	NA
	Relative % Difference	14	---	---	---
F2-3S	A	8	NA	310	NA
	B	4	NA	285	NA
	Relative % Difference	67	---	8.4	---
F2-4S	A	5	<5	NA	NA
	B	4	<5	NA	NA
	Relative % Difference	22	0	---	---
F3-1S	A	2	<5	NA	NA
	B	4	<5	NA	NA
	Relative % Difference	67	0	---	---
F3-2S	A	2	NA	445	NA
	B	2	NA	460	NA
	Relative % Difference	0	---	3.3	---
F3-3S	A	2	NA	NA	NA
	B	2	NA	NA	NA
	Relative % Difference	0	---	---	---
F3-4S	A	<2	<5	NA	NA
	B	4	<5	NA	NA
	Relative % Difference	---	0	---	---
C1-4S	A	NA	NA	600	NA
	B	NA	NA	575	NA
	Relative % Difference	---	---	4.3	---
C2-4S	A	NA	<5	NA	NA
	B	NA	<5	NA	NA
	Relative % Difference	---	0	---	---
C3-1M	A	NA	NA	445	NA
	B	NA	NA	450	NA
	Relative % Difference	---	---	1.1	---
C3-4B	A	NA	NA	500	NA
	B	NA	NA	515	NA
	Relative % Difference	---	---	3.0	---

\* 95% confidence intervals indicated in brackets (American Public Health Association, 1998. Standard Methods for the Examination of Water and Wastewater. 20th Edition. Washington, D.C. Table 9221.IV.)

<sup>a</sup> All TRC measurements performed in triplicate and tested below detection level.

NA Not analyzed

--- Not applicable

Three replicate fecal coliform samples were also collected at Station C2-2. The samples yielded results of 7, 4 and 2 FC MPN/100 mL. Mean, standard deviation, and coefficient of variation are not determined for fecal coliform due to the nature of the analysis which yields only a most probable number of bacteria per 100 mL. Instead, the  $\pm 95$  % confidence limits for each sample are typically provided. Fecal coliform values were within the confidence limits.

Variability and calibration checks of the electronics probe were done by performing repeated profiles of temperature, pH, DO, and salinity at one station (C2-2). Results of these calibration checks for the Seabird CTD show that probe variability for temperature, salinity, pH, and DO was extremely low in all cases with a maximum coefficient of variation of  $\leq 0.74$  %; seen at the bottom, with lower variability seen at shallower depths in the water column (Table 20). In addition, a precision thermometer was used to verify CTD temperature readings, and all of the CTD sensors were factory-calibrated prior to field deployment. The salinity, temperature, DO, and pH probes were found to be accurate and within calibration during the survey.

#### **4.5.2 Laboratory Quality Control Results**

Full analytical data and laboratory case narratives are provided by most laboratories and are included in the appendices. Laboratory duplicate analyses, where performed, were found to have a high degree of precision and were within the acceptance criteria for relative percent difference (RPD). Laboratory duplicates were performed for a number of organic analyses and were found to be within acceptance limits.

The compound 4,4-DDT exceeded 40 % RPD between the primary and confirmation columns in the July 2010 effluent and sludge samples. During the August 2010 sampling event 4,4-DDT in the influent and in the effluent also exceeded the 40 % RPD between the primary and confirmation columns criteria. In such cases the higher of the two results are reported. All data were appropriately qualified and full laboratory case narratives appear in Appendices A1 and B1.

In addition to the standard laboratory QC procedures, color and turbidity samples collected at Station C3-1 during the receiving water sampling were split in the laboratory and analyzed in triplicate. Mean, standard deviation, and coefficient of variation are reported in Table 19 for these samples. Coefficient of variation was shown to be 0 % for color and 2.4 % for turbidity. For analyses where samples were run in duplicate, such as TRC, turbidity, and color, the relative percent difference between duplicates was calculated. The RPDs for laboratory duplicate analyses were found to generally be very low and within acceptable limits. Duplicate results for turbidity ranged from 0.2 to 8.4 % RPD. Color duplicate results showed 0 % difference between duplicates. Duplicate TRC analyses were all read in triplicate and tested below detection limits for a 0 % RPD.

Laboratory accuracy was assessed through the use of surrogate recoveries, sample and control spikes and duplicates, and SRMs. Detailed QA/QC results for all contract laboratory analyses are provided in the appendices corresponding to each analysis. Surrogates are compounds that were added to each sample and QC sample that were analyzed by GC methodology, such as volatile organic compounds (EPA 602 and 624/8260B), semi-volatile organic compounds (EPA 625/8270C), pesticides (EPA 608/8081A/8082 and 614/8141A), and dioxins (EPA 8280A). Some instances of surrogate recoveries outside QC recovery limits were found during the 2010 influent, effluent, and sludge monitoring effort. These deviations are discussed in the case

**Table 20. Seabird SEACAT SBE-19 CTD Probe Variability Check, 14 July 2010.**

Depth (m)	Temperature (°C)			Salinity (ppt)			pH (units)			DO (mg/l)			Mean (units)				Standard Deviation				Coeff. of Variation (%)			
C2-	2A	2B	2C	2A	2B	2C	2A	2B	2C	2A	2B	2C	Temp	Sal	pH	DO	Temp	Sal	pH	DO	Temp	Sal	pH	DO
0.5	13.87	13.91	13.88	10.00	10.02	9.92	8.08	8.11	8.10	9.40	9.39	9.42	13.89	9.98	8.10	9.40	0.017	0.054	0.013	0.015	0.12	0.54	0.15	0.16
1.0	13.83	13.84	13.83	10.00	10.01	9.99	8.09	8.10	8.10	9.42	9.41	9.40	13.83	10.00	8.10	9.41	0.002	0.008	0.007	0.012	0.01	0.08	0.08	0.13
1.5	13.82	13.82	13.82	9.99	10.00	9.99	8.10	8.11	8.11	9.44	9.42	9.42	13.82	9.99	8.10	9.43	0.001	0.004	0.007	0.014	0.01	0.04	0.09	0.15
2.0	13.82	13.82	13.81	9.99	9.99	9.98	8.10	8.11	8.11	9.46	9.46	9.45	13.82	9.99	8.11	9.45	0.007	0.004	0.009	0.007	0.05	0.04	0.11	0.08
2.5	13.82	13.82	13.79	9.98	9.99	9.98	8.11	8.11	8.12	9.48	9.47	9.44	13.81	9.99	8.11	9.47	0.016	0.004	0.005	0.024	0.11	0.04	0.06	0.25
3.0	13.81	13.82	13.78	9.98	9.99	9.98	8.11	8.12	8.12	9.48	9.49	9.45	13.80	9.98	8.11	9.47	0.021	0.006	0.005	0.020	0.15	0.06	0.06	0.22
3.5	13.81	13.81	13.76	9.97	9.98	9.99	8.11	8.11	8.12	9.50	9.50	9.45	13.79	9.98	8.11	9.48	0.028	0.010	0.005	0.032	0.20	0.10	0.06	0.34
4.0	13.81	13.80	13.75	9.97	9.98	9.99	8.12	8.12	8.12	9.54	9.49	9.48	13.79	9.98	8.12	9.50	0.032	0.013	0.002	0.033	0.23	0.13	0.03	0.35
4.5	13.81	13.79	13.74	9.97	9.98	10.01	8.12	8.12	8.12	9.51	9.51	9.48	13.78	9.98	8.12	9.50	0.037	0.019	0.002	0.015	0.27	0.19	0.02	0.16
5.0	13.80	13.78	13.73	9.97	9.98	10.01	8.12	8.12	8.12	9.53	9.52	9.49	13.77	9.99	8.12	9.51	0.039	0.021	0.000	0.023	0.29	0.21	0.00	0.24
5.5	13.79	13.77	13.72	9.98	9.99	10.02	8.12	8.12	8.12	9.54	9.55	9.49	13.76	10.00	8.12	9.52	0.036	0.020	0.001	0.033	0.26	0.20	0.01	0.35
6.0	13.76	13.75	13.71	9.99	10.00	10.02	8.12	8.12	8.12	9.51	9.56	9.50	13.74	10.00	8.12	9.52	0.028	0.016	0.001	0.030	0.20	0.16	0.01	0.32
6.5	13.74	13.74	13.70	10.00	10.00	10.03	8.13	8.12	8.12	9.52	9.53	9.50	13.73	10.01	8.12	9.52	0.023	0.015	0.003	0.013	0.17	0.15	0.04	0.14
7.0	13.72	13.73	13.66	10.01	10.01	10.05	8.13	8.12	8.12	9.53	9.55	9.52	13.70	10.02	8.12	9.53	0.034	0.021	0.003	0.016	0.25	0.21	0.04	0.17
7.5	13.71	13.72	13.63	10.02	10.02	10.07	8.13	8.12	8.12	9.56	9.56	9.52	13.69	10.03	8.12	9.55	0.048	0.032	0.003	0.025	0.35	0.32	0.04	0.26
8.0	13.70	13.72	13.61	10.02	10.02	10.07	8.13	8.12	8.12	9.55	9.56	9.48	13.68	10.04	8.12	9.53	0.057	0.031	0.003	0.040	0.42	0.31	0.04	0.42
8.5	13.70	13.71	13.60	10.02	10.02	10.07	8.13	8.12	8.12	9.56	9.55	9.47	13.67	10.04	8.12	9.53	0.058	0.030	0.003	0.053	0.43	0.30	0.03	0.56
9.0	13.68	13.69	13.58	10.03	10.03	10.07	8.13	8.12	8.12	9.59	9.55	9.45	13.65	10.04	8.12	9.53	0.065	0.021	0.003	0.070	0.47	0.21	0.04	0.74

narratives that were provided by each laboratory which fully detail all QC issues for both sampling events and explain any QC deviations; these are provided in the appendices. As indicated by the laboratories, in all instances the data were not significantly affected by any QC issue.

Matrix spike (MS), matrix spike duplicate (MSD), laboratory control spike (LCS), and duplicate control spike (DCS) are samples and blanks that are spiked with target compounds of interest to determine percent recovery and relative percent difference between duplicates. The QC criteria include an acceptable recovery range and an RPD that should not be exceeded. Total metals, dissolved metals, cyanide, volatile organic compounds, and semi-volatile organic compounds met QC criteria for MS, MSD, LCS, and DSC with few exceptions for all analyses on the program. Detailed case narratives were provided by each laboratory which fully detail all QC issues for both sampling events and explain any QC deviations; these are provided in Appendices A1, A2, B1, B2, C1, C2, C3, and C4.

Trace metals analyses for the influent, effluent, sludge, and receiving water testing were supported through the use of SRMs, which are quality control reference materials with known metals values that are obtained from the National Bureau of Standards, National Institute of Standards and Technology, or other certified standards. These SRMs were analyzed by the laboratories at the same time as the project samples in order to ensure laboratory accuracy. Results of the analyses of SRMs should fall within acceptable limits and can be expressed as percent recovery. All metals SRM results were within acceptance limits (Appendix C4).

Method blanks (or procedural blanks) were also analyzed for most analyses on the program. Method blanks consist of pure, organic- or metal-free reagent grade water that is run through the analysis process and used to verify that analyte concentrations are accurate and do not reflect contamination. The method blank for EPA624 analyzed during the July 2010 influent and effluent sampling indicated low levels of acetone and methylene chloride were present. Also, the laboratory method blanks for the August 2010 EPA 624 (influent and effluent) and EPA 8260 (sludge), tested positive for trace levels methylene chloride. A detailed case narrative for these samples provided by the laboratory which fully details all QC issues is provided in Appendix A1 and B1.

During the receiving water program, trace levels of metals were detected at their MDLs in the method blanks for the ultra-low level metals analyses that were performed by Battelle. This is typical for ultra-low level metals analyses, in that trace levels are seen in method blanks in the low parts per trillion range and is not an indication of contamination. Metals concentrations seen in the receiving water were 2-3 orders of magnitude higher than that seen in these method blanks and all other QC criteria were within acceptance limits, therefore no further action was necessary or taken.

The method blank analyses performed with the TSS analyses showed no results above method detection limits. In addition to the typical method blanks, buffered dilution water was used as a blank for fecal coliform and enterococci bacteriological analyses. All blanks run for fecal coliform and enterococci showed no growth.



## **5.0 DISCUSSION**

### **5.1 INFLUENT, EFFLUENT, AND SLUDGE MONITORING**

The NPDES permit for the Asplund WPCF requires compliance with applicable State water quality standards as promulgated in Chapter 70 of the Alaska Administrative Code entitled "Water Quality Standards" (18 AAC 70; ADEC, 1999). This chapter requires that criteria outlined in "EPA Quality Criteria for Water" (also known as "The Red Book"; EPA, 1976), the revised quality criteria for water (EPA 1986b), and other applicable criteria as referenced in the AWQS be met in applicable receiving waters at every point outside of the ZID boundary. Also, as noted in Section 1.1.1, the State of Alaska water quality regulations include SSWQC for the Point Woronzof area of Cook Inlet for turbidity and the dissolved fraction of arsenic, cadmium, hexavalent chromium, copper, lead, mercury, nickel, selenium, silver, and zinc. Since the issuance of the current permit, EPA has approved ADEC's proposed use of dissolved metals for all of the State's marine water quality criteria, approved all of ADEC's proposed SSWQC for Upper Cook Inlet, and removed Alaska from the National Toxics Rule list (EPA 2006; Sept. 15, 2006 letter to ADEC). Except for cadmium and mercury, where the dissolved standard changed from 9.3 to 8.8 µg/L for cadmium and from 0.025 to 0.94 µg/L for mercury, all other AWQS metals criteria are the same as those in the SSWQC. Even though EPA has approved the use of dissolved metals criteria for the AWQS, the current SSWQC will most likely remain in affect for the Point Woronzof area for permit renewal as those are also listed in the current AWQS. To be conservative, we have used the more restrictive criteria for dissolved cadmium and mercury to evaluate the data in this report. For other parameters such as TRC we have utilized the current AWQS (ADEC 2009), since those criteria will be utilized for the permit renewal process. Finally, the permit itself includes some effluent limitations that must be met. The following sections discuss the parameters of concern in regards to the requirements of the NPDES permit or the AWQS as well as historical data from the WPCF, other POTWs, and other EPA data.

#### **5.1.1 Influent and Effluent Monitoring**

Table 21 lists permit effluent limitations and marine water quality criteria that are applicable to the current NPDES permit; it includes each of the parameters required to be monitored by the permit. Most of the values shown are the chronic toxicity criteria for salt water aquatic life. Chronic toxicity criteria concentrations are lower than acute toxicity criteria concentrations; therefore, the more stringent of the two values were used here for comparison. The MAEC for each constituent was calculated from the outfall design dilution factor of 142:1 (conservative substances) or 180:1 (non-conservative; TRC, ammonia, etc.), the water quality criteria, and the natural background concentrations as determined historically at the control site near Point MacKenzie. It was assumed that the final effluent would be diluted by a minimum factor of 143 by the time it reached the boundary of the ZID. For most metals, the MAECs were calculated from the SSWQC for dissolved metals contained in the AWQS for the Point Woronzof area.

To determine compliance with State water quality standards, Table 21 values have been compared with effluent values found in Table 8 through Table 11 as well as those in Table 16 and Table 17. The AWWU 2010 maximum effluent concentrations shown in Table 21 were the maximum encountered during the calendar year either during AWWU's in-plant monitoring, the toxic pollutant and pesticide monitoring events, pretreatment monitoring, or the supplemental effluent monitoring that was performed as part of the receiving water sampling. For metals, both total and dissolved concentrations in the effluent were compared against their MAEC, since it is

**Table 21. NPDES Requirements, State of Alaska Water Quality Standards, and AWWU 2010 Maximum Concentrations for Effluent Comparisons.**

Parameter	Receiving Water Quality Standard <sup>a</sup>	Maximum Allowable Effluent Concentration <sup>b</sup> (MAEC)	AWWU 2010 Maximum Effluent Concentration <sup>c</sup>
Antimony (µg/L)	146	Human health, not listed for saltwater aquatic life 20,607	ND (1.00) <sup>d,e</sup>
Arsenic (µg/L)	36	Chronic toxicity, measured as dissolved 4,882	1.86 <sup>f</sup>
Beryllium (µg/L)	11	For the protection of aquatic life in soft fresh water 1,513	ND (0.4) <sup>d,e</sup>
Cadmium (µg/L)	9.3 (8.8) <sup>k</sup>	Chronic toxicity, measured as dissolved 1,322 (1,250)	0.370 <sup>f</sup>
Chromium (VI) <sup>h</sup> (µg/L)	50	Chronic toxicity, measured as dissolved 7,038	3.00 <sup>f</sup>
Copper (µg/L)	3.1	Chronic toxicity, measured as dissolved 317	85.5 <sup>e</sup>
Lead (µg/L)	8.1	Chronic toxicity, measured as dissolved 1,140	3.79 <sup>e</sup>
Mercury (µg/L)	0.025	Chronic toxicity, measured as dissolved 2.73	0.302 <sup>f</sup>
Nickel (µg/L)	8.2	Chronic toxicity, measured as dissolved 978	4.59 <sup>e</sup>
Selenium (µg/L)	71	Chronic toxicity, measured as dissolved 10,136	ND (5.0) <sup>e</sup>
Silver (µg/L)	1.9	Acute toxicity, measured as dissolved 257	0.158 <sup>e</sup>
Thallium (µg/L)	2,130	Acute toxicity to saltwater aquatic life 306,567	ND (1.0) <sup>d,e</sup>
Zinc (µg/L)	81	Chronic toxicity, measured as dissolved 11,249	118 <sup>e</sup>
Cyanide (µg/L)	1	For marine aquatic 181	21 <sup>e</sup>

**Table 21. NPDES Requirements, State of Alaska Water Quality Standards, and AWWU 2010 Maximum Concentrations for Effluent Comparisons. (continued)**

Parameter	Receiving Water Quality Standard <sup>a</sup>		Maximum Allowable Effluent Concentration <sup>b</sup> (MAEC)	AWWU 2010 Maximum Effluent Concentration <sup>c</sup>
		life		
Total Aqueous Hydrocarbons (TAQH) (µg/L)	15	Growth and propagation of fish, shellfish, aquatic life, and wildlife including seabirds, waterfowl, and furbearers	2,715	23.68 <sup>f</sup>
Total Aromatic Hydrocarbons as BETX (µg/L)	10	Same as above	1,810	18.5 <sup>d</sup>
pH (pH units)		<i>g</i>	6.5 - 8.5	6.8 – 7.5 <sup>i</sup>
Total Residual Chlorine (TRC) (mg/L)		<i>g</i>	Daily Max. 1.2	Daily Max. 0.88 <sup>i</sup>
BOD <sub>5</sub> (mg/L)		<i>g</i>	Monthly Avg. 240 Weekly Avg. 250 Daily Max. 300 Monthly Removal Rate ≥30 %	Monthly Avg. 133 <sup>i</sup> Weekly Avg. 173 <sup>i</sup> Daily Max. 219 <sup>i</sup> (Monthly Avg. Rate 43 – 53% ) <sup>i</sup> Annual Avg. Removal (48% in 2010) <sup>i</sup>
Total Suspended Solids (TSS) (mg/L)		<i>g</i>	Monthly Avg. 170 Weekly Avg. 180 Daily Max. 190 Monthly Removal Rate ≥30%	Monthly Avg. 53 <sup>i</sup> Weekly Avg. 61 <sup>i</sup> Daily Max. 86 <sup>i</sup> Monthly Avg. Removal 77 – 81% <sup>i</sup> , Annual Avg. Removal 79% <sup>i</sup>
Total Ammonia (mg/L) <sup>j</sup>		8.1 Acute 1.2 Chronic <sup>j</sup>	1466 217	Monthly Max. 22.9 <sup>i</sup>

**Table 21. NPDES Requirements, State of Alaska Water Quality Standards, and AWWU 2010 Maximum Concentrations for Effluent Comparisons. (continued)**

Parameter	Receiving Water Quality Standard <sup>a</sup>	Maximum Allowable Effluent Concentration <sup>b</sup> (MAEC)	AWWU 2010 Maximum Effluent Concentration <sup>c</sup>
Fecal Coliform (FC MPN/100 mL)	<sup>g</sup>	Monthly geometric mean of at least five samples shall not exceed 850.  Not more than 10% of samples shall exceed 2600.	Monthly mean maximum was 18 <sup>i</sup>  The criterion of not more than 10% of samples exceeding 2600 was met in 2010. <sup>i</sup>
<b>Other Detected Effluent Constituents with Specific Alaska Water Quality Criteria</b>			
4,4'-DDT	0.13 Acute 0.001 Chronic	23.4 0.18	0.094 P,D <sup>d</sup>
1,4 Dichlorobenzene	400 <sup>l</sup>	56,800	0.73 J
Diethyl phthalate	23,000 <sup>l</sup>	3,266,000	7.04
Phenol	21,000 <sup>l</sup>	2,982,000	18.0
Toluene	6,800 <sup>l</sup>	965,600	9.8

- <sup>a</sup> Alaska Administrative Code, 2003. Water Quality Standards, Chapter 70, 18 AAC 70.020(b)
- <sup>b</sup> For conservative substances, effluent water quality criteria were determined by assuming a dilution of 142:1 at the ZID boundary, where: MAEC = 142 \* (Criteria - Natural Background Concentration) + Criteria; pollutant concentrations in the effluent should not exceed these values. For non-conservative substances, a dilution of 180:1 was utilized in the MAEC calculation.
- <sup>c</sup> For metals, the maximum effluent concentration was determined from both total and dissolved concentrations.
- <sup>d</sup> Values from July 2010 or August 2010 toxic pollutant and pesticide samplings.
- <sup>e</sup> Values from AWWU's pretreatment program.
- <sup>f</sup> Values from effluent tested during receiving water sampling event.
- <sup>g</sup> MAECs are not based on water quality criteria but instead are specified in MOA's 2000 NPDES permit.
- <sup>h</sup> All samples tested as total chromium.
- <sup>i</sup> Values from AWWU's in-plant monitoring.
- <sup>j</sup> Ammonia receiving water criteria based on pH of 8.0, temperature of 15.0°C, and salinity of 20psu.
- <sup>k</sup> Cadmium standard based on revised EPA level that was approved for State of Alaska (2001).
- <sup>l</sup> Alaska water quality Human Health criteria for consumption of water & aquatic organisms.
- D Reported result from a dilution.
- J Estimated value (below MRL but above MDL)
- P > 40% RPD between primary and confirmation column results. The higher result is reported.

assumed that all of the metals contained in the effluent are potentially bioavailable upon entering the receiving water. All effluent concentrations were found to be much lower than the MAECs from the permit or computed from the water quality standards provided for in the AWQS. In addition, the permit limitations for all parameters were met for the 2010 program year. When the MAECs in Table 21 were compared to all of the 2010 data including; AWWU's self-monitoring effluent data, the toxic pollutant and pesticides sampling events (July and August 2010), the pretreatment monitoring data, and the effluent data from the receiving water sampling, no metals or cyanide values exceeded their MAECs. The highest concentrations of either total or dissolved metals seen in 2010 were all well below their respective MAECs.

The one metal that most closely approached its MAEC at any time was copper, and this was considerably below its MAEC. The maximum concentration of total copper was 85.5 µg/L compared to a MAEC of 317 µg/L. The highest dissolved copper concentration that was seen was 48 µg/L. All other effluent dissolved metals concentrations were found to be much less than their respective MAECs.

Those metals without AWQS, while analyzed as both total and dissolved metals as called for by the permit, are compared to total recoverable metal MAECs as provided by EPA criteria and include antimony, beryllium and thallium. Total metals concentrations for antimony, beryllium, and thallium were below detection limits, and thus well below their MAECs. As in past years, total recoverable metals detected in the influent and final effluent were compared with data from an EPA study of 40 Publicly Owned Treatment Works (POTWs) in Table 22 (EPA, 1982a). Without exception, metals and cyanide values were lower than or within the range of those detected in other POTWs from across the nation, even though the Asplund WPCF provides only primary treatment as compared to secondary treatment provided at the other plants that were examined in this study.

Historic influent and effluent total recoverable metals and cyanide concentrations collected as part of AWWU's self-monitoring program are presented in Table 23 and Table 24. It should be noted that prior to 2000 when the permit requirements changed, dissolved metals had only been analyzed in a single sample of effluent collected each year during the receiving water sampling. Beginning in August 2000, dissolved metals from the effluent have been analyzed in both the summer wet and summer dry sampling events and as part of the pretreatment monitoring. Concentrations are low and fairly consistent over time. Concentrations of total recoverable metals concentrations seen in the influent and effluent during 2010 generally fell within the range of concentrations seen during prior years, although zinc was found to be high in one influent sample. Concentrations of dissolved metals were generally found to fall within range of concentrations seen over the prior five years as seen in Table 23. In 2008 through 2010, cyanide concentrations were typical of the long term average whereas in 2007 cyanide appeared to be slightly elevated although barely outside the historic range.

Total arsenic concentrations in the final effluent had remained fairly steady over the last five years, and 2010 values remained well within the historic range. The maximum total arsenic concentration in final effluent seen during 2010 was < 5 µg/L, compared to an MAEC of 4,882 µg/L (Table 21). Arsenic values are not a serious concern for this permit in terms of effluent concentrations, since the concentration in the final effluent is so much lower than the MAEC.

**Table 22. Comparison Between Influent/Effluent Analysis Results for Anchorage and 40 POTWs.<sup>a</sup>**

Parameter	Anchorage Values				40 POTW Study Values				
	2010 Concentration <sup>b</sup> (µg/L)				Frequency of Detection (%)		Range Detected (µg/L)		Influent Median
	Summer-Dry		Summer-Wet		Influent	Secondary Effluent	Influent	Secondary Effluent	(µg/L)
	INF	EFF	INF	EFF					
VOLATILES <sup>c</sup>									
Benzene	ND	ND	ND	ND	61	23	1-1560	1-72	2
Chloroform	2.0 J	3.5 J	2.0 J	3.0 J	91	82	1-430	1-87	7
1,4-Dichlorobenzene	0.91 J	0.73 J	0.78 J	0.67 J	17	3	2-200	3-9	NA
Ethylbenzene	ND	ND	ND	ND	80	24	1-730	1-49	8
Methylene chloride	4.8 J,B	6.0 B	15 B	7.2 B	92	86	1-49000	1-62000	38
Tetrachloroethene	ND	ND	ND	ND	95	79	1-5700	1-1200	23
Toluene	10	9.8	5.3	6.3	96	53	1-13000	1-1100	27
Xylene (Total)	7.8 J	4.7 J	8.6 J	5.7 J	NA	NA	NA	NA	NA
SEMI-VOLATILES <sup>c</sup>									
Phenol	180 D	ND	336	18.0	79	29	1-1400	1-89	7
TOTAL METALS & OTHER COMPONENTS									
Antimony	ND	ND	ND	ND	14	13	1-192	1-69	NA
Arsenic	ND	ND	ND	ND	15	12	2-80	1-72	NA
Beryllium	ND	ND	ND	ND	3	1	1-4	1-12	NA
Cadmium	3.67	0.660	ND	ND	56	28	1-1800	2-82	3
Chromium	3.09	ND	4.16	ND	95	85	8-2380	2-759	105
Copper	61.7	82.9	67.7	41.9	100	91	7-2300	3-255	132
Lead	4.02	2.38	3.19	1.81	62	21	16-2540	20-217	53
Mercury	ND	ND	ND	ND	70	31	0.2-4	0.2-1.2	0.517
Molybdenum	ND	ND	ND	ND	NA	NA	NA	NA	NA
Nickel	5.12	3.62	5.95	3.96	79	75	5-5970	7-679	54
Selenium	ND	ND	ND	ND	9	10	1-10	1-150	NA
Silver	1.38	ND	1.20	ND	71	25	2-320	1-30	8
Thallium	ND	ND	ND	ND	3	2	1-19	1-2	NA
Zinc	175	118	178	94.9	100	94	22-9250	18-3150	273
Cyanide	ND	ND	ND	5.0	100	97	3-7580	2-2140	249

<sup>a</sup> Source: EPA, 1982. *Fate of Priority Pollutants in POTWs*. Final Report, Volume I, Effluent Guidelines Division, WH-552, EPA 440/1-82/303

<sup>b</sup> Data from NPDES 2009 toxic pollutant and pesticide monitoring

<sup>c</sup> Only analytes detected in either the influent or effluent and in the 40 POTW study are included.

J Estimated value

NA Not available

ND Not detected

**Table 23. Comparison of Toxic Pollutants and Pesticides in Anchorage's Final Effluent to the Previous Five Years.** Values in brackets are from EPA Method 602 where available.

Pollutant	2005		2006		2007	
	Dry	Wet	Dry	Wet	Dry	Wet
	5/23-24	8/1-2	6/27-28	8/15-16	6/19-20	8/7-8
<b>ORGANICS (µg/L)</b>						
Acetone	ND	ND	ND	ND	ND	ND
Benzene	ND [ND]	3.3 J	ND [0.43J]	0.59 J	0.58 J[ND]	0.27 J
Benzoic acid	ND	ND	ND	ND	ND	ND
Benzyl alcohol	ND	ND	ND	ND	ND	ND
Bis-(2-ethylhexyl) phthalate	15	14 B	18	10	20 J	0.20 J
Butyl benzyl phthalate	7.4 J	2.7 J	ND	ND	ND	ND
Chloroform	2.6 J	4.1 J	3.6 J	3.8 J	3.4 J	3.4 J
Chloromethane	ND	1.4 J	ND	1.1 J	ND	ND
*1,3-Dichlorobenzene	ND	ND	ND	ND	9.5	9.5
*1,4-Dichlorobenzene	ND	ND	1.0 J	1.1 J	9.5	9.5
Diethyl phthalate	8.5 J	10	7.7 J	7.9 J	10 J	10 J
Ethylbenzene	ND/ND	ND	ND [3.8]	0.62 J	2.9 J[ND]	2.9 J[ND]
3&4 Methylphenol	ND	ND	ND	ND	3.1 J	3.1 J
Methylene Chloride	2.6 J,B	5.0 B	4.5 J	3.3 J,B	ND	ND
Phenol	19	21	18	13	24 J	24 J
Tetrachloroethene	ND	ND	4.0 J	1.4 J	6.4	6.4
1,1,2,2-Tetrachloroethane	ND	ND	ND	ND	3.3 J	3.3 J
Toluene	4.0 J [5.6]	10	8.7 [11]	8.7	21[4.9]	21[4.9]
* Total Xylenes	1.5 J [0.93]	2.4 J	6.7 J [17]	3.1 J	23 [2.6]	23 [2.6]
Total Hydrocarbons as Oil and Grease <sup>a</sup>	21400	17600	19900	19400	14900	14900
Total Aromatic Hydrocarbons as BETX	1.5 J [0.93]	20.7	25.4 [32.23]	13.0	47.5 [8.5]	12.1
<b>OTHER</b>						
Cyanide (µg/L)	ND	ND	5	ND	9J/9J/10	59
Asbestos (million fibers/L)	ND	ND	ND	ND	ND	ND

**Table 23. Comparison of Toxic Pollutants and Pesticides in Anchorage's Final Effluent to the Previous Five Years. (continued)** Values in brackets are from EPA Method 602 where available.

Pollutant	2008		2009		2010	
	Dry	Wet	Dry	Wet	Dry	Wet
	6/17-18	8/13-14	6/23-24	7/28-29	7/13-14	
<b>ORGANICS (µg/L)</b>						
Acetone	ND	ND	ND	100	130 B	180
Benzene	ND[ND]	ND	0.240 J	ND	ND	ND
Benzoic acid	ND	ND	241	141	ND	ND
Benzyl alcohol	ND	ND	28.4	25.8	ND	ND
Bis-(2-ethylhexyl) phthalate	33 J,B	14 J	ND	ND	14	ND
Butyl benzyl phthalate	ND	ND	ND	ND	ND	ND
Chloroform	3.0 J	3.0 J	3.24	2.0 J	3.5 J	3.0 J
Chloromethane	ND	ND	ND	ND	95 J	ND
* 1,3-Dichlorobenzene	ND J	ND	ND	ND	ND	ND
* 1,4-Dichlorobenzene	ND	0.96 J	0.790 J	0.68 J	0.73 J	0.67 J
Diethyl phthalate	7.7 J	7.8 J	ND	ND	ND	ND
Ethylbenzene	0.76 J [0.86]	ND	1.18	ND	ND	ND
Methylene Chloride	3.4 J, B	7.7 B	11.0	4.2 J	6.0 B	7.2 B
3&4 Methylphenol	ND	ND	119	78.1	99	84.0
Phenol	23 J	25 J	18.1	ND	ND	ND
Tetrachloroethene	0.94 J	ND	1.14	ND	ND	ND
1,1,2,2-Tetrachloroethane	ND	ND	ND	ND	ND	ND
Toluene	6.8[6.4]	6.4	8.84	5.5	9.8	6.3
* Total Xylenes	4.5 J [2.5 COL]	2.6 J	3.68	1.7 J	4.7 J	5.7 J
Total Hydrocarbons as Oil and Grease <sup>a</sup>	28000	32000 B	37600	64900	35500	49100
Total Aromatic Hydrocarbons as BETX	17.1	19.0	13.9	17.2	18.5	16.0
<b>OTHER</b>						
Cyanide (µg/L)	9.0 J (0.19 / 0.25)	6.9 J	9.0 J (0.19 / 0.25)	6.9 J	ND	5.0
Asbestos (million fibers/L)	ND	ND	ND	ND	14.8	ND



**Table 23. Comparison of Toxic Pollutants and Pesticides in Anchorage's Final Effluent to the Previous Five Years. (cont.)** Values in brackets from EPA Method 602 where available.

Pollutant	2005		2006		2007	
	Dry	Wet <sup>b</sup>	Dry	Wet <sup>b</sup>	Dry	Wet
	5/23-24	8/1-2	6/27-28	8/15-16	6/19-20	8/7-8
<b>TOTAL METALS (µg/L)</b>						
Antimony	0.65	ND/ND	0.65	0.58/0.60	ND	ND
Arsenic	3	2	2.4	2.6/2.8	2.18	1.83
Beryllium	0.1	ND	ND	ND/ND	ND	ND
Cadmium	0.3	ND	0.27	0.24/0.25	ND	ND
Chromium	5	3	2.2	2.8/2.7	2.09	1.90
Copper	52	54	51	49/48	40.7	34.8
Lead	6	2	2.7	2.9/2.9	2.08	3.57
Mercury	0.10	0.11	0.13	0.10	ND	ND
Molybdenum	NT	NT	NT	NT	10.8	6.81
Nickel	4	4	2.8	4.2/4.3	3.91	4.05
Selenium	0.47	ND/ND	1.4	1.4/ND	ND	ND
Silver	1.7	1.4	3.1	2.3/2.4	2.22	1.26
Thallium	ND	ND/ND	ND	ND/ND	ND	ND
Zinc	90	90	100	97/96	83.8	84.8
<b>DISSOLVED METALS (µg/L)</b>						
Antimony	0.45	ND/ND	ND	ND	ND/ND	ND
Arsenic	2	2	1.2	1.9	1.93/1.95	1.69
Beryllium	0.1	ND	ND	ND	ND/ND	ND
Cadmium	ND	ND	ND	ND	ND/ND	ND
Chromium	4	2	0.79	0.55	1.32/1.50	1.33
Copper	37	42	1.4	26	25.4/25.2	26.9
Lead	6	2	0.064	0.42	ND/ND	2.34
Mercury	ND	ND	0.035	0.027	ND	ND
Molybdenum	NT	NT	NT	NT	9.95/10.2	6.19
Nickel	ND	ND	3.7	2.9	3.24/3.33	3.54
Selenium	ND	ND/ND	9.6	1.1	ND/ND	ND
Silver	ND	0.9	ND	ND	1.01/1.07	ND
Thallium	ND	ND/ND	ND	ND	ND/ND	ND
Zinc	40	40	3.3	35	48.7/48.0	49.5
<b>PESTICIDES (µg/L)</b>						
alpha-BHC	ND	ND	ND	0.023 J	ND	ND
beta-BHC	ND	0.037 J	ND	ND	ND	ND
Dieldrin	ND	ND	0.010 J,COL	0.021 J	ND	ND
Endosulfan II	0.075 J	ND	ND	0.042 J	ND	ND
Endrin	ND	ND	ND	ND	ND	0.011 J, COL
Endrin ketone	ND	ND	0.012 J,COL	ND	ND	ND
Heptachlor	0.17 J	0.99	ND	0.54	0.015 J, COL	0.58
2,4'-DDD	ND	ND	ND	ND	ND	ND
4,4'-DDD	ND	ND	ND	ND	ND	0.047 J
4,4'-DDE	ND	ND	0.0080 J,COL	ND	ND	0.016 J, COL
4,4'-DDT	ND	ND	ND	ND	ND	ND
Malathion	0.49 J, CHI	ND	ND	ND	ND	ND

**Table 23. Comparison of Toxic Pollutants and Pesticides in Anchorage's Final Effluent to the Previous Five Years. (cont.)** Values in brackets from EPA Method 602 where available.

Pollutant	2008		2009		2010	
	Dry	Wet	Dry	Wet	Dry	Wet
	6/17-18	8/13-14	6/23-24	7/28-29	7/13-14	8/
<b>TOTAL METALS (µg/L)</b>						
Antimony	0.57 J	0.47 J	ND	ND	ND	ND
Arsenic	2.5 J	2.2 J	ND	ND	ND	ND
Beryllium	ND	ND	ND	ND	ND	ND
Cadmium	0.51 J	0.60 J	ND	ND	0.660	ND
Chromium	2.3 B,J	2.2 B,J	ND	ND	ND	ND
Copper	36	37	48.9	34.9	82.9	41.9
Lead	2.8	2.3	2.35	1.8	2.38	1.81
Mercury	0.047 J	0.041 J	ND	ND	ND	ND
Molybdenum	6.4 B	4.5	ND	ND	ND	ND
Nickel	2.5	3.2	3.62	2.98	3.62	3.96
Selenium	1.3 J	ND	ND	ND	ND	ND
Silver	1.3	1.4	1.56	ND	ND	ND
Thallium	ND	ND	ND	ND	ND	ND
Zinc	120	110 B	97.4	81.4	118	94.9
<b>DISSOLVED METALS (µg/L)</b>						
Antimony	0.39 J	0.33 J	ND	ND	ND	ND
Arsenic	ND	2.2 J	ND	ND	ND	ND
Beryllium	ND	ND	ND	ND	ND	ND
Cadmium	ND	ND	ND	ND	ND	ND
Chromium	1.1 J	3.5	ND	ND	ND	ND
Copper	14	15	14.0	26.8	48.0	13.3
Lead	0.78 J	0.64 J	ND	0.917	0.964	0.356
Mercury	ND	ND	ND	ND	0.302	ND
Molybdenum	6.4	4.2	ND	ND	ND	ND
Nickel	2.7	2.8	2.94	2.77	3.73	3.85
Selenium	ND	2.5 J	ND	ND	ND	ND
Silver	0.51 J	0.59 J	ND	ND	ND	ND
Thallium	ND	ND	ND	ND	ND	ND
Zinc	20	27	38.4	58.4	75.3	28.0
<b>PESTICIDES (µg/L)</b>						
alpha-BHC	ND	0.0082 J,COL	ND	ND	ND	ND
beta-BHC	ND	0.036 J,COL	ND	0.014 CHI	ND	ND
Dieldrin	ND	ND	ND	ND	ND	ND
Endosulfan II	ND	ND	ND	ND	ND	ND
Endrin	ND	ND	ND	ND	ND	ND
Endrin ketone	ND	ND	ND	ND	ND	ND
Heptachlor	ND	ND	ND	ND	ND	ND
2,4'-DDD	ND	ND	ND	ND	0.012	ND
4,4'-DDD	ND	ND	ND	ND	ND	ND
4,4'-DDE	ND	ND	ND	0.010	ND	ND
4,4'-DDT	ND	ND	0.25 CHI	ND	0.086 P	0.094 P,D
Malathion	ND	ND	ND	ND	ND	ND

*a* EPA method 1664 HEM *b* Duplicate effluent collected (field duplicate) or analyzed (lab duplicate) shown as value/duplicate value

\* Non-priority pollutants J Estimated value D Sample diluted for analysis.

CHI or P More than 40% RPD between primary and confirmation results. The higher of the two results is reported.

COL More than 40% RPD between primary and confirmation results. The lower of the two results is reported.

B Compound also detected in method blank ND Not detected NT Not tested

**Table 24. Historical Discharge Monitoring Data (1986 - Present) for Influent and Effluent Total Metals and Cyanide.** Concentrations are rounded values in µg/L. Values represent a range of minimum (Min) and maximum (Max) results for 1986-2004 as available. Results for 2005-2010 are from pretreatment monitoring (Avg of six results from both the wet and dry sampling events).

Year	Average Flow (mgd)	Arsenic		Beryllium		Cadmium		Copper		Lead		Mercury		Nickel		Silver		Zinc		Chromium		Cyanide	
		Inf.	Eff.	Inf.	Eff.	Inf.	Eff.	Inf.	Eff.	Inf.	Eff.	Inf.	Eff.	Inf.	Eff.	Inf.	Eff.	Inf.	Eff.	Inf.	Eff.	Inf.	Eff.
1986-2004 Min	23	<1	<1	<0.03	<0.03	<0.3	0.2	36	10	<1	<1	<0.1	<0.1	<1	<1	<1.5	1	54	38	<1	<1	<0.4	1
1986-2004 Max	40	26	16	0.6	0.3	20	30	280	150	149	50	3.0	1.5	77	60	30.4	98	260	407	112	120	85	50
2005 Avg	28	3	3	0.1	0.1	0.4	0.3	90	53	7	4	0.3	0.1	9	5	3.3	1.7	155	92	6	4	<0.9	<0.9
2005 Min	24	2	<2	<0.1	<0.1	<0.3	<0.3	78	47	4	2	0.1	0.1	6	4	2.8	1.3	140	80	4	1	<0.9	<0.9
2005 Max	30	4	4	0.1	0.1	0.4	0.3	116	57	10	6	0.5	0.1	17	8	4.3	2.2	180	110	7	7	<0.9	<0.9
2006 Avg	28	3	3	0.5	0.5	0.4	0.3	84	57	5	3	0.1	0.1	7	4	4	3	190	99	7	3	1.0	1.7
2006 Min	26	3	2	<0.5	<0.5	0.3	0.2	75	48	2	2	0.1	0.1	4	3	2	2	180	94	4	2	<1.0	<1.0
2006 Max	32	3	3	<0.5	<0.5	0.5	0.4	93	77	8	4	0.2	0.1	11	6	6	3	200	110	15	7	<1.0	5
2007 Avg	28	2	2	<1.0	<1.0	<1.0	<1.0	63	39	4	2	0.3	<0.2	5	4	3	2	156	90	4	2	14	22
2007 Min	25	1	2	<1.0	<1.0	<1.0	<1.0	30	34	2	2	<0.2	<0.2	4	4	1	1	55	79	2	2	6	9
2007 Max	32	3	3	<1.0	<1.0	<1.0	<1.0	94	43	10	4	0.9	<0.2	9	4	3	2	299	105	7	3	25	59
2008 Avg	29	2	2	<1.0	<1.0	1.7	1.2	66	37	6	2	0.1	0.1	5	3	2	1	187	104	5	3	6	7
2008 Min	27	2	2	<1.0	<1.0	0.3	0.2	55	35	4	2	0.1	0.04	4	2	2	1	160	79	4	2	2	3
2008 Max	30	3	3	<1.0	<1.0	6.8	4.9	92	41	13	3	0.2	0.1	6	3	3	2	290	120	8	3	10	10
2009 Avg	28	<5	<5	<0.4	<0.4	<0.5	<0.5	55	59	4	4	<0.2	<0.2	4	4	1.3	1.5	124	119	2	2	<5	11
2009 Min	26	<5	<5	<0.4	<0.4	<0.5	<0.5	33	35	2	2	<0.2	<0.2	3	3	<1	<1	63	81	<2	<2	<5	<5
2009 Max	31	<5	<5	<0.4	<0.4	<0.5	<0.5	74	128	9	8	<0.2	<0.2	5	6	1.8	3.2	165	220	3	3	<5	21
2010 Avg	27	<5	<5	<0.4	<0.4	1.16	0.53	67.5	54.8	4.7	2.3	<0.2	<0.2	6.1	4.2	1.4	<1	406	109	3.9	<2	<5.0	5.8
2010 Min	24	<5	<5	<0.4	<0.4	<0.5	<0.5	58.0	41.9	3.19	1.81	<0.2	<0.2	5.1	3.6	<1	<1	159	95	3.0	<2	<5.0	<5.0
2010 Max	30	<5	<5	<0.4	<0.4	3.67	0.660	83.6	82.9	8.02	3.79	<0.2	<0.2	7.2	4.5	2.19	<1	1520	118	4.9	<2	<5.0	7.7

Inf. Influent  
 Eff. Effluent  
 mgd million gallons per day

During 2010, cyanide concentrations in the effluent ranged from  $< 5.0$  to  $7.7 \mu\text{g/L}$ , well below the MAEC of  $181 \mu\text{g/L}$ . Cyanide had been a constituent of concern in the prior permit because it approached or even exceeded the prior MAEC of  $50 \mu\text{g/L}$  in some years. In 1986 it was observed that the effluent cyanide concentrations often exceeded the influent concentrations by an order of magnitude. This trend continued during subsequent years of sampling and was the subject of a special investigation conducted by the AWWU (CH2M Hill, 1987; CH2M Hill in association with Loren Leman, P.E., 1988). The conclusion of this investigation was that the measured increase in cyanide between the influent and effluent was the result of the treatment plant's incinerator. Cyanide formed in the incinerator during sludge incineration was returned to the plant during the stack scrubbing process. Subsequently, cyanide decreased and this was believed to be due to the change in the scrubber water source from recirculated primary effluent to well water.

The most restrictive criteria of growth and propagation of fish, shellfish, other aquatic life, and wildlife was used for the hydrocarbon limits presented in Table 21. The MAECs for TAqH and TAH as BETX were met again this year, with maximum levels in the effluent well below the AWQS. The parameter of TAqH was analyzed in effluent only during the receiving water quality sampling, and the TAqH concentration was  $23.68 \mu\text{g/L}$  as compared to the MAEC of  $2,715 \mu\text{g/L}$ . The maximum BETX (TAH) value of  $18.5 \mu\text{g/L}$  (measured by EPA method 624) was seen during the toxic pollutant and pesticide July 2010 sampling, and this value fell well below the MAEC of  $1,810 \mu\text{g/L}$ .

The MAEC for total ammonia was met again this year, with effluent values exhibiting a maximum of  $22.9 \text{ mg/L}$  as compared to the MAEC of  $217 \text{ mg/L}$  for the chronic limit and a MAEC of  $1,466 \text{ mg/L}$  for the acute limit. These MAECs are based on saltwater acute criteria of  $1.2 \text{ mg/L}$  and saltwater chronic criteria of  $8.1 \text{ mg/L}$ , which are a function of temperature, salinity, and pH as presented in the AWQS. For comparison in this report the criteria were based on a salinity of 20 psu, temperature of  $15^\circ\text{C}$ , and a pH of 8.0 units.

In addition to the MAECs that are based on the State and Federal water quality criteria, a number of other effluent limitations are specified in the NPDES permit. These daily, weekly, and monthly limitations include pH, TRC,  $\text{BOD}_5$ , TSS, and fecal coliform (Table 21). All of these parameters met all permit limitations for 2010.

For fecal coliform, the permit limitation of a monthly geometric mean (of at least five samples) that shall not exceed 850 FC MPN/100 mL was not exceeded this year, with the maximum monthly geometric mean of 18 FC MPN/100mL. Fecal coliform also met the monthly criteria "that not more than 10 % of the effluent samples shall exceed 2600 FC MPN/100 mL during any month" for all of 2010.

The permit limitations for monthly and weekly averages and daily maximum were met for  $\text{BOD}_5$  and TSS. Amendments to the Clean Water Act (40 CFR; Final Rule, 8/9/94), require at least 30 % average monthly removal for both of these parameters.  $\text{BOD}_5$  and TSS met this requirement on both an average monthly and annual basis with the lowest monthly removal of 43% for BOD and 77% for TSS. Removal of  $\text{BOD}_5$  averaged 48 % for the 2010 calendar year which is similar to that seen in 2009 although higher than was seen during the previous five years where average annual removals ranged from 33 to 39 %. The monthly removal for TSS ranged from 77 to 81 %

with an annual average of 79 %, about the same as reported for the last ten years and well above the average monthly criteria of 30 %.

Concentrations of other toxic pollutants and pesticides detected in the influent and final effluent were generally lower than or within the range of those detected in other POTWs from across the nation, even though the Asplund WPCF provides only primary treatment as compared to secondary treatment provided at the other plants (Table 22). Toxic pollutants and pesticides also generally fell within the historical range of values seen in past years; levels of toxic pollutants and pesticides detected in the Anchorage effluent this year and over the previous five years are shown in Table 23. These data indicated some variability over time, but a generally similar pattern overall. Levels were low and often below reporting limits. As in the past, the types and concentrations of measured organic compounds varied between the two sampling periods. Also, in some instances, large differences in pollutant concentrations occurred between the influent and effluent. Inconsistencies can be explained by looking at sampling methodology and plant operation in the case of point-source contaminants. If spikes of contaminants are occurring in the influent, these might be hit or missed during sampling. On the other hand, an effluent sample could still contain the contaminant because of mixing in the clarifiers. Differences in concentrations in influent and effluent samples could also be due to lower suspended solids in the effluent samples. This can be seen in Table 9 and Table 10, where greater variability usually occurs in the influent concentrations as compared to the effluent.

Historic discharge monitoring data (1986 - 2010) for other parameters of concern measured in the influent and effluent are presented in Table 25. Most parameters have remained fairly steady over time. Dissolved oxygen levels increased from 1986 with a peak in 1992, and then generally decreased over the next ten years. Levels of DO over the past six years including 2010 have been fairly consistent and generally in the range of 2 to 6 mg/L. Other constituents of potential concern such as TSS have remained fairly steady in both the influent and effluent over the past six years and within the range of that seen historically. The BOD<sub>5</sub> effluent average during 2010 (133 mg/L) is similar to that seen in recent years and well within the range seen historically. Average BOD<sub>5</sub> levels in the influent in 2010 were slightly higher than that seen over the prior five years yet still within the range seen historically.

The yearly average effluent fecal coliform bacteria concentration, reported at 18 MPN/100 mL for 2010, was similar to that seen in 2007 through 2009 and considerably lower than the prior three years. Elevated levels seen in 2004 were the highest yearly average seen on the program to date and were most likely the result of a program to optimize chlorine usage as described below. Lower fecal coliform values for 2006 through 2010 showing a downward trend are more in line with prior data, indicating that the disinfection efficacy at the WPCF has been improving.

As described in earlier reports, a project to improve the efficiency of the Asplund WPCF effluent disinfection system was implemented during 2001-2002. The chlorine injection process was improved by installation of rapid mixing equipment (the “Water Champ”, installed in November 2001) to mix chlorine gas directly with the effluent. Oxidation Reduction Potential (ORP) technology using a *Strantrol 890 Controller* was installed in December 2001 to control the chlorine dosage rate by adjusting it in response to both flow and oxidation reduction potential of the wastewater. Prior to this improvement, it was never possible to determine an exact correlation between TRC and coliform kill. Dosage control by the ORP has resulted in adequate coliform kills with far lower residuals and has substantially reduced the annual chlorine usage, but optimizing the disinfection process continues to be an on-going focus.

**Table 25. Historical Mean Monthly Discharge Monitoring Data (1986-Present) for Influent and Effluent Non-Metals.**

Year	Temperature (°C)		pH <sup>a</sup>		TRC (mg/L)		DO (mg/L)		BOD <sub>5</sub> (mg/L)		TSS (mg/L)		Fecal Coliform (MPN/100 mL)		Ammonia (mg/L)	
	Influent	Effluent	Influent	Effluent	Influent	Effluent	Influent	Effluent	Influent	Effluent	Influent	Effluent	Influent	Effluent	Influent	Effluent
1986-2004 Min	9	9	6.4	6.4	NT	0.07	NT	2.1	98	69	117	37	NT	5	NT	13.8
1986-2004 Max	17	18	8.2	8.5	NT	1.0	NT	8.6	296	174	307	86	NT	1213	NT	29.3
2005 Avg	12.7	13.2	NA	NA	NT	0.30	NT	4.6	205	148	235	54	NT	175	NT	20.0
2005 Min	10.7	11.1	7.4	7.2	NT	0.17	NT	3.1	194	131	216	45	NT	19	NT	18.5
2005 Max	15.6	16.1	7.9	7.5	NT	0.55	NT	6.2	284	174	273	58	NT	484	NT	22.0
2006 Avg	12.0	12.3	NA	NA	NT	0.34	NT	4.0	220	146	229	54	NT	44	NT	19.4
2006 Min	9.8	10.0	6.7	6.5	NT	0.18	NT	2.6	199	132	196	50	NT	4	NT	17.0
2006 Max	14.8	15.1	7.8	8.1	NT	0.56	NT	5.3	237	165	268	59	NT	229	NT	21.5
2007 Avg	11.8	11.9	NA	NA	NT	0.38	NT	3.7	229	148	231	57	NT	18	NT	19.3
2007 Min	9.4	9.9	6.6	6.5	NT	0.25	NT	1.5	180	124	195	48	NT	3	NT	17.2
2007 Max	14.6	15.4	8.0	7.9	NT	0.57	NT	6.1	279	174	284	66	NT	76	NT	22.9
2008 Avg	12.0	11.9	NA	NA	NT	0.35	NT	2.9	211	131	223	51	NT	20	NT	19.9
2008 Min	10.0	9.3	6.8	6.6	NT	0.20	NT	2.2	189	119	194	46	NT	6	NT	16.1
2008 Max	14.2	15.1	8.5	7.7	NT	0.49	NT	3.8	244	149	265	56	NT	63	NT	22.5
2009 Avg	11.7	12.2	NA	NA	NT	0.34	NT	2.8	235	125	239	56	NT	14	NT	21.6
2009 Min	8.9	9.4	6.6	6.5	NT	0.24	NT	1.3	172	83	187	48	NT	5	NT	18.2
2009 Max	14.7	15.4	7.9	7.4	NT	0.52	NT	4.3	274	145	269	63	NT	30	NT	25.8
2010 Avg	12.8	13.0	7.0/7.8	6.8/7.5	NT	0.29	NT	3.0	253	133	252	53	NT	18	NT	22.9
2010 Min	10.2	10.6	6.6	6.5	NT	0.22	NT	2.3	216	109	218	47	NT	7	NT	14.6
2010 Max	15.2	15.4	8.0	7.7	NT	0.43	NT	3.6	280	159	289	61	NT	49	NT	26.7

<sup>a</sup> Values represent monthly pH minimum and maximum

NA Not applicable

NT Not tested

The average TRC had dropped from 0.8 mg/L in 2001 to 0.4 mg/L in 2002, and TRC levels fell even lower in 2003 and 2004, with average TRC values of 0.26 mg/L for both of these years. The average TRC in 2010 concentration of 0.29 mg/L was within the range seen in 2005 through 2008. The average fecal coliform monthly average rose from 39 FC MPN/100 mL in 2001 to a high of 325 FC MPN/100 mL in 2004, and in 2010 has dropped back even further to 18 FC MPN/100 mL. As noted above, there were no exceedances of fecal coliform permit limits in 2010.

Quarterly WET testing was conducted on 24-hr flow composite effluent samples during all four quarters of calendar year 2010. Echinoderm fertilization tests were performed during the second, third, and fourth quarters of 2010. Use of this test during the last three quarters of 2010 was based on the screening test results from the first quarter, which had determined the sea urchin to be the most sensitive species tested. Although a toxic response was seen from the effluent during three of the four quarters of WET testing, all results were found to be acceptable and within the permit specified limit of TUc < 143 and required no additional testing.

***In summary, effluent chemistry monitoring indicated that with no exceptions, effluent concentrations of toxic pollutants and pesticides, metals and cyanide, and conventional parameters were much lower than their applicable permit limits and their MAECs. Fecal coliform also met all permit specified criterion in 2010. All toxic pollutants and pesticides concentrations including metals and cyanide were lower than or within the range of those detected at secondary treatment plants from across the nation.***

### **5.1.2 Sludge Monitoring**

The current permit requires sludge monitoring twice per year, once each during the dry and wet conditions in summer as part of the toxic pollutant/pretreatment monitoring. There are no Part 503 monitoring requirements included in the permit, but sludge monitoring is required because the Part 503 regulations are self-implementing as described in Section 2.1.5. Part 503 monitoring of sludge at Asplund WPCF has been included in this report (Table 13) and will also be reported separately to EPA as required by the regulations by 19 February 2010.

While limits for levels of toxic pollutants and pesticides in sludge are not part of the current permit, comparisons can be made for these data based on other treatment facilities' monitoring results and to the site specific allowable limits for metals that were determined for the facility. In all cases, sludge metals were found to be substantially lower than the site specific allowable limits (Table 13). Again, data indicate that average concentrations of toxic pollutants and pesticides in Anchorage's sludge are generally lower than "typical" concentrations seen at other treatment facilities (Table 26)(EPA 1985c). In 2010, arsenic ranged from < 2.23 to 9.21 µg/g with an average concentration of 4.87 µg/g compared to a typical concentration of 4.6 µg/g and a 95<sup>th</sup> percentile value of 20.77 µg/g. Beryllium concentrations were all < 0.33 µg/g with all samples being ND in 2010 (Table 13). The detection limit for beryllium is similar to the "typical" result from other municipal sludges of 0.313 µg/g and well within the 95<sup>th</sup> percentile value reported from other treatment works of 1.168 µg/g. Nickel ranged from a low of 7.52 µg/g to a high of 14.5 µg/g compared to a "typical" concentration of 44.7 µg/g and a 95<sup>th</sup> percentile value reported from other treatment works of 662.7 µg/g. Chromium was also found to be

**Table 26. Comparison Between Sludge Analysis Results for Anchorage and Typical and Worse Case Concentrations Used by EPA in Developing Median or Mean Environmental Profiles<sup>a</sup>. All concentrations are in µg/g dry weight.**

Pollutant	2010 Anchorage Values			Typical Concentration	95 <sup>th</sup> Percentile "Worse Case"
	July <sup>b</sup>	August <sup>b</sup>	2010 AVG <sup>c</sup>		
Aldrin/Dieldrin	ND	ND	---	0.07	0.81
Arsenic	5.53	4.73	4.87	4.6	20.77
Benzene	ND	0..00047	---	0.326	6.58
Benzo(a)anthracene	ND	ND	---	0.68	4.8
Benzo(a)pyrene	ND	ND	---	0.14	1.94
Beryllium	ND	ND	ND	0.313	1.168
Bis(2-ethylhexyl)phthalate	31 D	ND	---	94.28	459.25
Cadmium	1.26	1.72	1.52	8.15	88.13
Carbon Tetrachloride	ND	ND	---	0.048	8.006
Chlordane (α,γ)	ND	ND	---	3.2	12
Chloroform	ND	0.00096 J	---	0.049	1.177
Chromium	13.60	15.10	12.53	230.1	1499.7
Copper	207	273	---	409.6	1427
Cyanide	ND	ND	---	476.2	2686.6
DDT/DDE/DDD	180P,D / 0.047D / 0.027D	0.270 / ND / ND	---	0.28	0.93
3,3-Dichlorobenzidine	ND	ND	---	1.64	2.29
Methylene chloride	ND	0.0065 B	---	1.6	19
Endrin	ND	ND	---	0.14	0.17
Hexachlorobenzene	ND	ND	---	0.38	2.18
Hexachlorobutadiene	ND	ND	---	0.3	8
Lead	19.3	18.60	19.10	248.2	1070.8
gamma-BHC (Lindane)	ND	ND	----	0.11	0.22
Malathion	ND	ND	---	0.045	0.63
Mercury	NT	0.833	0.96	1.49	5.84
Nickel	12.0	14.5	10.94	44.7	662.7
PCBs	ND	ND	---	0.99	2.9
Pentachlorophenol	ND	ND	---	0.0865	30.434
Phenanthrene	ND	ND	---	3.71	20.69
Phenol	ND	ND	---	4.884	82.06
Selenium	2.95	ND	---	1.11	4.848
Tetrachloroethene	ND	0.0052	---	0.181	13.707
Trichloroethene	ND	ND	---	0.46	17.85
2,4,6-Trichlorophenol	ND	ND	---	2.3	4.6
Vinyl Chloride	ND	ND	---	0.43	311.942
Zinc	610	653	---	677.6	4580

<sup>a</sup> Source: EPA 1985c. *Summary of Environmental Profiles and Hazard Indices for Constituents of Municipal Sludge: Methods and Results*. Office of Water Regulations and Standards, Appendix F.

<sup>b</sup> Data from NPDES 2010 toxic pollutant and pesticide monitoring

<sup>c</sup> Average from 2010 Part 503 sludge monitoring results

--- Not monitored in-plant for Part 503 ND Not detected

B Method blank contamination. J Estimated value (below MDL or MRL)

NT Not tested



elevated during the June sampling event 2009, but concentrations had returned to normal in 2010 where chromium in the sludge ranged from 7.94 to 16.2 µg/g compared to the typical concentration seen at other POTWs of 230.1 µg/g. The average mercury concentration measured during the Part 503 sampling was 0.96 µg/g, well below the typical concentration of 1.49 µg/g. The other Part 503 metals tested, cadmium and lead, also fell well below typical concentrations.

Other metals that were monitored but not a requirement of the Part 503 regulations were copper, selenium, and zinc. Copper and zinc concentrations were below typical concentrations during both the July and August sampling. The selenium value reported for the July 2010 sampling event was 2.95 µg/g, compared to the typical concentration of 1.11 µg/g and a 95<sup>th</sup> percentile concentration of 4.848 µg/g (Table 26).

Table 27 provides an overview of historical sludge data for total metals. In general, year 2010 data indicated similar concentrations of arsenic, cadmium, lead, and mercury compared to historical data over the last few years. As discussed previously, both chromium and nickel were found to be elevated during June 2009, but had dropped down to concentrations that were similar to the historic range for the remainder of the 2009 and for all sampling in 2010.

## **5.2 WATER QUALITY MONITORING**

### **5.2.1 Plume Dispersion Sampling**

To test the hypothesis that the water quality at the ZID boundary was not degraded with respect to the water quality at the nearfield and control stations, the non-parametric Kruskal-Wallis Test (Zar, 1984) was employed which determined whether significant differences occurred within the sample group. If significant differences were observed, Dunn's test, a test that performs pairwise tests of significance (alpha = 0.05), was employed (Dunn, 1964). The results of these tests for the July survey period as a function of water quality parameters are presented in Table 28. Non-detect values were replaced with half the detection limit value for statistical testing.

Data from the receiving water survey showed statistically significant differences between the three outfall groupings (within-ZID, ZID boundary, and nearfield stations) and a grouping that included the control station for salinity at the surface, middle, and bottom. As can be seen in the Table 28, significant salinity differences between stations were between the control and nearfield station for all three depths and between the control and ZID boundary for the surface. These differences were the result of the control stations being slightly less saline; however, though significant differences were found, these differences were very small. In the past the control stations have often been found to be less saline as a result of increased river influence on the north side of Knik Arm as a result of freshwater inputs from the Matanuska and Knik Rivers.

Temperature was found to be significantly different between the control and within-ZID station for the surface only with the control site being slightly warmer. No statistically significant differences were seen for dissolved oxygen, pH, turbidity, color, fecal coliform, or TRC for any depth.

**Table 27. Historical Discharge Monitoring Data (1986 - Present) for Metals in Sludge.**  
Concentrations are in mg/kg dry weight. Results for years 1986-2004 represent the range of the results for monthly minimum (Min) and maximum (Max) as available. Results for 2005-2010 represent Part 503 sludge monitoring values.

Year	Arsenic	Beryllium*	Cadmium	Chromium	Lead	Mercury	Nickel
<b>1986-2004 Min</b>	1.7	<0.02	0.7	3.38	19	<0.1	7
<b>1986-2004 Max</b>	151	0.23	10.0	55	468	7.3	53
2005 Avg	2.4	0.21	2.0	14	25	1.5	16
2005 Min	1.8	0.13	1.69	12	10	<0.02	11
2005 Max	2.7	0.37	3.0	19	33	3.1	20
2006 Avg	3.8	0.06	2	17	26	0.86	15
2006 Min	1.9	ND	1.23	13	18.2	0.390	11.9
2006 Max	5.94	0.15	3	25	39.3	1.70	19
2007 Avg	5.00	N/A	1.56	14.5	18.9	0.818	11.0
2007 Min	ND(<3.2)	ND(<0.307)	0.986	12.8	9.8	0.576	9.43
2007 Max	7.85	ND(<3.38)	2.37	15.8	25.1	1.04	13.4
2008 Avg	4.73	N/A	1.35	13.7	21.4	0.812	11.5
2008 Min	ND	0.08 J	1.02	9.77	15.4	0.357	8.34
2008 Max	5.40	ND< (0.42)	1.80	24.0	31.2	1.93	13.6
2009 Avg	3.85	0.49	1.11	26.17	17.70	0.541	59.34
2009 Min	ND	ND	0.66	7.90	10.60	0.231	7.23
2009 Max	6.34	1.75	1.77	122.00	25.80	0.885	400.00
2010 Avg	4.87	ND	1.52	12.53	19.10	0.96	10.94
2010Min	2.23	ND (0.314)	1.01	7.94	12.60	0.403	7.52
2010 Max	9.21	ND(0.329)	2.53	16.20	24.80	2.140	14.5

N/A – Not applicable ND – Not Detected

All pH values fell well within the AWQS of 6.5 - 8.5 and values did not vary more than 0.2 pH units that would be attributed to the outfall, as required by the AWQS. For color, concentrations were found to be all either at or below the detection limit of 5 color units. In 2007, color was found to be elevated at all locations compared to most prior years, and was believed to be due to the naturally high suspended sediment levels in the samples from Knik Arm that were not completely removed from the samples during the laboratory analysis. During 2008 through 2010, an additional preparation step was added where color samples were allowed to settle and decanted in the field prior to submittal to the laboratory. Upon reaching the laboratory the samples were processed by either filtration or centrifuging which resulted in much more representative sample analysis results. It is anticipated that this procedure will be continued in future years.

**Table 28. 2010 Station Group Differences at the 5 Percent Significance Level ( $p \leq 0.05$ ) as determined by the Nonparametric Kruskal-Wallis (ANOVA) and Dunn's (ad hoc multiple comparison) Statistical Analysis Tests.**

Water Quality Parameter	Water Column Depth		
	Surface	Middle	Bottom
<u>Conventional Analyses*</u>			
Temperature	<u>1 2 3</u> vs. <u>2 3 4</u>	NS	NS
Salinity	<u>4 1</u> vs. <u>1 2 3</u>	<u>4 1 2</u> vs. <u>1 2 3</u>	<u>4 1 2</u> vs. <u>1 2 3</u>
Dissolved Oxygen	NS	NS	NS
pH	NS	NS	NS
Turbidity	NS	NS	NS
Color Units <sup>HV</sup>	NS	----	----
Fecal Coliform <sup>HV</sup>	NS	----	----
Total Residual Chlorine <sup>HV</sup>	NS	----	----
<u>Metal and Hydrocarbon Analyses**</u>			
Arsenic	NS <sup>D</sup> , <b>SIG</b> <sup>TR</sup>	----	----
Cadmium	NS <sup>D</sup> , <b>SIG</b> <sup>TR</sup>	----	----
Chromium	NS <sup>D</sup> , NS <sup>TR</sup>	----	----
Copper	NS <sup>D</sup> , <b>SIG</b> <sup>TR</sup>	----	----
Mercury	NS <sup>D</sup> , NS <sup>TR</sup>	----	----
Nickel	NS <sup>D</sup> , NS <sup>TR</sup>	----	----
Lead	NS <sup>D</sup> , NS <sup>TR</sup>	----	----
Silver <sup>HV</sup>	NS <sup>D</sup> , NS <sup>TR</sup>	----	----
Zinc	NS <sup>D</sup> , NS <sup>TR</sup>	----	----
Cyanide <sup>HV</sup>	NS <sup>TR</sup>	----	----
Total Suspended Solids	NS <sup>TR</sup>	----	----
Total Aromatic Hydrocarbons (BETX)	NS <sup>TR</sup>	----	----
Total Aqueous Hydrocarbons (TAqH)	<b>SIG</b> <sup>TR</sup>	----	----
Total Petroleum Aromatic Hydrocarbons (TPAH)	<b>SIG</b> <sup>TR</sup>	----	----

\* Tests were performed on four Station Groups (Group 1: Within-ZID Sites; Group 2: ZID Boundary Sites; Group 3: Nearfield Sites; and Group 4: Control Sites). Bolded Groups signify **SIG** result. Underlined Groups are not significantly different from each other and **red** denotes significantly different Groups with a Bonferroni corrected significance level of 0.00833 (Dunn's Test result,  $p > 0.05$  for six tests).

\*\* Tests performed on stations located between the outfall (Drogue F1 sites) and the control (Drogue C1 sites).

---- Not Applicable (surface samples only)

NS Not Significant K-W Test Result ( $p > 0.05$ )

**SIG** Significant K-W Test Result ( $p \leq 0.05$ )

<sup>D</sup> Dissolved

<sup>TR</sup> Total Recoverable

<sup>HV</sup> Half the detection limit used in place of data with "<" and NDs.

In addition to the standard water quality sampling, concentrations of total aromatic hydrocarbons (TAH) as BETX and TAqH were measured at the surface at six stations (three at the control site and three at the flood tide outfall site, along the first drogue track). No statistically significant differences in TAH concentrations were detected between the control and outfall stations. There were no BTEX concentrations above MRLs seen at the Control Stations. Two outfall stations, F1-1 within the ZID and F1-3 nearfield, had BTEX values slightly above the MRL, however, they were well below, the AWQS of 10 µg/L. TAqH concentrations were calculated for all six stations using the MDL as the value for all non-detects yielding a maximum of 7.15 µg/L, significantly less than the AWQS of 15 µg/L at all locations. TAqH and TPAH were found to be significantly different between the control and outfall sites as a result of the slightly elevated levels at Station F1-1. In addition, comparison of TAqH and TPAH concentrations indicate that levels were elevated only at the outfall station during low slack water (F1-1), as has sometimes been seen in the past. All TPAH levels were relatively low.

Total suspended solids, cyanide, and total recoverable and dissolved metals samples collected at the outfall and control sites were also subject to statistical testing. No statistically significant differences were noted for TSS, cyanide, or any of the dissolved metals that were tested.

Statistically significant differences were seen for the total recoverable metals arsenic, cadmium, and copper due to the higher concentrations at the control site. Although not statistically significant, levels of most dissolved metals were found to be slightly higher at Station F1-1 that is located within the ZID. This probably is an outfall related effect, although all concentrations (except copper as noted previously) were still well below AWQS, even within the ZID.

A comparison of the water quality data listed in Table 15 with the marine receiving water quality criteria for the State of Alaska (Table 21 and Table 29) indicates that none of the parameters listed in Table 15 exceeded AWQS outside the ZID. All of the TRC concentrations were below the PQL of 0.01 mg/L for all locations both within and outside of the ZID. It should be noted that the lowest achievable method detection limit in 2010 for the analysis of TRC in seawater (0.01 mg/L) is higher than the State-specified limit of 0.0075 mg/L for chronic 4-day average and below the 0.013 mg/L limit for acute 1-hr average for marine water use. As previously noted, all TRC methods are subject to positive interferences in estuarine or marine waters. However, based on the maximum daily effluent TRC concentration (0.88 mg/L) measured during 2010 and a dilution credit of 180:1 in the NPDES permit, the highest potential TRC concentration at the ZID boundary would be < 0.005 mg/L which meet AWQS at all receiving water locations outside of the ZID. Also, although the analysis was only able to achieve a PQL of 0.01 mg/L, ADEC considers a PQL of 0.1 mg/L to be the reasonable and achievable limit for regulatory purposes.

The State's receiving water quality standard for the "growth and propagation of fish, shellfish, aquatic life, and wildlife including seabirds, waterfowl, and furbearers" is 15 µg/L for TAqH and 10 µg/L for TAH. As seen in Table 17, these standards were not exceeded during the receiving water sampling. For all control stations, TAH as BETX was below the method reporting limit and well below the AWQS. Two outfall stations (F1-1 and F1-3) had measurable toluene values which pushed the total BTEX value to a maximum of 6.5 µg/L; still well below the AWQS of 10 µg/L. In addition, for "contact recreation", the AWQS for hydrocarbons is as follows: "Shall not cause a film, sheen, or discoloration on the surface or floor of the water body or adjoining shorelines. Surface waters shall be virtually free from floating oils." No film, sheen, or discoloration was observed during the receiving water sampling program during 2010.

**Table 29. State of Alaska Water Quality Standards for Receiving Water.**

Parameter	Most Restrictive Marine Water Quality Standards								
Fecal Coliform	Based on a 5-tube decimal dilution test the fecal coliform median MPN shall not exceed 14 FC/100 mL (harvesting for consumption of raw shellfish); a geometric mean of 20 FC/100 mL (for aquaculture of products not normally cooked and seafood processing); and not more than ten percent (10%) of the samples shall exceed 40 FC/100 mL (aquaculture of products not normally cooked and seafood processing).								
Dissolved Oxygen	Dissolved oxygen concentrations in estuaries and tidal tributaries shall not be less than 5.0 mg/L except where natural conditions cause this value to be depressed.								
pH	pH shall not be less than 6.5 or greater than 8.5, and shall not vary more than 0.2 pH unit from natural condition.								
Turbidity	Turbidity may not exceed the natural condition.								
Temperature	Temperature shall not cause the weekly average temperature to increase more than 1° C. The maximum rate of change shall not exceed 0.5° C per hour. Normal daily temperature cycles shall not be altered in amplitude or frequency.								
Salinity	<p>Maximum allowable variation above natural salinity:</p> <table> <tr> <th>Natural Salinity (‰)</th><th>Man-induced Salinity (‰)</th></tr> <tr> <td>0 to 3.5</td><td>1</td></tr> <tr> <td>3.5 to 13.5</td><td>2</td></tr> <tr> <td>13.5 to 35.0</td><td>4</td></tr> </table>	Natural Salinity (‰)	Man-induced Salinity (‰)	0 to 3.5	1	3.5 to 13.5	2	13.5 to 35.0	4
Natural Salinity (‰)	Man-induced Salinity (‰)								
0 to 3.5	1								
3.5 to 13.5	2								
13.5 to 35.0	4								
Sediment	No measurable increase in concentrations above natural conditions.								
Color	Color shall not exceed 15 color units or the natural conditions, whichever is greater.								
Petroleum Hydrocarbons, Oils and Grease	Total aqueous hydrocarbons (TAQH) in the water column shall not exceed 15 µg/L. Total aromatic hydrocarbons (TAH) in the water column shall not exceed 10 µg/L. Shall not cause a film, sheen, or discoloration on the surface or floor of the water body or adjoining shorelines. Surface waters shall be virtually free from floating oils.								
Total Residual Chlorine	Shall not exceed 13 µg/L (one hour average) acute, and 7.5 µg/L (four day average) chronic; for marine aquatic life.								
Toxic and Other Deleterious Substances	See Table 21.								

All the dissolved metals tested in receiving water (Table 16) as part of this program met the AWQS as shown in Table 21 at all locations on the ZID boundary and outside of the ZID including the control stations. These included: arsenic, cadmium, copper, chromium, lead, mercury, nickel, silver, and zinc. One copper sample taken at Station F1-1 was found to be higher (6.95 µg/L) than the AWQS of 3.1 µg/L, but this sample was taken from within the NPDES permit specified ZID and therefore is not a violation of AWQS. Testing of antimony, beryllium, selenium, and thallium in receiving water is not required by the permit and was not performed in 2010. Previous years of monitoring showed exceedances of water quality criteria for total recoverable metals due to the specified test methods in conjunction with high amounts of suspended particulates in Cook Inlet. Since the adoption of the more-appropriate SSWQC for dissolved metals in May 1999 and the adoption of dissolved metals in the AWQS, the receiving waters of Cook Inlet near the Asplund WPCF discharge have been in compliance with the AWQS.

All cyanide samples collected during the receiving water sampling were below the State-specified criteria of 1 µg/L for marine aquatic life. The highest cyanide concentration that was seen in 2010 was at Station C1-1 with a value of 0.19 µg/L. The cyanide concentrations in the effluent samples, that were measured as part of the receiving water sampling, were reported at a concentration of 0.20 µg/L, well below the MAEC of 181 µg/L.

*In summation, statistical analyses of the 2010 receiving water quality data indicated that water quality outside the ZID was not degraded with respect to control stations for any parameter as a result of the outfall. Differences that were noted in some parameters such as salinity and temperature were attributed to riverine influences and were not caused by the Asplund WPCF discharge. All AWQS were met in 2010 for the Asplund WPCF receiving water quality program. Although dissolved metal parameters appeared to be slightly elevated at the within ZID station as compared to control stations, no statistically significant increases at the outfall stations were seen, and all parameters were well below AWQS at all locations at or outside of the mixing zone. Statistically significant differences between the outfall and control site were seen for the total metals arsenic, cadmium, and copper due to elevated levels at the control site due to higher TSS concentrations. No statistically significant differences were seen for cyanide or TAH between the outfall and control locations and all cyanide and hydrocarbon concentrations were below AWQS at all locations in the receiving water including those within the mixing zone.*

### **5.2.2 Fecal Coliform Bacteria**

In the past, the ADEC has indicated that one of their primary concerns is bacterial contamination of the shoreline by the Asplund WPCF discharge, indicated by fecal coliform bacteria concentrations. Because the Knik Arm's water uses have not been classified, regulations provide that the most restrictive standard must apply. State marine water quality standards for contact recreation require that the geometric mean fecal coliform concentration taken within a 30-day period not exceed 100 FC MPN/100 mL and that not more than one sample, or more than 10 % of the samples if there are more than 10, exceed 200 FC MPN/100 mL. Criteria for secondary recreation and for industrial water supply require that the mean fecal coliform concentration not exceed 200 FC MPN/100 mL and that not more than 10 % of the samples exceed 400 FC MPN/100 mL. State marine water quality criteria for the harvesting for consumption of raw mollusks and other raw aquatic life require that, based on a 5-tube decimal dilution test, the

median shall not exceed 14 FC MPN/100 mL, and that not more than 10 % of the samples shall exceed 43 FC MPN/100 mL. For seafood processing water supply for products not normally cooked, criteria are that the geometric mean may not exceed 20 FC MPN/100 mL, and not more than 10 % of the samples shall exceed 40 FC MPN/100 mL. For seafood processing water supply for products normally cooked, criteria are that the geometric mean may not exceed 200 FC MPN/100 mL, and not more than 10 % of the samples shall exceed 400 FC MPN/100 mL. For aquaculture water supply, criteria are that the geometric mean may not exceed 20 FC MPN/100 mL and not more than 10 % of the samples may exceed 40 FC MPN/100 mL.

Since the harvesting of shellfish and other raw aquatic life is not performed in these waters, and there is no aquaculture or seafood processing, it seems that the criteria for secondary recreation is most applicable; however, these criteria are not the most restrictive. Therefore, the most restrictive criteria used were that the median shall not exceed 14 FC MPN/100 mL (consumption of raw shellfish and other aquatic life), the geometric mean shall not exceed 20 FC MPN/100 mL (seafood processing and aquaculture for raw consumption), and not more than 10 % shall exceed 40 FC MPN/100 mL (seafood processing and aquaculture for raw consumption; Table 29).

Statistical tests indicated that fecal coliform concentrations were not significantly different between the within-ZID, ZID boundary, and the nearfield outfall station groups as compared to the control stations (refer to Table 28). Fecal coliform concentrations were very low again this year, and values ranged from <2 to 50 FC MPN/100 mL at the outfall stations (including the ZID stations) and a range of <2 to 11 FC MPN/100 mL at the control stations. The median at the control stations was 3 FC MPN/100 mL, and the median at the all outfall stations was 4 including stations both within and outside the ZID for both ebb and flood tides, well within the 14 FC MPN/100 mL criterion. The control site had a geometric mean of 3.8 FC MPN/100 mL, while that at the all the outfall stations (inside and outside the ZID) was 3.9 FC MPN/100 mL, both well below the criterion of 20 FC MPN/100 mL. One sample within the ZID (3 %) at the outfall stations including within and outside the ZID exceeded 40 FC MPN/100 mL, compared to the criteria of not more than 10 % of the measurements may exceed 40 FC MPN/100 mL. No samples collected at the control stations exceeded this criterion. The highest fecal coliform concentration (50 FC MPN/100 mL) was seen this year at a within-ZID outfall station (F1-1) at low slack tide.

The range of fecal coliform concentrations for all intertidal samples collected during 2010 was higher than that seen in 2009 and ranged from a low at <2 to 240 FC MPN/100 mL, with a median of 10 FC MPN/100 mL and a geometric mean of 10.1 FC MPN/100 mL. These values met the most restrictive water quality criterion of a median of 14 FC MPN/100 mL and a geometric mean of 20 FC MPN/100 mL. The highest coliform concentrations were seen at Station IT-4, 250 m east of the outfall. The criterion of not more than 10 % of the samples exceeding 40 FC 100/mL was not met, as 2 of 18 (11 %) intertidal measurements exceeded this value. However, due to the very low values encountered in 2010 in both the effluent and in the receiving water program, it does not appear that the outfall would necessarily be the source of the bacteria concentrations. In the past, elevated fecal concentrations have sometimes been seen in the intertidal area that were attributed to heavy waterfowl use of the area, and not believed to be the result of the effluent discharge. The area is also heavily used in the summer by hikers that access the beach at Pt. Woronzof and often use the area for walking their dogs.

Elevated fecal coliform bacteria concentrations were seen in all three area creeks that were sampled prior to their entering Knik Arm which is similar to historical data, and creek

concentrations in general were higher than most receiving water and intertidal samples that were collected. The two replicate fecal coliform concentrations measured in Fish Creek were 540 and >2400 FC MPN/100 mL. Replicate concentrations measured in Ship Creek were much lower at 49 and 79 FC MPN/100 mL, while those at Chester Creek were <0.5 and 49 FC MPN/100 mL. These fecal coliform concentrations from area streams that discharge into Knik Arm were much greater than that seen in the receiving water at the intertidal, outfall, or control locations.

Fecal coliform concentrations in the effluent samples collected in conjunction with the receiving water, intertidal sampling, and stream sampling were also low at 8 and 11 FC MPN/100 mL for the two replicates. These values were within the range of average monthly effluent values reported for the Asplund WPCF, which in 2010 ranged from 7 to 49 FC MPN/100 mL (Table 8).

*In summary, fecal coliform concentrations in 2010 were found be very low in the receiving water area. No statistically significant differences were seen between station groupings for the ZID, ZID-boundary, or nearfield stations as compared to the control location. Fecal coliform samples collected during the receiving water sampling program met all AWQS criteria, including all outfall stations both within and outside the ZID. Fecal coliform samples collected from intertidal areas were found to be slightly elevated at one location and exceeded the AWQS of not more than 10% may exceed 40 FC MPN/100 mL at 11%. Heavy waterfowl and recreational use of the intertidal area has been noted in the past, and based on the relatively low effluent concentrations, the outfall does not appear to be the source of these fecal coliform levels. Area creeks in 2010 were again found to be elevated and within the historical range for fecal coliform stream concentrations seen in prior years. It is clear from both this data and historic data that area streams are an important source of fecal coliform loading to the receiving waters of Knik Arm and that waterfowl use of the intertidal areas may cause elevated fecal coliform levels that are higher than those being discharged by the Asplund WPCF into Cook Inlet.*



## 6.0 CONCLUSIONS

The following conclusions were based on results from the 2010 monitoring effort as compared to the current NPDES permit and State of Alaska water quality standards:

- The influent, effluent, and sludge monitoring showed, that with no exceptions, the Asplund WPCF met the NPDES permit requirements and complied with all applicable AWQS. AWWU's self-monitoring of TRC, pH, fecal coliform bacteria, BOD<sub>5</sub>, and TSS showed compliance with all permit effluent limitations throughout 2010.
- AWWU's self-monitoring of TRC and pH showed that the permit limit for daily maximum TRC levels in the effluent was never exceeded and pH was within permit limits throughout 2010.
- The permit limit for the monthly maximum geometric mean of 850 fecal coliform colonies per 100 mL by most probable number (FC MPN/100 mL) technique was not exceeded in 2010. The fecal coliform monthly criteria "that not more than 10 % of the effluent samples shall exceed 2600 FC MPN/100 mL" was also not exceeded in any month during 2010.
- AWWU's self-monitoring of TSS and BOD<sub>5</sub> showed compliance with both regulatory and permit effluent limitations. TSS and BOD<sub>5</sub> were well within the daily, weekly, and monthly criteria for the entire year. Average monthly removals for BOD<sub>5</sub> and TSS of greater than 30 % are required by the amendment to the CWA (40 CFR Part 125; Final Rule, 8/9/94). The removal rate for both TSS and BOD<sub>5</sub> met the 30 % minimum removal requirement for all months during 2010. Annual removals were 79 % for TSS and 48 % for BOD<sub>5</sub> which indicate an exceptional level of primary treatment.
- Total aromatic hydrocarbons (TAH), total aqueous hydrocarbons (TAqH), and total ammonia concentrations in the effluent were all found to be below their maximum allowable effluent concentrations (MAECs) throughout 2010.
- Concentrations of metals and cyanide in the effluent never exceeded their MAECs at any time during any of the 2010 monitoring events.
- Concentrations of toxic pollutants and pesticides, including metals and cyanide, in the influent and effluent were all within the established range or lower than values from a national study of secondary treatment plants (EPA 1982a).
- Toxic pollutant sludge concentrations were found to be very low compared to the limits established under 40 CFR Part 503 and most were either not-detected or within the established range or lower than values from a national study of secondary treatment plants. Most metals fell at or below the typical concentrations and all metals were below 95<sup>th</sup> percentile worst case values (EPA 1985c).
- Whole effluent toxicity testing conducted quarterly met all permit limitations for toxicity for all tested species and all monitoring events in 2010.
- Little variation among stations was observed for most hydrographic parameters indicating that the receiving water environment is uniform and well mixed near the outfall.

- To test the hypothesis that the water quality at the ZID boundary was not degraded with respect to the water quality at the nearfield and control stations, statistical comparisons were employed. Conventional parameters such as salinity and temperature did show significant differences between sites, however, these were not ascribed to the outfall but were due to river influences at the control stations. No statistically significant differences were seen for dissolved oxygen, pH, turbidity, TRC, TSS, color, or fecal coliform.
- Fecal coliform concentrations in receiving water and intertidal samples were found to be low at most locations. AWQS criteria of a median of not more than 14 FC MPN/100 mL, a geometric mean of not more than 20 FC MPN/100 mL, and of not more than 10 % of the samples exceeding 40 FC MPN/100 mL were met at all receiving water locations. Intertidal stations exceeded the criteria of not more than 10 % of samples exceeding 40 FC MPN/100 mL as a result of two elevated samples of unknown source at one location.
- Supplemental receiving water quality samples obtained as part of the plume dispersion monitoring indicated that dissolved metals were all below the AWQS at all locations on the ZID boundary and outside of the ZID including the control stations. No statistically significant differences between the outfall and control station groupings were seen for any dissolved metal. Total metals were elevated compared to the dissolved as a result of the naturally high suspended sediment load. Statistically significant differences were seen for arsenic, cadmium, and copper due to higher ambient TSS at the control stations.
- All cyanide concentrations in the receiving waters were found to be below the receiving water quality criterion limit of 1.0 µg/L and no statistically significant differences were detected between concentrations at the control and outfall stations.
- Supplemental receiving water samples also demonstrated that TAH and TAqH met the AWQS at all locations. No statistically significant differences were detected between the control and outfall stations for TAH. TAqH did exhibit significant differences due to higher levels seen at the within mixing zone station, but levels were still within AWQS.
- Turbidity and color met the AWQS at all stations. TRC was at or below the detection limit of 10 µg/L at all locations including those located within the ZID, as compared to the AWQS of 7.5 µg/L for chronic, 13.0 µg/L for acute marine water use, and 100 µg/L as ADEC's practical quantitation limit for regulatory purposes. Based on the highest maximum daily effluent TRC concentration of 880 µg/L seen during 2010 and a 180:1 effluent dilution credit, it is expected that TRC in the receiving water would be <5 µg/L before reaching the ZID boundary and would always meet all AWQS for TRC.

## CONCLUSIONS

Results from the past year of the monitoring program confirm previous studies, data in the NPDES permit and 301(h) variance renewal application, and the decision by the EPA to reissue the NPDES permit with 301(h) variance. The Asplund WPCF is operating within regulatory requirements with no exceptions seen in 2010 and is showing no measurable impacts to the marine environment.

## 7.0 REFERENCES

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